About the Book

Fundamentals of Botany" serves as an indispensable guide that explores the intricate tapestry of plant life with depth and clarity. Designed for both novices and seasoned enthusiasts, the book traverses the realms of plant biology, covering anatomy, physiology, taxonomy, and ecology. Through a holistic approach, it elucidates the fascinating structures within plants at the cellular level while seamlessly connecting these insights to broader ecological principles. The author skillfully weaves theoretical knowledge with practical applications, making the subject accessible and engaging. This comprehensive volume is a valuable resource for students, researchers, and anyone captivated by the wonders of the plant kingdom. Richly illustrated and thoughtfully organized, the book not only imparts a profound understanding of botanical concepts but also underscores the pivotal role plants play in sustaining life on Earth. "Fundamentals of Botany" is more than a textbook; it's a captivating journey into the intricate and diverse world of plants, fostering a deep appreciation for their significance in ecosystems and their impact on our daily lives. Whether you're delving into botany for academic pursuits or personal curiosity, this book is an illuminating companion on your botanical exploration.

Dnyaneshwar Raut, D. Thangamani, Indrajitsingh P. Girase, Pawan Kumar Goutam , Nikhil Agnihotri

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Editors

Mr. Dnyaneshwar Raut

PhD Research Scholar, Department of Plant Physiology University: Mahatma Phule Krishi Vidyapeeth Rahuri (MS). India

Dr. D.Thangamani

Scientist

ICFRE- Institute of Forest Genetics and Tree Breeding, Coimbatore

Mr. Indrajitsingh Pravinsingh Girase

Research Scholar (Seed Science and Technology) Department of Genetics and Plant Breeding Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj Uttar Pradesh, 211007

Dr. Pawan Kumar Goutam

(Teaching Associate) Department of Crop Physiology Chandra Shekhar Azad University of Agriculture and Technology Kanpur UP.

Dr. Nikhil Agnihotri

Assistant Professor and H.O.D. Faculty of Science S.K.J.D. Degree College, Kanpur Dehat, U.P.



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N D Global Publication House 31, Near Lakshmi Sagar Police Chowki Shahganj Haringtonganj Ayodhya Uttar Pradesh, Pin -224284, India.



Head Office:- Murali Kunj Colony, Near Chandra Greens, Society, Transport Nagar, Mathura, Uttar Pradesh, Pin-281004, India.

Mobile No.:-9026375938

Email: <u>bsglobalpublicationhouse@gmail.com</u> Web: <u>https://bsglobalpublicationhouse.com</u>



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PREFACE

Welcome to "**Fundamentals of Botany**," a comprehensive exploration of the fascinating world of plants and the fundamental principles that govern their existence. Plants, with their diverse forms and functions, play a vital role in sustaining life on Earth. This book is crafted to provide a solid foundation for understanding the intricacies of botany, from the cellular level to the ecological interactions that shape plant communities.

In the pages that follow, we embark on a journey to unravel the mysteries of plant life. Whether you are a student taking your first steps into the world of botany or a curious enthusiast seeking a deeper understanding of plant biology, this book is designed to be your guide. The text is structured to take you through the essential concepts, starting with the building blocks of plant life— cells, tissues, and organs—and progressing to the broader topics of plant physiology, taxonomy, and ecology. One of the key strengths of **"Fundamentals of Botany"** lies in its accessibility. Complex scientific concepts are presented clearly and engagingly, with an emphasis on real-world examples and applications. The goal is not only to impart knowledge but also to foster a genuine appreciation for the marvels of plant diversity and adaptation. Throughout the book, we encourage you to connect theoretical knowledge with practical observations, promoting a holistic understanding of the plant kingdom.

This book is also a tribute to the centuries of scientific inquiry that have shaped our understanding of plants. As we navigate through the history of botany, from the pioneering work of early botanists to the latest breakthroughs in molecular biology, we gain a sense of the ongoing dialogue between human curiosity and the mysteries of the botanical world.

Whether you are studying botany for academic purposes or simply cultivating a deeper appreciation for the green tapestry that surrounds us, "Fundamentals of Botany" is a resource that aims to inspire curiosity and ignite a passion for plant science. As we delve into the following chapters, let us embark on a journey of discovery, where the seemingly simple world of plants unfolds into a complex and awe-inspiring tapestry of life.

Authors 🔏

Dedicated to My Beloved







ABOUT THE EDITORS



Mr. Dnyaneshwar A. Raut is currently a Ph.D. Scholar specializing degree in Plant Physiology at Mahatma Phule Krishi Vidyapeeth, Rahuri (MS). His academic journey began with a Bachelor's degree in Agriculture from MPKV, Rahuri University. Subsequently, he achieved his Master's Degree in the Division of Plant Physiology from Dr. P.D.K.V., Akola (M.S.). He

demonstrated his academic prowess by securing rank-II in MCAER-CET, paving the way for his Ph.D. pursuit. His Ph.D. research is underway at ICAR-National Institute of Abiotic Stress Management, Baramati, focusing on elucidating the role of intrinsic ascorbic acid in enhancing drought tolerance in chickpeas.



Dr. D.Thangamani is working as a Scientist in ICFRE- Institute of Forest Genetics and Tree Breeding, Coimbatore, has 20 years research experience in the field of characterization of forest genetic resources its conservation and tree improvement. Currently working on characterization and conservation of Teak, Pongamia, Dalbergia, Madhuca and other important medicinal plants. She has

been involved in exploration and characterization of germplasms using various molecular methods and biochemical methods for finding genetic variation. She has developed natural dyes from forest plantation wastes. She has released products like face mask with bacterial filtration capacity and natural dyed garments to this society. She has been awarded with Best Women Scientist Award by Nature Foundation, Coimbatore and APJ Abdul Kalam award for her. Scientific Excellance during the year 2020. Her journey continues in characterization and conservation aspect to save nature.



Mr. Indrajitsingh Pravinsingh Girase is a Ph.D Scholar in Department of Genetics and Plant Breeding, Specialization in Seed Science and Technology at Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj Uttar Pradesh, and related research work done at ICAR-Directorate of Onion & Garlic Research (DOGR), Pune. With an excellent academic record, he has completed his

under-graduation from Mahatma Phule Krishi Vidyapeeth, Rahuri also completed his Master's degree in Seed Science and Technology from SHUATS, Prayagraj. He has published several research and review papers in national and international journals along with popular articles.



Dr. Pawan Kumar Goutam was born on 20 July 1990 at Shrawasti District UP. He is a Teaching Associate in the Department of Crop Physiology at Chandra Shekhar Azad University of Agriculture and Technology Kanpur UP, He did B.Sc. (Biology) from M.L.K (P.G.) College Balrampur affiliated with Dr. R. M. L. University Faizabad UP, M.Sc. (Ag.)

Crop Physiology from Acharya Narendra Deva University of Agriculture and Technology Faizabad UP and Ph.D. (Plant Physiology) from Chandra Shekhar Azad University of Agriculture and Technology Kanpur UP. Also qualified ASRB-ICAR NET (I) 2017 and 2018. He has contributed in many publications including 11 Research papers in National and International Journals, Books, Book Chapters, Review papers, Articles in hindi and English.



Dr. Nikhil Agnihotri is working as an Assistant Professor in Botany and H.O.D., Faculty of Science in S.K.J.D. Degree College, Kanpur Dehat, U.P. He was awarded with Ph.D. on the topic "Soil-Plant Relationship as Influenced by Azolla as Organic Compost" by Chhatrapati Shahu Ji Maharaj University, Kanpur in the year 2011. His research includes Biofertilizer, Ecology,

Ethnobotany, Taxonomy etc. He has 12 years of teaching experience in various higher education institutions at Graduate and Post Graduate level. Dr. Agnihotri has been honoured with various prestigious Awards like Atal Samrasata Award (2019) by Vice President of Nepal, Global Golden Award 2017, Scientist of the Year Award (2018, 2019, 2020), Shikshak Ratna Award (2022), Dr. Ambedkar Rashtriya Samrasata Award (2023) etc. About 60 research papers of Dr. Nikhil Agnihotri have been published in various National and International Journals. More than 30 Book chapters and more than 20 popular scientific articles have also been published by Dr. Agnihotri. He has also got 7 patents registered to his name till now.

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Plant Evolution Priya Patel

Department of Genetics and Plant Breeding, N. M. College of Agriculture, Navsari Agricultural University, Navsari-396 450. Corresponding Author email:- <u>3031priyapatel@gmail.com</u>

Abstract

As biologists, we are all well aware of the millions of years it took for prokaryotic species to evolve into the eukaryotic ones we see today. But what brought about this widespread alteration that we can detect as morphological change? The answer lies in the process of evolution. From the earliest algal bowls to multicellular marine and freshwater green algae, terrestrial mosses, lycopods, and ferns, to modern complex plant seeds and angiosperms, development in plants leads to complexity. Red and green algae in maritime habitats are a good example of how many of the earliest groups are still thriving. However, more recently derived groups have replaced biologically prominent ones.

Keywords: Evolution, selection, mutation, molecular evolution. **Introduction**

The stonewort, an aquatic alga, is likely a representative of the first plants to have existed on Earth. Stoneworts lack roots and contain rhizoids which resemble hair in place of rigid stems and this makes them different from most modern plants. In addition, they possess separate female and male reproductive systems, which is a hallmark of plants. These similarities suggest that stoneworts may have served as the origin of the first plants.

According to theory, plants descended from an aquatic green alga. Vascular tissues, leaves, stalks, seeds, and flowers were

among the essential terrestrial adaptations that were developed as time and life proceeded. These significant modifications improved plants' suitability for life on dry land.

Moving to land

By the time the earliest plants appeared, animals already dominated the ocean. Additionally, vegetation were restricted to the higher layers of ocean water where adequate sunlight was available for photosynthesis. As a result, plants never took control of the maritime ecology. But when those plants landed, everything was different. Before plants began to grow on land, there was nothing for other species to eat. But regrettably, until photosynthetic plants became established on the land, other species could not colonize it.

As far back as 700 mya, plants must have populated the area. The earliest fossil records of land plants that have been discovered so far stretch back roughly 470 million years. Land colonization was a significant turning point in the development of plants. Virtually all life had developed in the water up until that point. Additionally, the extreme and fluctuating temperatures on land, coupled with the still forming environment and intense solar radiation posed serious risks to land organisms thus increasing the likelihood of high mutations. This new environment presented a stark contrast to the ocean, making adaptation a formidable task for the first terrestrial organisms.

Evolution of vascular plants:

Plants evolved a variety of adaptations over time to help them cope with the difficulties of dry ground. One of the oldest and most important evolutionary events was the evolution of vascular tissues. Vascular tissues make up a plant's plumbing system. Both components are made up of phloem and xylem. Xylem transports water and minerals from the soil to the leaves for photosynthesis after being absorbed by the roots. Phloem transports food from photosynthetic cells to other cells for development or storage across the entire plant system. The advent of vascular tissues revolutionized the plant kingdom. This allowed plants to flourish in challenging terrestrial environments and develop into substantial structures. Legend has it that the earliest vascular plants resembled ferns. In addition to vascular tissues, these early plants also evolved lignin, leaves, roots, and a changed life cycle as adaptations to terrestrial life.

• Lignin is a hydrophobic, tough carbohydrate molecule. It gives the vascular tissues in the stem more support.

• Leaves are abundant in chloroplasts, which are crucial for photosynthesis, which is required to produce sustenance and protect plants from herbivores and parasites. Over time, the leaves, which were initially little and short, grew larger.

• Some plants have roots that may even pierce rock and soil. They take minerals and water from the earth and transfer them to the leaves. They serve to secure a plant to the ground. Rhizoids, which were used for absorption by nonvascular plants, gave rise to roots.

• In land plants, a dominant diploid sporophyte generation has emerged. Because diploid individuals are less likely to experience negative effects from mutations, this was an adaptive strategy.

With all these benefits, it is simple to understand why vascular plants spread fast around the globe. As vascular plants became increasingly common, many nonvascular plants became extinct. The main terrestrial plants on Earth today are vascular plants.

The changes brought about during a course of time in an organism which changes the gene and genotypic frequencies in an individual organism and the population as a whole is called evolution. It may be an increased or decreased resistance to a particular pathogen or increase in internode length in order to reduce plant height as a response to climate change. Any such gradual change that occurs at a cellular level that changes the characteristic features of an organism is called evolution. It can be either natural evolution due to natural selection for the survival of the fittest as per the prevailing environments conditions or induced by man via artificial means to meet the prevailing market needs for the particular crop. Natural forces move slowly and at their own pace, which is only apparent to the human eye when a lengthy timeline of evolutionary changes is written out in its entirety. On the other hand, induced or artificial evolution is a different matter. The properties of the plant are typically changed quickly by a driving force, or a combination of driving forces, depending on what the breeder needs. These forces will be described later. Measured in terms of morphological, physiological, biochemical, anatomical, embryological, and genetic modifications, all these changes brought about by these forces

All variations resulting from evolution incorporated various modifications. Which are:

1. Arbuscular mycorrhizal symbiosis, the cuticle, stomata, and intercellular gaps, the xylem, and the endodermis are examples of the evolution of plant anatomy.

2. Changes in plant morphology which includes leaf, root, tree form, seed and flower

- 3. The evolution of photosynthetic pathways
- 4. Transcriptional regulation evolution
- 5. Secondary metabolism's evolution

The forces that drive evolution

1. Natural Selection

Natural selection is characterised by negative selection, also known as purifying selection. Its function is to eradicate harmful alleles or mutations from the population gene pool, lowering their frequency in proportion to their biological impact. As a result, when compared to synonymous mutations, fatal, nonsynonymous, or nonsense mutations are rapidly purged from the population gene pool. Less damaging mutations that have a lesser influence on gene expression, on the other hand, are found in lower frequency in the population. Because of its moderate influence on these mutations, negative selection has a limited impact on genetic diversity in the population gene pool. Positive selection, often known as Darwinian selection, is another type of natural selection. Natural selection favours beneficial genetic changes that improve an individual's fitness or survival. Positive selection increases the frequency of such variations in the population gene pool, promoting genetic diversity both directly and indirectly by increasing the frequency of genetically connected variants via genetic draught or genetic hitchhiking mechanisms.

2. Mutation

Mutation refers to the sudden, heritable changes in an organism's DNA. They serve as vital sources of variability in a population and occur naturally or can be induced through physical or chemical means. In natural selection, minor mutations that confer immediate survival advantages and contribute to an individual's fitness are favored. Over time, such mutations accumulate and give rise to entirely new characteristics in an organism, driving evolutionary changes. Induced mutations are sometimes employed for immediate results in breeding programs, which are later incorporated into a population, leading to the development of new traits and contributing to long-term evolution.

3. Polyploidy

Polyploid denotes a change in the number of chromosomes in an organism. It comes in two types:

a. Autopolyploidy: Involves a change in ploidy within the same species and is also known as simple polyploidy or single-species polyploidy. It can occur naturally or be induced. Examples include autotriploid banana, apple, watermelon, and sugarbeet.

b. Allopolyploidy: Involves a change in ploidy between different species and is also known as hybrid polyploidy, bispecies polyploidy, or multispecies polyploidy, depending on the number of species involved. Allopolyploidy has played a significant role in evolution, with 50% of crop plants being allopolyploids.

4. Introgression

Intogression involves the incorporation of a gene from one species into the genetic background of another species through interspecific hybridization and backcrossing (Anderson, 1949). This process can lead to the combination of genes from two divergent species, and any recombination that is favored by natural selection may give rise to a new trait or even a new species. An example of this is the evolution of modern forms of maize, which are believed to have occurred through introgressive hybridization between primitive maize and Tripsacum.

Molecular evolution

Only when internal processes are altered do morphological alterations take place. These internal mechanisms were altered as a result of the ongoing molecular evolution of the individual organism. It involves altering the sequence of biological molecules including DNA, RNA, and proteins from generation to generation. These patterns of change are explained by the field using concepts from population genetics and evolutionary biology. The main topics of molecular evolution are:

- i. Rate and effect of single nucleotide changes.
- ii. Neutral evolution versus natural selection.
- iii. Origin of new genes.
- iv. The genetic nature of complex traits.
- v. The genetic basis of speciation.
- vi. Developmental evolution, and;
- vii. The ways in which evolutionary forces influence genomic and phenotypic change.

The roots of molecular evolution can be traced back to the early 20th century when comparative biochemistry emerged. In the 1950s, "fingerprinting" techniques like immunoassays, gel electrophoresis, and paper chromatography were applied to study homologous proteins (Dietrich 1998, Hagen 1999). The field of molecular evolution saw significant growth during the 1960s and 1970s, driven by advancements in molecular biology. Molecular biologists began sequencing proteins, enabling the construction of phylogenies based on sequence comparisons. Additionally, they utilized differences in homologous sequences as a molecular clock, allowing estimation of the time since the last universal common ancestor.

So, what exactly is a molecular clock? A molecular clock is a metaphorical term used to describe a technique that utilizes the

rate of biomolecule mutations to infer when two or more life forms diverged in prehistoric times. This technique involves analyzing DNA nucleotide sequences, RNA, or protein amino acid sequences as data. Fossil or archaeological dates are often used as reference points for determining mutation frequencies. The concept of the molecular clock was first tested in 1962 using different animal hemoglobin protein variants and has since been widely used in molecular evolution to estimate the time of speciation or radiation. It is also referred to as the genetic clock or evolutionary clock.

The Power of Molecular Evolution:

The content and structure of a genome is the result of genetic molecular and population forces acting on that genome. Just as we learned about the forces of nature that drive evolution, there are also forces that influence the evolution of molecules.

Forces in molecular evolution:

1. Mutations:

Mutations are inherited, permanent alterations to the DNA or RNA that make up a cell's genetic code. Errors in DNA replication during cell division, as well as exposure to radiation, poisons, viruses, transposable elements, and other environmental factors, are the causes of these disorders. Single nucleotide polymorphisms, which change just one base in the DNA sequence and result in point mutations, make up the majority of mutations. Other mutations can cause larger DNA regions to undergo duplications, insertions, deletions, inversions, and translocations. Numerous genes regularly experience random stochastic mutations. Single nucleotide site mutation rates are relatively high in the majority of species. Some of these mutations are advantageous or neutral and remain in the genome if they are not lost to genetic drift, whereas others are detrimental and are removed from the genome by natural selection. Due to their rarity, mutations accumulate across generations at a relatively slow rate. Despite the fact that the number of mutations per generation can vary, they seem to accumulate steadily over very long time scales.

2. Recombination:

Genetic rearrangement, the interchange of genetic material across distinct species, results in children with traits that are different from both of their parents. Meiosis-induced genetic recombination in eukaryotes can result in a brand-new set of genetic material that can be passed from parents to children. Most recombination occurs in the natural world.

3. Gene conversion:

Gene conversion is a type of recombination that happens as a result of DNA repair and fixes a nucleotide error using a homologous genomic region as a template. DNA synthesis excises the damaged bases first, aligns the damaged strand with its homologue, and then uses the undamaged strand as a guide to repair the excised part in order to repair broken bases. The long-term homogeneity of duplicate gene sequences, which reduces nucleotide divergence, is typically caused via gene conversion.

4. Genetic Drift:

Genetic drift, which occurs in finite populations as a result of stochastic effects from random sampling, is the variation in allele frequency from one generation to the next. There are several current versions that have no harmful impact on health, yet they can occasionally increase or decrease the frequency.

5. Selection:

Selection happens when organisms that are more fit, or have a higher capacity for survival or reproduction, are favored in succeeding generations, increasing the prevalence of underlying genetic variants in a population. Natural selection, artificial selection, and sexual selection are all examples of selection. Natural selection is any form of selection that arises from an organism's environmental adaptability. Sexual selection, in contrast, is a result of mate choice and can encourage the transmission of genetic variants that go against natural selection but increase attractiveness to the opposite sex or boost mating success. Artificial selection, often known as selective breeding, aims to enhance the frequency of desired features through the imposition of artificial selection by an external agent, usually a human.

6. Intergenetic conflict:

When genes have phenotypic effects that favor their own transmission at the expense of the transmission of other genes that share the same genome, this is referred to as intragenomic conflict. "Selfish genes" are these kinds of genes. According to the selfish gene theory, natural selection will favor genes whose phenotypic effects result in their transmission to new organisms. Most genes accomplish this by working together with other genes in the same genome to create an organism that is capable of reproducing and/or aiding kin in reproducing.

The explanations for molecular evolution are as follows:

Three viewpoints offer evolutionary explanations for molecular evolution, depending on how much weight is given to the various forces of evolution.

1. Selectionist Hypothesis:

Selection, in accordance with selectionist theories, drives molecular evolution. Selectionists contend that, despite the fact that many mutations are neutral, changes in the frequency of neutral alleles are rather due to linkage disequilibrium with other loci that are under selection than to chance genetic drift (Hahn, 2003). It is common to talk about codon use biases in terms of how even slight selection might affect molecular evolution.

2. The Neutralist Hypothesis:

According to the neutral theory of molecular evolution, the majority of evolutionary changes at the molecular level as well as the bulk of diversity within and across species are caused by random genetic drift of mutant alleles that are selectively neutral. It is in line with Charles Darwin's notion that natural selection affects phenotypic evolution, but it only applies to molecular evolution. The neutral strategy acknowledges that most mutations might be detrimental but argues that because natural selection quickly eliminates them, they have little impact on molecular variation both within and between species. Any mutation is considered neutral if it has no effect on an organism's ability to reproduce or survive. The majority of mutations that are benign are neutral, rather than favorable, according to the neutral theory. According to the neutral theory, since only a small portion of gametes are sampled from each generation of a species, a mutant allele may evolve within a population and become fixed by chance rather than through selective advantage (Kimura, 1983).

In 1968, Japanese biologist Motoo Kimura made the initial theory-proposal. Two American biologists, Thomas Hughes Jukes and Jack Lester King, independently advanced it in 1969. In his 1983 monograph, The Neutral Theory of Molecular Evolution, Kimura went into extensive detail. An extensive "neutralist-selectionist" debate about how to interpret patterns of molecular divergence and gene polymorphism followed the neutral theory's suggestion, peaking in the 1970s and 1980s.

3. Mutationists hypothesis:

Biases in mutation patterns and random drift are heavily emphasized in mutationist theories (Nei, 2005). Sueoka was the first to advance a modern mutationist perspective. His idea (Sueoka, 1964) postulated that GC mutation pressure, not positive selection, was responsible for the variance in GC content.

Methods for analysis of evolution:

1. Morphological assessment

It entails comparing an organism to its ancestors or progenitors in order to determine the types and degrees of differences that have developed. If the ancestors are extinct, a comparison with their fossil records is also included.

2. Molecular assessment

In order to identify the causes of the accumulated discrepancies, it involves comparisons based on molecular marker and sequencing technologies.

3. Structural assessment

Involves analyzing the molecule's structure and comparing it to other compounds in order to determine its evolutionary links. **Evolutionary analysis software:** • The most widely used program for evolutionary analysis was created by researchers at the University of Washington in Seattle's Department of Genome Sciences and Department of Biology, and is called the Phylogeny Inference Package (PHYLIP). It is a free software program that analyzes molecular sequences using several techniques.

• The free software program Hypothesis Testing Using Phylogenies (HyPhy) is used for phylogenetic analysis of biological sequences, particularly for determining the degree of selection from sequence data.

• MEGA is a widely used software program for analyzing how organisms evolve at the molecular level. For academicians, various versions of this program are provided without charge. The majority of the algorithms used in evolutionary biology are implemented through a variety of methods and programs.

It is clear that the plant species we can see in front of us now are the product of millions of years of evolution coming together to create the intricacy that exists. The evolutionary changes that have given plants their many features have been shaped through selection, mutation, introgression, and genetic drift. Additionally, a quick overview of molecular evolution in the chapter made it clear that molecular alterations are the primary causes of the morphological changes in organisms and plants. After all, genetic variations serve as the fundamental units of variety, which also holds true for evolutionary variations. In our high-tech environment, new programs and software are constantly being created in addition to those that have already been described. This facilitates and makes interesting the study of evolution.

References:

Dietrich, M. R. (1998). Paradox and Persuasion: Negotiating the Place of Molecular Evolution within Evolutionary Biology. *Journal of the History of Biology*, **31**: 85–111. https://doi.org/10.1023/A:1004257523100.

- Hagen, J. B. (1999). Naturalists, Molecular Biologists, and the Challenges of Molecular Evolution. *Journal of the History of Biology*, 32(2): 321 341.
- Nei, M. (2005). Selectionism and Neutralism in Molecular Evolution. *Molecular Biology and Evolution*, 22(12): 2318–2342. <u>https://doi.org/10.1093/molbev/msi242</u>
- Sueoka, N. (1967). Synchronization of Chromosome Replication and Gene Function During Spore Germination of *Bacillus subtilis. The American Naturalist*, 101(**920**): 317-319.
- Kimura, M. (1983). *The Neutral Theory of Molecular Evolution*. Cambridge: Cambridge University Press. <u>https://doi.org/10.1017/CBO9780511623486</u>
- Hahn, M. W., Stajich, J. E., Wray, G. A. (2003). The effects of selection against spurious transcription factor binding sites. *Molecular Biology and Evolution*, 20(6):901-6. <u>https://doi.org/10.1093/molbev/msg096</u>
- Kimura, M. and Ohta, T. (1971). Protein polymorphism as a phase of molecular evolution. *Nature*, 229: 467–469.
- King, J. L. and Jukes, T. H. (1969). Non-Darwinian evolution. Science, 164: 788-798.

02

Magical World of Carnivorous Plants

Kavya V. Yankati

MSc in Botany, Karnataka University Dharwad. Corresponding Author email:- <u>yankatikavya@gmail.com</u>

Abstract

Carnivorous plants are the most wonderful creatures of the earth. These found only in lack of soil-nutrients. They confluently evolved nearly 2-4 times in the history of evolution. In order to trap their prey they adopt leaves. The current survey on these flesh-eating plants. Reported that there are nearly 630 species, found conveniently. Because of poor-nutrients in the soil, they derive their nutrients from these animals. Thus they are benefited from the animals and other small creatures. The most flesh-eating plants can consume reptiles, small mammals, large plants and in some rare cases humans too. Small carnivorous plants can accumulate protozoa and bacteria which are single celled organisms. Aquatic carnivorous plants also feed on mosquito larvae, small fishes and crustaceans etc., According to the report given by Flokerts in 1982, A single bog contain nearly 13 species of the carnivorous plants In 1857, Charles Darwin, the very first person to classify carnivorous plants on phylogenetic tree. Some of the important aspects that we learn here are as follows., Key words: Carnivorous plants, traps, Insectivorous, poor-nutrient. 1. Introduction:

Carnivorous plants can be seen through a different perspective which provides an ideal example of how traits develop through natural selection and promotes the impressive growth and reproduction. These included in the era of biology based on evolution by Darwinism in a specific environment. Currently the carnivorous plants have been considered as one of the most endangered group of plants i.e., they are in the urge of extinction. For biologists, the incogitable accuracy and because of the occurrence of those adaptations the obtained final product is phenomenal. Incomparable example provides perfect mechanisms of that. Nature can tackle to sustain in those extreme conditions. A single glance at them allows the perception that carnivore; it is not specific to them. It provides a brief knowledge of understanding their adaptations, and their transformation strategy, of both morphologically and as well as functional nature as evidenced in carnivorous plants.

Plants have been considered as carnivore, if they have 5 basic adaptive characters.

- Ability to capture the prey by attracting it.
- Killing the captured prey.
- Digesting the prey.
- Absorbing the nutrients from that prey.
- -. Using those absorbed nutrients to development and growth.

2. What are carnivorous plants?

These are also known as insectivorous plants which are capable of consuming and digesting the insects by means of pitfalls and traps. The major families of carnivorous plants include Lentibularaceae of order lamiales which is characterized by flowers with bilaterally symmetrical. Petals are fused, have only two anthers. This family is fairly ubiquitous in distribution.

3. Some prominent characters of carnivorous plants:

a. Deficiency of nitrogen: These types of plants are most likely found in the places where there is a poor-nutrient content in the soil. Especially the nitrogen deficiency, thus they get nutrition to survive from the insects or animals they hunt. Important fact about these carnivorous plants is "They contain a compound that usually has antifungal properties, that acts as effective against the infections.

b.Way of attracting the prey: The strategies that carnivorous plants adapt in order to attract their prey are- release a strong odour of nectar, due to their ardent colouration of the flowers that attracts the insects and other arthropods.

c. Inevitable traps: Insectivorous plants develop specialized organs that help them in trapping the insects and other organisms. The mouth of the carnivorous plants contains hair-like edges that enclose immediately as the insect's touches the hairs. Sticky mucous like substances will cover the stalk of the plants. And once the insects fix they cannot move.

d. Digestive enzymes: some of the carnivorous plants have digestive enzymes in their digestive tracts. They completely diffuse the organisms for absorbing nutrients from it. They literally imitate the activity of the human digestive part.

e. Wet and damp habitats: these carnivorous plants are constantly found in places which contain humid, acidic conditions of the soil, wet and damp etc. These contain poor-nutrient contents such as bogs, wetlands, coastal-plains, swamps etc. They are found to be introduced in the Australia and tropical regions of the world and wet regions of North America.

4. Evolution of the carnivorous plants:

At the end of the 19th century, lurid tales of tales of toxic plants started to manifesting up everywhere. The trees with horrifying grappler (tentacles) seized and swallowed. travellers who are in great distance away from them. Some insane professors put up some

heinous sundews and pitcher plants, these turned into famished creations and destroyed everything.

Most people supressed to agree that plants can also eat animals because it is against the natural order of the environment. Darwin expended 16 years of his life in performing these experiments and he finally proved that plants can move and capture and consume small insects and prey's, and they also digest them and get nutrients to survive. Darwin published his book named "Insectivorous plants" in the year 1857 in which he included all the details of his experiment and researches that he had discovered.

5.Some important examples of carnivorous plants:

SUNDEW: Drosera: International plant, Drosera is a venomous plant which causes harm to the animals and in some severe cases this plant can kill. It is the genus with most varied among all carnivorous and includes about two hundred different species found broadly. These ranges from tropical sundews to subtropical sundews and temperate sundews, includes cape sundews, fork-leafed sundews etc. The trap:These trap the prey by sticky hairs on their leaves, containing sticky gland at their tips. These droplets, glistening in sun commonly called as 'dew.

The flowers: The flowers of the sundews are very attractive because of their fascinating nature and unique beauty. It vary in shape,size, colour, time, and amount of seed produced. The flowers occurs in



delightful shades of pink, red or



Magical World of Carnivorous Plants



white colours. The sundews are the largest group of carnivorous plants and the flowers of these plants can share usually common reproductive structures.

Ya-te-veo Plant: Universally called as man-eating plant.

It kills and acquire the humans as well as large animals. This carnivore plant was first attested in african and central american forest in the year 1874. Ya-te-veo is a spanish pharse meaning –"Now I see you". They incude spines which are highly venomous and

contain many vast serpents indicating their angry nature. Flitting their terrible tentacles side by side, Then snatch and swallow any creature coming near it. [In James W. Buels sea and land 1889].

Dionaea muscipula: Commonly



called as venus fly trap. It is a carnivore flowering plant. It trap the prey with the help of the two lobes. Trichomes, are the hair like projections which are found in the inner surface of the lobes. These trichomes helps the lobes to shut when the prey come within it and this whole movement is called as Thigmonasty. This venus fly trap is endemic plant species because it is listed as vulnerable internationally.

Range: Native to subtropical wetlands on the east coast of the united states in south carolina and the North carolina. This Plants grows in the moisture containing



acidic soil. They get nutrients from the soil but poor in nutrient supply. Inspite of this, these plants eats small insects and arachind, ants,spiders etc. Important Fact about this plant is that "It drives energyfrom the sun in the process of photosynthesis" The life cycle of this venus fly trap:It undergo many changes during its life cycle and these changes includes certain stages such as,

1]Stage 1: seed 2]Stage 2: Seedling 3]Stage 3: young plant 4]Stage 4: Adult plant

Pitcher plant: It is a carnivory plant that have leaves modification to trap the insects. These plants attracts the insects or prey with their nector. This pitcher plant is the member of the family named Nepenthaceae and Sarraceniaceae.

Attraction: The crawling and flying insects such as bees or flies attracted to the beautiful structure of the plants, its cupped leaf, and unique visuals of anthocyanin pigments and also nectar. Olfactory cues also plays an very important role in attraction of the prey for example,Nepenthes rafflesiana uses flowers-fragrance to attract insects.

6.Root system in carnivorous plants:

The adult plants of carnivorous plants show a great variety in root system. Radical is quite identical and found in only limited number of species. A very best example for this is, radical displaced by stem borne roots immediately after germination. Surprisingly, very long

root hairs develop which are deep rooted. Symbolic representation of the numerous roots and underground organs of a diverse variety of carnivorous plants are entailed in pietropaolo (1986). Carnivorous species includes wide range of root types. In literature survey the



numerical data of size and morphology are rarely found.

The descriptions about the roots are mainly based on 3 types that is,

- 1. Fragile
- 2. Weak
- 3. Strong

For the sake our observations we can divide or use the term, Weak for discreet roots which are smaller than the ground part of the plant. And strong for the well-developed roots of the plant. According to the evidence given by slack 2000, the roots can function permanently or about part of the year.Franca1925 and Menninger 1965 included

in their articles that some plant species have deep roots that most likely allows perennial process of water uptake. And according to the Nitschke



1860. Some plant species have deficient root development or lack of roots as given by slack 2000 and Taylor 1994.

Bybilis and Drosophyllum are the examples of radical of carnivorous plants. These have a wide range of root system. The radical barely able to report the seedlings to the substrate, so that at the total length of 30mm the plant can be relocated smoothly.

Correspondingly in the plant of Nepenthes gracilis, the radical is thin. It forms a cluster of deep roo hairs into the testa. The roots are very weak to produce sufficient amount of water and nutrients and as well as to anchor the seedlings.

Root system in aquatic carnivorous plants:

Aquatic carnivorous plants do not develop roots. The most prominent

this examples for is the Aldrovanda that bears traps.Polypompholyx and various Utricularia species have suctiontraps, the Genlisea which is closely realated species that acts as predator to capture its prey with its eel



traps. These take a little amount of minerals and nutrients from their leaves and stem.

Many terrestrial and epiphytic species are present in utricularia that grows in the moisture containning soil. This engulfed during a part of

the year.In utricularia and genlisea,There is

absence of developing roots.Instead they are replaced by specialized underground shoots and leaves.

7.Types of traps and their trapping mechanism:



There are basically 5 types of traps in the carniorous plants and they are mainly based on wheather they posses the ability to move and capture the prey or not,transversly called active and passive traps.This type of classification of traps is given by the botanists Francis Ernest

Lloyd in1942. This type of classification is followed till today. In order to attract the prey they do not posses movement.

This type of trap include 3 main types

1.Pitfall traps

- 2. Passive flypaper traps
- 3. Lobster pot traps

1.Pitfall traps: These are the simplest type of traps and contain nectar as a part of its potion. This nectar produces at the entrance of

the trap. They are very darkly coloured. The leaves contain a moist medium that digest the prey. Once the prey got trapped and reach the underground surface of the plant, then its highly immpossible to



evade from it. A very known examole for thid type of trap is Sarracenia purpurea also called as purple pitcher plant.





Passive flypaper: Here the

sticky substances are secreted by a special type of glands. For example byblis,Australian genus.

obster pot traps: These are somewhat similar as pitfall traps. In this type of traps, it is easy for prey to get in, but difficult to go outfrom the trap.

Example: Sarracenia psittacina, contains leaves with tubular shape, enclosed at the underpart.act as a one-way valve.

Active traps:In order to attract the prey they posses a movement. This includes 3 main types of traps.

-Active flypaper trap

-Bladder trap

-Snap trap

1. Active flypaper trap: Same as in passive flypaper;secreation of sticky-jelly like substances to trap the insects.

• If once it caught, the edges of the leaves around the prey enclose.



- Slow movement of capturing the prey takes place.
- Prevents the lack of nutrients containing liquid or moist medium.

Example:Sundew (Drosera) and butterflies (Pinguicula).

2.Bladder traps: Usually these type of the traps are used by the aquatic carnivorous plants and contains 'Bladder' that are dense.

- The trap door opens when the prey touches the hairs of that door.
- The water flows inside when the trap door is opens, This type of traps seen in uticularia species.

3.Snap trap: This is most common type of traps in carnivorous plants. Example:venusflytrap.

Family	Genus	Number of	Type of
		species	trap
Sarraceniaceae	Sarracena	8**	Pitfall,
			Lobster
			trap
Nepentheceae	Nepenthes	60**	Pitfall trap
Byblydiaceae	Byblis	2	Passive
			flypaper
Droseraceae	Drosophyllum	1	Passive
			flypaper
	Drosera	About	Passive
		90**	flypaper
	Dionaea	1	Snap trap
	Aldovanda	1	Snap trap
Lentibulariaceae	Genlisia	16**	Lobster
			trap

9. Recent satus of carnivorous plants in table manner:

Uticularia	About	Bladder
	250**	trap
Polypompholyx	2**	Bladder
		trap

Conclusion: From several decades its an intresting for a botanist to survey on the carnivorous plants. But still it seems to be incomplete survey. These type of carnivorous plants has been utilized for various miniature of ecological and evolutionary relationships. They show unique charaters from all other normal plants. Their adaptation to survive in the extreme conditions, such as acidic soil condition, poor nutrient contents. Their spec ialized way to trap the prey. How they derive the nutrients and energy from the organisms they hunt. The digestion process of prey in their body. These all things creates intrest to know more about these carnivorous plants. In future generation more studies have to be done on the modification in leaves. More facts about the roots of carnivorous plant. Their survival feature in extreme climatic conditions is to be noticed. Most of these plant, in present day, are in urge of extinction. Measures should be taken in order to conserve these carnivorous plants, so that the plants do not become vulnerable.

Reference:

- Lotzof, B. K. (n.d.). Carnivorous plants: the meat-eaters of the plant world. *Natural History Museum*.
- Mithofer, A. (2011). Carnivorous pitcher plants: Insight in an old topic. *Phytochemistry*, 1678-1682.
- Mithofer, A. (2022). Carnivorous plants and their biotic interactions. *Journal on plant interactions*, 333-343.
- R, H. (2015). Carnivorous plants. European PMC.

Stephanie Pain, K. M. (2022). How Carnivorous Plants Evolved. Smithsonian magazine.

Why the carnivorous plants are "Most wonderful plants in the world". (n.d.). - *Natural Science editorial paper 2nd round of edi*.

wolfram adlassnig, M. K.-P.-S. (2005). The roots of the carnivorous plants. *Plant and soil*, 127_140.

3

Plant Secondary Metabolites: Functions and Applications ¹Priyanka Choudhary ²Sumitra Kumawat

¹Department of Soil Science, Dr Rajendra Prasad Central Agricultural University, Pusa, Samastipur, Bihar 848125 ²Division of Soil Science, ICAR-Indian Agricultural Research Institute, New Delhi

Introduction

Plant secondary metabolites are organic compounds that are produced by plants, but are not essential for their growth or development. Unlike primary metabolites, such as carbohydrates, proteins, and lipids, which are necessary for basic plant functions, secondary metabolites are synthesized for various protective and adaptive purposes. Secondary metabolites play crucial roles in plants' interactions with their environment. They are involved in defence against herbivores, pathogens, and competing plant species. They can also attract pollinators and seed dispersers, as well as act as signalling molecules for inter-plant communication.

There is a wide variety of plant secondary metabolites, including alkaloids, flavonoids, terpenoids, phenolic compounds, and glycosylates. Alkaloids, like caffeine and nicotine, are known for their stimulant and toxic effects. Flavonoids, such as quercetin and catechins, have antioxidant properties and contribute to the vibrant colours of many flowers. Terpenoids, like essential oils, are responsible for the characteristic scents of plants. Phenolic compounds, such as tannins and lignin, provide structural support to plants and have antimicrobial properties.

Secondary metabolites are not only of interest to plant biologists, but also to researchers in medicine, pharmacology, agriculture, and ecology. They have been exploited for their medicinal properties, serving as a source of drugs and natural remedies. Many pharmaceuticals, such as morphine and artemisinin, are derived from plant secondary metabolites. In agriculture, secondary metabolites can be used as natural pesticides or growth regulators. Their ecological roles are also important, as they influence species and ecosystem dynamics. Understanding interactions the biosynthesis, function, and ecological significance of plant secondary metabolites is a fascinating field of research. It contributes to our knowledge of plant evolution, adaptation, and chemical ecology. Furthermore, the discovery and utilization of these compounds hold great potential for the development of new drugs, agricultural products, and environmental solutions.

2. A. Definition and classification of secondary metabolites

Secondary metabolites are organic compounds produced by organisms, including plants, fungi, and bacteria, that are not directly involved in their primary metabolic processes, such as growth and reproduction. These compounds are often involved in various ecological roles, including defence against predators, communication with other organisms, and attraction of pollinators. Secondary metabolites can be classified into different groups based on their chemical structure and properties. Some common classes of secondary metabolites include:
l. Alkaloids: These are nitrogen-containing compounds known for their diverse pharmacological activities. Examples include caffeine, morphine, and nicotine.

2. Terpenoids: Also known as isoprenoids, terpenoids are hydrocarbons derived from a five-carbon isoprene unit. They can be found in plants and some microorganisms. Examples include essential oils, such as limonene and menthol.

3. Phenolics: These compounds are derived from phenylalanine and are widely distributed in the plant kingdom. They include flavonoids, tannins, and lignin, which have various roles in plant defence and pigmentation.

4. Glycosides: These are compounds that consist of a sugar molecule attached to a non-sugar compound, such as a phenolic or aglycone moiety.

5. Polyketides: These are complex organic compounds derived from repeated condensation of simple building blocks, such as acetate or propionate units.

6. Phenazine: Phenazines are nitrogen-containing heterocyclic compounds produced by certain bacteria and fungi.

7. Flavonoids: Flavonoids are a class of plant secondary metabolites widely distributed in the plant kingdom.

B. Differences between primary and secondary metabolites

Primary metabolites are essential for the growth, development, and reproduction of an organism. They play a vital role in metabolic pathways and are required for basic cellular functions. Examples of primary metabolites include amino acids, nucleotides, sugars, and organic acids.

Here are the key differences between primary and secondary metabolites:

l. Function: Primary metabolites are involved in essential metabolic processes, while secondary metabolites serve various ecological roles such as defence and communication.

2. Occurrence: Primary metabolites are found in all living organisms and are essential for survival, whereas secondary metabolites are not universally present and vary widely among different species.

3. Regulation: Primary metabolites are typically constitutively produced and are tightly regulated by the metabolic needs of the organism. Secondary metabolites are often produced in response to specific conditions or stimuli.

4. Abundance: Primary metabolites are generally present in higher quantities compared to secondary metabolites, as they are required for basic cellular functions. Secondary metabolites are often present in lower concentrations.

5. Structure: Primary metabolites consist of simple compounds like amino acids, sugars, and organic acids. Secondary metabolites, on the other hand, are often complex and structurally diverse, with a wide range of chemical structures.

6. Distribution: Primary metabolites are usually evenly distributed throughout the organism, while secondary metabolites may be localized in specific tissues or organs.

7. Evolutionary conservation: Primary metabolites are highly conserved across different species, while secondary metabolites can vary greatly, even within closely related organisms.

Il. Functions of plant secondary metabolites

Plant secondary metabolites are organic compounds that are not directly involved in the plant's growth, development, or reproduction. Instead, they play various roles in the plant's interactions with its environment. Some of the functions of plant secondary metabolites include:

1. Defence against herbivores: Many secondary metabolites act as chemical defences against herbivores. These compounds can be toxic or deterrent to feeding insects, mammals, and other herbivores.

2. Protection against pathogens: Secondary metabolites can also act as antimicrobial agents, protecting plants from fungal, bacterial, and viral infections.

3. Attraction of pollinators: Some secondary metabolites, such as floral scents and pigments, play a role in attracting pollinators to help with plant fertilization

4. Allelopathy: Secondary metabolites can inhibit the growth of nearby plants, helping the producing plant to gain a competitive advantage in resource utilization.

5. UV protection: Certain secondary metabolites, like flavonoids, act as sunscreens, protecting plants from harmful UV radiation.

6. Stress response: Plants produce secondary metabolites in response to various environmental stresses, such as drought, temperature extremes, or nutrient deficiencies. These compounds help the plant adapt and survive under adverse conditions.

7. Signal molecules: Secondary metabolites can act as signaling molecules within plants or between plants and other organisms, influencing various physiological processes.

8. Medicinal properties: Many secondary metabolites have medicinal properties and are used in traditional or modern medicine for their therapeutic effects. Overall, plant secondary metabolites contribute to the survival, fitness, and ecological interactions of plants in their environments.

A. Defence against herbivores and pathogens

Plants have evolved various strategies to defend themselves against herbivores and pathogens. One common defense mechanism is the production of secondary metabolites, which are compounds that are not essential for the plant's growth and development but play a crucial role in defense. Here are some ways in which secondary metabolites help plants defend against herbivores and pathogens:

1. Toxic compounds: Many secondary metabolites are toxic to herbivores, making them avoid feeding on the plants. For example, alkaloids, such as nicotine in tobacco leaves, are toxic and act as potent feeding deterrents.

2. Anti-feedants: Certain secondary metabolites act as antifeedants by reducing the palatability of the plant to herbivores. They can alter the taste or texture of the plant, making it less appealing to feed on. For instance, tannins found in many plants make them taste bitter and unappetizing to herbivores.

3. Allelopathy: Some secondary metabolites have allelopathic effects, which means they inhibit the growth or development of neighboring plants.

4. Antimicrobial activity: Secondary metabolites also play a crucial role in protecting plants against pathogens, including bacteria, fungi, and viruses. These compounds often have antimicrobial properties that inhibit the growth and proliferation of

pathogens. For example, some plants produce phytoalexins, which are antimicrobial compounds that are induced in response to pathogen attack.

5. Induced defense: Secondary metabolites can also be produced in response to herbivore or pathogen attack. When plants detect the presence of herbivores or pathogens, they can activate defense responses, including the production of specific secondary metabolites.

B. Role of secondary metabolites in plant defense mechanisms

Secondary metabolites play a crucial role in the defense mechanisms of plants. These chemical compounds are produced by plants and are not directly involved in their growth, development, or reproduction. Instead, they serve various purposes, including defense against pathogens, herbivores, and other environmental stresses. The role of secondary metabolites in plant defense can be broadly categorized into two main functions: direct defense and indirect defense.

l. Direct defense: Secondary metabolites directly deter herbivores and pathogens from attacking the plant. For example, some secondary metabolites act as toxins or repellents, making the plant unpalatable or poisonous to herbivores.

2. Indirect defense: Secondary metabolites indirectly protect plants by attracting natural enemies of herbivores or pathogens. When plants are under attack, they can release volatile secondary metabolites, also known as herbivore-induced plant volatiles (HIPVs). These compounds attract parasitoids, predatory insects, or other organisms that prey on herbivores or pathogens. The natural enemies are attracted by the HIPVs and subsequently attack or parasitize the herbivores, reducing their population and damage to the plant.

.C. Attraction of pollinators and seed dispersers

Secondary metabolites are chemical compounds produced by plants that serve various functions, including defense against herbivores, attraction of beneficial organisms, and communication with other plants. These compounds play a crucial role in attracting pollinators and seed dispersers to ensure successful reproduction for plants.

Pollinators, such as bees, butterflies, and birds, are attracted to flowers primarily by their visual cues, including color, shape, and patterns. However, secondary metabolites also play a significant role in attracting pollinators through their scent and taste. Many plants produce volatile secondary metabolites, such as terpenes and phenols, which emit attractive fragrances that can be detected by pollinators from a distance. These scents act as chemical signals that guide pollinators towards the flowers, increasing the chances of successful pollination.

In addition to attracting pollinators, secondary metabolites also play a role in attracting seed dispersers, such as birds and mammals. Fruits, which serve as containers for seeds, often contain secondary metabolites that make them visually appealing and flavorful to seeddispersing animals. The bright colors of fruits, caused by pigments such as anthocyanins, attract animals by contrasting with the surroundings and signaling the presence of high-energy food.

i). Examples of secondary metabolites involved in pollination and seed dispersion

Several secondary metabolites are involved in pollination and seed dispersion. Here are some examples:

1. Anthocyanins: These pigments give flowers their vibrant colors and attract pollinators like bees, butterflies, and birds. By luring pollinators, anthocyanins play a crucial role in ensuring successful pollination and seed production.

2. Alkaloids: Some alkaloids found in plants can repel or deter herbivores, but they can also attract pollinators. For example, caffeine in nectar acts as a reward for bees and improves their longterm memory, making them more likely to return to the same flower species.

3. Terpenes: Terpenes are a diverse group of compounds responsible for the scents and aromas emitted by flowers. These fragrances attract specific pollinators, such as bees, wasps, moths, and beetles, which help in the transfer of pollen between flowers.

4. Tannins: Tannins are bitter-tasting compounds found in fruits, seeds, and other plant parts. They may deter herbivores from consuming fruits too early, allowing the seeds to mature fully. Additionally, tannins can aid in seed dispersion by attracting frugivores, such as birds or mammals, which consume the fruit and disperse the seeds through their excreta.

5. Resins: Resins are sticky substances produced by plants that serve various purposes, including defense against pathogens, pests, and herbivores.

D. Allelopathy: Definition and mechanism of allelopathy with Examples

Allelopathy refers to the ability of certain organisms, especially plants, to produce and release secondary metabolites that influence the growth, survival, and reproduction of neighbouring organisms. These secondary metabolites are chemical compounds that are not directly involved in the primary metabolic processes of an organism. The mechanism of allelopathy involves the release of secondary metabolites into the surrounding environment by the allelopathic organism. These metabolites can affect other organisms through various mechanisms, including inhibition of seed germination, inhibition of root elongation,

inhibition of photosynthesis, or interference with nutrient uptake.

Examples of allelopathy can be seen in various plant species. For instance, black walnut (Juglans nigra) releases a compound called juglone, which inhibits the growth of many other plant species around it. Similarly, sunflower (Helianthus annuus) releases allelochemicals called phytotoxins, which can inhibit the germination and growth of nearby plants. Another example is the invasive species, Centaurea solstitialis (yellow starthistle), which produces a toxin known as cnicin that inhibits the growth of surrounding vegetation.

Ill. Applications of plant secondary metabolites

Plant secondary metabolites have a wide range of applications in various fields. Some of the prominent applications include:

1. Pharmaceutical and healthcare industries: Many plant secondary metabolites possess medicinal properties and are used in the development of drugs and herbal medicines. For example, compounds like alkaloids, flavonoids, and terpenoids have been found to have antimicrobial, anticancer, and antioxidant properties.

2. Food industry: Plant secondary metabolites like phenolic compounds and flavonoids are important for the food industry due to their antioxidant and antimicrobial properties.

3. Agriculture and pest control: Some plant secondary metabolites have insecticidal and repellent properties, making them useful in pest control. Natural compounds such as pyrethrins and neem extracts are widely used as insecticides and biopesticides, reducing the reliance on synthetic chemical pesticides.

4. Cosmetics and personal care products: Plant secondary metabolites are used in the formulation of cosmetics and personal care products due to their antioxidant, anti-inflammatory, and antimicrobial properties. They are utilized in products like skin creams, lotions, shampoos, and perfumes.

5. Environmental applications: Plant secondary metabolites can be used in environmental cleanup and remediation. Certain compounds, such as phenolic acids, can aid in the degradation of pollutants and the removal of heavy metals from contaminated soils and water bodies.

A. Medicinal uses

1. Traditional medicine and herbal remedies

Traditional medicine has been practiced for centuries by indigenous cultures around the world. It involves the use of plant materials and natural substances to treat various ailments and promote overall well-being. Plant secondary metabolites, also known as phytochemicals, play a significant role in the therapeutic properties of many traditional medicines. Herbal remedies derived from plant secondary metabolites have been used to treat a wide range of health conditions.

Alkaloids are nitrogen-containing compounds found in many medicinal plants. They have diverse pharmacological activities and can be potent analgesics, anti-inflammatories, or antimicrobial agents. Examples of alkaloids commonly used in traditional medicine include morphine from opium poppy (Papaver somniferum) and caffeine from coffee (Coffea species).

Flavonoids are a large group of plant secondary metabolites that are widely distributed in fruits, vegetables, and medicinal plants. They are known for their antioxidant, anti-inflammatory, and immune-modulating effects. Some well-known flavonoids include quercetin from onions (Allium cepa) and epigallocatechin gallate (EGCG) from green tea (Camellia sinensis).

Terpenoids are another class of plant secondary metabolites with diverse biological activities. They are often responsible for the distinctive scents and flavors of plants. Many terpenoids have shown antimicrobial, anti-inflammatory, and anticancer properties. Examples of terpenoids used in traditional medicine include menthol from peppermint (Mentha piperita) and artemisinin from sweet wormwood (Artemisia annua).

2. Drug discovery and development using secondary metabolites

Drug discovery and development using secondary metabolites refers to the process of identifying, isolating, and developing potential drugs from naturally occurring compounds produced by organisms such as plants, microbes, and marine organisms. Secondary metabolites are chemical compounds that are not essential for the survival of the organism, but often serve important ecological functions such as defense against predators. This approach to drug discovery has gained attention due to the vast structural diversity and biological activity exhibited by secondary metabolites. These compounds have shown promising potential in treating a wide range of human diseases, including cancer, infectious diseases, neurodegenerative disorders, and cardiovascular diseases.

The drug discovery process using secondary metabolites typically involves several steps:

l. Collection and identification of the source organism: Researchers identify organisms known to produce secondary metabolites with potential medicinal properties. These organisms can be sourced from terrestrial environments (such as plants and fungi) or marine environments (such as corals, sponges, and algae).

2. Extraction and isolation: Secondary metabolites are extracted from the source organism using various techniques such as solvent extraction, distillation, or fermentation. The crude extract is then fractionated using chromatographic techniques to isolate individual compounds.

3. Biological activity screening: Isolated compounds are screened for their biological activity and potential therapeutic effects using in vitro and in vivo assays. These tests help determine the compounds' efficacy, safety, and mechanism of action.

4. Lead compound selection: Compounds that show promising biological activity are selected as lead compounds, which serve as starting points for drug development.

5. Structural modification and optimization: Lead compounds undergo structural modifications and optimization to improve their drug-like properties, such as potency, selectivity, and pharmacokinetic properties. This can involve chemical synthesis or semi-synthesis to generate analogs with improved activity or reduced toxicity.

6. Preclinical studies: Developed compounds undergo extensive preclinical studies to evaluate their safety, efficacy, and

pharmacokinetic properties. This includes testing in animal models to assess toxicity, dosing, and pharmacodynamics.

7. Clinical trials: Promising compounds that pass preclinical studies enter clinical trials, which involve testing in humans to evaluate their safety and efficacy. Clinical trials are typically divided into three phases and involve larger patient populations.

B. Food and flavor industry

Plant secondary metabolites are compounds produced by plants that are not directly involved in their growth, development, or reproduction. These metabolites often play a role in plant defense against pests, pathogens, and environmental stresses. They can also contribute to the aroma, flavor, and color of different plant parts. In the food and flavor industry, plant secondary metabolites are highly sought after as they can add unique taste, smell, and color to various food products. Here are some examples of plant secondary metabolites commonly used in the industry:

1. Phenolic compounds: These include flavonoids, tannins, and phenolic acids. They are widely distributed in fruits, vegetables, and beverages like tea and wine. Phenolic compounds provide antioxidant properties and contribute to the flavor, color, and astringency of foods.

2. Terpenes: Terpenes are responsible for the distinct aromas of many fruits, herbs, and

spices. For example, limonene gives citrus fruits their characteristic smell, while pinene is responsible for the aroma of pine needles. Terpenes are commonly used as flavoring agents and are also used in the production of essential oils.

3. Alkaloids: Alkaloids are a diverse group of compounds found in various plants, such as caffeine in coffee beans and nicotine in

tobacco leaves. They can contribute to the bitter taste of foods and are often used as stimulants or flavor enhancers.

- 4. Capsaicinoids: Capsaicinoids are the compounds responsible for the hot and spicy sensation in chili peppers. They are commonly used in the production of hot sauces, seasonings, and spicy snacks.
- 5. Anthocyanins: These are pigments responsible for the vibrant red, purple, and blue colors seen in fruits, vegetables, and flowers.

1. Flavor compounds derived from plant secondary metabolites

Flavor compounds derived from plant secondary metabolites are responsible for the characteristic taste and aroma of various fruits, vegetables, herbs, and spices. These compounds are synthesized by plants as part of their defense mechanisms against predators, pathogens, and environmental stressors. Here are some examples of flavor compounds derived from plant secondary metabolites:

1. Terpenes: These are a large class of compounds found in many plants, such as citrus fruits, pine trees, and herbs like basil and mint. Terpenes contribute to fruity, floral, and herbal flavors and aromas.

2. Phenolic compounds: This category includes compounds like flavonoids, phenolic acids, and tannins. They can be found in various plant sources, such as berries, grapes, tea, and coffee. Phenolics contribute to bitter, astringent, and earthy flavors.

3. Aldehydes: These are volatile compounds found in many fruits and vegetables, such as apples, tomatoes, and leafy greens. Examples include hexanal (green, grassy flavor) and furfural (nutty flavor)

4. Sulfur compounds: Found in garlic, onions, and cruciferous vegetables like broccoli and cabbage, sulfur compounds contribute to pungent and savory flavors.

5. Pyrazines: These compounds are responsible for the nutty, roasted, and caramel-like flavors found in coffee, chocolate, nuts, and toasted bread.

6. lactones: These are responsible for the fruity and coconutlike flavors found in peaches, pineapples, and whiskey.

.2. Natural food preservatives and additives from plant secondary metabolites

Plant secondary metabolites are compounds produced by plants for various purposes such as defense against pests, diseases, or environmental stress. Many of these compounds can also act as natural food preservatives and additives. Here are some examples:

1. Polyphenols: Polyphenols are a diverse group of compounds found in fruits, vegetables, and beverages like tea and wine. Examples include resveratrol in grapes and catechins in green tea.

2. Essential oils: Essential oils are volatile compounds extracted from plants. They have antimicrobial properties and can be used as natural preservatives in food. For example, oregano oil, thyme oil, and cinnamon oil have shown antimicrobial effects against various microorganisms.

3. Curcumin: Curcumin is the main active component of turmeric, known for its antioxidant and anti-inflammatory properties. It also exhibits antimicrobial effects and can be used as a natural food preservative, particularly against spoilage bacteria.

4. Allicin: Allicin is a sulfur compound found in garlic that exhibits antimicrobial activity against various bacteria, fungi, and

viruses. It is used as a natural preservative in some food products like pickles and dressings.

5. Citric acid: Citric acid is a natural organic acid found in citrus fruits. It is commonly used as a food additive for its acidic and preservative properties.

6. Terpenes: Terpenes are aromatic compounds found in many plants, including herbs and spices. Some terpenes, such as thymol in thyme and carvacrol in oregano, exhibit antimicrobial properties and can be used as natural food preservatives.

C. Agricultural applications of plant secondary metabolites

Plant secondary metabolites have a wide range of agricultural applications. Here are some examples:

l. Insecticides: Many secondary metabolites possess insecticidal properties and can act as natural insecticides to control pests in agriculture. For example, pyrethrins derived from chrysanthemum flowers are used as a natural insecticide.

2. Herbicides: Certain secondary metabolites can act as herbicides, inhibiting the growth of weeds. For instance, allelochemicals such as juglone from walnut trees can suppress weed growth.

3. Antifungal agents: Some secondary metabolites have antifungal properties, making them useful in controlling fungal diseases in crops. Examples include saponins from various plant sources and alkaloids like vincristine from the Madagascar periwinkle.

4. Antibiotics: Plant secondary metabolites have been used as natural antibiotics in agriculture to prevent the growth of pathogens in plants. For instance, plant-derived compounds like berberine have shown antimicrobial activity against bacteria and fungi.

5. Plant growth regulators: Certain secondary metabolites, such as gibberellins and auxins, act as plant growth regulators by influencing processes such as seed germination, flowering, and fruit development. These compounds can be used to enhance crop productivity and yield.

6. Allelopathy: Secondary metabolites released by plants can have allelopathic effects on nearby plants, influencing their growth and development. This can be utilized in crop management to suppress weed growth or enhance desirable plant traits.

7. Flavors and fragrances: Secondary metabolites are responsible for the characteristic flavors and fragrances of many plants. These compounds can be extracted and used in food, cosmetic, and fragrance industries.

I) Used as biopesticides, natural insect repellents and plant growth regulators

Plant secondary metabolites refer to a diverse group of organic compounds that are produced by plants for various purposes including defense against herbivores, attraction of pollinators, and inhibition of competing plants. These compounds have gained significant attention in recent years due to their potential applications as biopesticides, natural insect repellents, and plant growth regulators and enhancers.

Biopesticides are substances derived from natural sources, such as plants, animals, or microorganisms, and are used for controlling pests in agriculture. Plant secondary metabolites have shown promising biopesticidal properties, as they possess insecticidal, fungicidal, and herbicidal activities. For example, compounds such as pyrethrins derived from chrysanthemum flowers are used as natural insecticides to control various pests. Furthermore, some plant secondary metabolites have been found to exhibit plant growth regulatory and enhancing effects. These compounds can influence various physiological processes in plants, such as seed germination, root development, leaf expansion, and flowering. For instance, auxins, a class of plant hormones, can promote root and shoot growth, while gibberellins are known to stimulate stem elongation and flowering.

However, it is important to note that the efficacy of plant secondary metabolites can vary depending on factors such as the target pest or plant species, formulation, application method, and environmental conditions. Therefore, further research and development are needed to optimize their use and ensure their practicality in agriculture and pest management.

D. Plant secondary metabolites in Cosmetics and perfumery

Plant secondary metabolites are organic compounds produced by plants that serve various functions such as defense against herbivores, attraction of pollinators, and protection against pathogens. These metabolites have also found extensive use in cosmetics and perfumery due to their aromatic and therapeutic properties. Here are some examples of plant secondary metabolites used in the cosmetic and perfumery industries:

l. Essential Oils: Essential oils are volatile plant secondary metabolites extracted from various plant parts, such as flowers, leaves, and roots. They contain a complex mixture of aromatic compounds that give plants their characteristic scent. 2. Flavonoids: Flavonoids are a diverse class of plant secondary metabolites known for their antioxidant and anti-inflammatory properties.

3. Polyphenols: Polyphenols are a group of plant secondary metabolites with antioxidant, antimicrobial, and anti-aging properties. Examples of polyphenols used in cosmetics include resveratrol from grapes, catechins from green tea, and curcumin from turmeric.

4. Terpenoids: Terpenoids are a large class of plant secondary metabolites that contribute to the scent and flavor of plants. They are widely used in perfumery and skincare products for their aromatic properties.

5. Alkaloids: Alkaloids are nitrogen-containing plant secondary metabolites that have diverse biological activities. Some alkaloids, such as caffeine and theobromine derived from coffee and cacao, are used in cosmetic formulations for their stimulating and antioxidant properties.

i) Use of extract of plant secondary metabolites in cosmetics as natural fragrance and essential oils

Plant secondary metabolites are organic compounds synthesized by plants that go beyond their primary metabolic functions. These compounds have been found to have various biological activities, making them valuable for their potential use in cosmetics and fragrances.

Extraction of plant secondary metabolites:

1. Solvent extraction: Plant materials are soaked in a solvent (usually ethanol, methanol, or hexane) to extract the desired metabolites. The solvent is then evaporated to obtain the concentrated extract.

2. Steam distillation: This method is commonly used to extract essential oils. Steam is passed through the plant material, causing the oil glands to burst and release the essential oil. The oil is then separated from water and collected.

3. Cold pressing: For citrus fruits, the outer layer of the fruit is mechanically pressed to release the essential oils contained in the peel.

Use in cosmetics:

1. Anti-aging properties: Plant secondary metabolites like polyphenols and flavonoids have antioxidant properties that help protect the skin from free radicals and prevent premature aging.

2. Anti-inflammatory effects: Some metabolites, such as terpenoids and alkaloids, have anti-inflammatory properties that can reduce skin redness, irritation, and acne.

3. Skin brightening: Certain plant metabolites, like arbutin and kojic acid, can help lighten dark spots and improve overall skin tone.

4. Moisturizing properties: Many plant extracts, such as aloe vera and chamomile, have hydrating effects on the skin, keeping it moisturized and improving its elasticity.

Use in natural fragrances and essential oils:

1. Aromatherapy: Essential oils extracted from plants are commonly used in aromatherapy due to their calming, uplifting, and stress-relieving effects.

2. Scenting products: Plant secondary metabolites provide natural fragrances used in perfumes, lotions, soaps, and candles, offering a wide range of scents derived from different plants.

3. Therapeutic benefits: Some essential oils have specific therapeutic properties, like lavender oil for relaxation or tea tree oil for its antiseptic properties.

It's important to note that the extraction and use of plant secondary metabolites require careful consideration of sustainability, ethical sourcing, and safety regulations to ensure the protection of both plants and consumers.

IV. Future prospects and challenges related to plant secondary metabolites

Plant secondary metabolites have gained considerable attention in recent years due to their diverse biological activities and potential applications in various industries, including pharmaceuticals, agriculture, and cosmetics. However, there are several future prospects and challenges associated with these compounds.

1. Pharmaceutical Applications: Plant secondary metabolites have shown promising effects in treating various diseases, including cancer, cardiovascular disorders, and neurodegenerative diseases.

2. Agricultural Applications: Plant secondary metabolites can play a significant role in plant defense against pests and pathogens. The future prospects include the utilization of these compounds as natural alternatives to synthetic pesticides, leading to more sustainable and environmentally friendly agricultural practices.

3. Industrial Applications: Plant secondary metabolites have applications in the food and beverage industry, fragrance and flavor industry, and cosmetic industry.

4. Challenges in Discovery and Isolation: The discovery and isolation of plant secondary metabolites can be challenging due to their low abundance and complex chemical structures.

5. Biosynthesis and Metabolic Engineering: Understanding the biosynthetic pathways of plant secondary metabolites is crucial for their sustainable production.

6. Regulatory and Intellectual Property Issues: The commercialization of plant secondary metabolites involves navigating complex regulatory frameworks and protecting intellectual property rights.

7. Ecological Considerations: Plant secondary metabolites play essential ecological roles, such as allelopathy and plant-insect interactions.

A. Exploration of untapped plant sources for secondary metabolites

Exploration of untapped plant sources for secondary metabolites involves researching and identifying plant species that have not been extensively studied for their potential bioactive compounds. Secondary metabolites are organic compounds produced by plants that are not essential for their growth and development but have various biological activities.

To begin the exploration, you can start by reviewing existing literature on plants that are known to possess secondary metabolites with medicinal or other beneficial properties.

Identify plants that have not been extensively studied in this context.

Overall, the exploration of untapped plant sources for secondary metabolites requires a multidisciplinary approach, involving botany, ethnobotany, chemistry, and pharmacology. It is an essential step in discovering potential new drugs, natural products, and bioactive compounds that can have an impact on various fields including medicine, agriculture, and cosmetics.

B. Sustainable production and extraction methods of plant secondary metabolites

Here are some methods that can be utilized:

1. Organic farming: Implementing organic farming practices helps reduce the use of synthetic fertilizers, pesticides, and herbicides, which can contaminate the environment and affect the quality of plant secondary metabolites.

2. Agroforestry and intercropping: Growing plants in combination with trees or other crops can create a more sustainable and diverse agricultural system. Trees provide shade and wind protection, improve soil quality, and promote beneficial interactions between species, enhancing the production and quality of plant secondary metabolites.

3. Biotechnology and genetic engineering: Advances in biotechnology can play a crucial role in sustainable production of secondary metabolites.

4. Plant tissue culture and micropropagation: Plant tissue culture techniques, such as micropropagation, allow for the rapid and efficient multiplication of plant material under controlled conditions.

5. Sustainable extraction techniques: Green extraction methods, such as supercritical fluid extraction (SFE), microwave-assisted extraction (MAE), and ultrasound-assisted extraction

(UAE), have been developed to minimize solvent usage and reduce energy consumption during the extraction of secondary metabolites.

6. Vaporization of waste materials: Utilizing waste materials from agricultural and food processing industries can be an effective way to extract secondary metabolites sustainably.

C. Overcoming regulatory hurdles in commercializing plant secondary metabolites

Here are some steps to consider:

1. Understand the regulatory landscape: Start by familiarizing yourself with the regulatory framework governing the commercialization of plant secondary metabolites.

2. Conduct a thorough risk assessment: Identify potential regulatory risks that may impact your commercialization efforts.

3. Engage with regulatory authorities: Establish communication channels with relevant regulatory authorities to seek guidance and clarification on regulatory requirements.

4. Seek expert advice: Consulting with regulatory experts or hiring a regulatory consultant who specializes in plant secondary metabolites can provide valuable insights and guidance.

They can help navigate through complex regulations and suggest strategies to overcome regulatory hurdles.

5. Conduct pre-market assessments: Before commercializing plant secondary metabolites, it's important to conduct thorough pre-market assessments.

6. Develop robust documentation: Maintain detailed records of all research, testing, and development activities.

7. Collaborate with research institutions: Collaborating with academic or research institutions can provide access to scientific expertise, resources, and infrastructure for conducting the required studies and tests.

8. Stay updated on regulatory changes: Regulatory requirements may evolve over time. Stay informed about any changes or updates in the regulations that may impact your commercialization plans.

V. Conclusion

The study of plant secondary metabolites is crucial for understanding the diverse chemical compounds produced by plants and their potential applications in various fields. This chapter has examined the importance and classification of secondary metabolites, as well as their functions in plants and their ecological roles.One of the primary conclusions drawn from this chapter is that secondary metabolites play essential roles in plant defense mechanisms against herbivores, pathogens, and abiotic stresses. These compounds often possess toxic, deterrent, or allelopathic properties, which help plants survive and thrive in challenging environments. Furthermore, this chapter highlights the multifaceted nature of secondary metabolites, as they also have significance beyond plant defense. Many secondary metabolites serve as pigments, attractants for pollinators, or signaling molecules for plant growth and development. Some compounds even have potential medicinal or industrial applications. The discovery and isolation of secondary metabolites from plants have led to the development of various pharmaceutical drugs,

agricultural pesticides, and natural products for various industries. This chapter emphasizes the importance of continued research in this field to unlock the full potential of plant secondary metabolites. In conclusion, the study of plant secondary metabolites is a fascinating and rapidly evolving field. Understanding the diverse range of compounds produced by plants and their functions will not only enhance our knowledge of plant biology but also contribute to the development of sustainable agriculture, medicine, and other industries.

References:

- Liu, M., & Hansen, S. H. (2018). Understanding the structure-activity relationship of plant secondary metabolites for drug discovery. In Plant Bioactive Compounds for Pancreatic Cancer Prevention and Treatment (pp. 1-31). Springer.
- Ghorbani, A., & Esmaeilizadeh, M. (2017). Pharmacological properties of Salvia officinalis and its components. Journal of Traditional and Complementary Medicine, 7(4), 433-440
- Lila, M. A. (2019). Plant pigments and their manipulation: an approach to improving human health. Annual Review of Food Science and Technology, 10, 121-143
- 4. Khan, M. H., & Al-Ghamdi, A. A. (2018). Phytoconstituents of medicinal plants: a review of their antibacterial activity against multidrug-resistant bacteria. Journal of Chemical and Pharmaceutical Research, 10(8), 42-54.
- Shanmugam, S., Rajendran, R. B., & Rajesh, M. (2018). Pharmacological importance of plant secondary metabolites-a review. Journal of Pharmaceutical Sciences and Research, 10(6), 1415-1426.

Plant Secondary Metabolites: Functions and Applications

- 6. Serrano, R. (2019). Health benefits of plant secondary metabolites: beyond the usual suspects. In Food Quality and Shelf Life (pp. 317-334). Woodhead Publishing.
- Ahmed, S., Hasan, M. M., & Ahmed, S. M. (2017). Antioxidant activities of various solvent extracts, phydroxycinnamic acid and ursolic acid from Premna integrifolia Linn. leaves. Journal of Traditional and Complementary Medicine, 7(4), 523-529.
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G., & Kaur, H. (2011). Phytochemical Screening and Extraction: A Review. Internationale Pharmaceutica Sciencia, 1(1), 98-106.
- Boukhris M., et al. (2020). Plant secondary metabolites: a review of their potential anti-inflammatory and antioxidant medicinal applications. Current Pharmaceutical Design, 26(6), 679-690.
- Atanasov A.G., et al. (2015). Natural products in drug discovery: advances and opportunities. Nature Reviews Drug Discovery, 14(2), 111-129.
- 11. Kesari R., et al. (2020). Plant secondary metabolites as potential drug candidates against COVID-19. Phytotherapy Research, 34(9), 2292-2295.
- Yang H.B., et al. (2019). Plant-derived bioactive compounds as potential sources of multi-targeted agents for cancer treatment: Opportunities and challenges. International Journal of Biological Sciences, 15(7), 1371-1386.
- Rajurkar N.S., et al. (2018). Plant secondary metabolites: biosynthesis, classification, function and pharmacological properties. Journal of Pharmacognosy and Phytochemistry, 7(3), 1162-1171.

- Mishra B.B., et al. (2011). Plant natural products: an overview. Austin Journal of Pharmacology and Therapeutics, 1(1), 1-10.
- 15. Mohammed A., *et al.* (2020). Plant secondary metabolites: a comparison of anticancer properties in vitro and in vivo studies of isothiocyanates and their nitrogen analogues. Phytochemistry Reviews, 19(4), 785-801.
- Pilon A.C., *et al.* (2019). Plant secondary compounds in bioactive apples-impact of brief post-harvest storage. Food Research International, 125, 108633.
- 17. Bown D. (2019). Plant secondary metabolites: an ecological perspective. The New Phytologist, 224(3), 705-707.
- Wang Y., *et al.* (2020). Bioactive phytochemicals and their potential role in cancer prevention and treatment. Oxidative Medicine and Cellular Longevity, 2020, 1-18.
- 19. Wink, M. (2010). Plant secondary metabolites: occurrence, structure and role in the human diet. John Wiley & Sons.
- Pandey, A. K., & Palni, L. M. S. (1997). Antifungal activities of plant extracts against some plant pathogenic fungi. Indian Journal of Forestry, 20(4), 347-350.
- Harborne, J. B. (1988). Introduction to ecological biochemistry (Vol. 1). Academic Press.
- Kuc, J. (1982). Plant immunization and its application for disease control. In Advances in Botanical Research (Vol. 9, pp. 235-257). Academic Press.
- Simmonds, M. S. (2003). Flavonoid-insect interactions: recent advances in our knowledge. Phytochemistry, 64(1), 21-30.
- 24. Dixon, R. A., & Paiva, N. L. (1995). Stress-induced phenylpropanoid metabolism. Plant cell, 7(7), 1085-1097.

- Balandrin, M. F., Klocke, J. A., Wurtele, E. S., & Bollinger, W. H. (1985). Natural plant chemicals: sources of industrial and medicinal materials. Science, 228(4704), 1 154-1160. Grace, S. C. (1996). Phenolics as antioxidants. CRC Critical Reviews in Plant Sciences, 15(2), 103-128.
- Liu, M., Li, X. Q., Weber, C., Lee, C. Y, Brown, J., & Liu, R. H. (2002). Antioxidant and antiproliferative activities of raspberries. Journal of Agricultural and Food Chemistry, 50(10), 2926-2930.
- Quideau, S. , Deffieux, D. , Douat-Casassus, C. , & Pouységu, L. (2011). Plant polyphenols: chemical properties, biological activities, and synthesis. Angewandte Chemie International Edition, 50(3), 586-621.
- Dixon, R. A., & Steele, C. L. (1999). Flavonoids and isoflavonoids—a gold mine for metabolic engineering. Trends in Plant Science, 4(10), 394-400.
- Pichersky, E., & Gang, D. R. (2000). Genetics and biochemistry of secondary metabolites in plants: an evolutionary perspective. Trends in Plant Science, 5(10), 439-445.
- Wink, M. (2010). Introduction: biochemistry, physiology and ecological functions of plant secondary metabolites. In Annual Plant Reviews Volume 40: Biochemistry of Plant Secondary Metabolism (pp. 1-19). Wiley-Blackwell.
- Gang, D. R. (2005). Evolution of plant secondary metabolite gene clusters: gene duplication and alternative chemical pathways. Integrative and Comparative Biology, 45(5), 837-844.

- 32. Bohlmann, J., & Steele, C. L. (2012). Natural product chemistry: Harnessing the potential of plant secondary metabolites. In Comprehensive Natural Products II Chemistry and Biology (pp. 171-224). Elsevier.
- Harborne, J. B. (1999). Phytochemical methods: a guide to modern techniques of plant analysis. Springer Science & Business Media.
- Wink, M. (2013). Annual Plant Reviews Volume 40: Biochemistry of Plant Secondary Metabolism (Vol. 40). John Wiley & Sons.
- Simkaniaite, S., & Kuzina, V. (2020). Alkaloid diversity and evolution in the Zingiberales: a detailed review. Phytochemistry Reviews, 19(2), 401-438.
- Hamberger, B., & Bak, S. (2013). Plant P450s as versatile drivers for evolution of species-specific chemical diversity. Philosophical Transactions of the Royal Society B: Biological Sciences, 368(1612), 20120426.
- Facchini, P. J. (2001). Alkaloid biosynthesis in plants: biochemistry, cell biology, molecular regulation, and metabolic engineering applications. Annual Review of Plant Biology, 52(1), 29-66.

04

How Plants Reproduce

Tejaskumar H. Borkhatariya¹ and Chirag M. Godhani¹

¹Ph. D. (Scholars) Genetics and Plant Breeding, College of Agriculture, Junagadh Agricultural University, Junagadh

Author's email id: ahirtejas33@gmail.com

Abstract

Plant reproduction is a fascinating and essential aspect of the natural world, intimately connected to the sustenance of life on Earth. This chapter delves into the intricate mechanisms by which plants give rise to new generations, emphasizing their profound ecological significance and relevance to human civilization. From asexual reproduction strategies, including vegetative propagation and apomixis, to the complexities of sexual reproduction involving flowers, pollination, and genetic diversity, this chapter offers a comprehensive exploration of plant reproductive biology. Asexual reproduction allows plants to create genetically identical offspring, fostering rapid propagation and trait preservation. Various methods of vegetative propagation, such as rhizomes, tubers, bulbs, cuttings, and grafting, are discussed in detail. Apomixis, a unique form of asexual reproduction that enables seed production without fertilization, is also explored. Sexual reproduction, on the other hand, involves flower formation, pollination, fertilization, and seed production. Special attention is given to the structure and function of flowers in plants and the various types of pollination mechanisms, including self-pollination and cross-pollination. The concept of alternation of generations, exemplified in plants like ferns and mosses, elucidates the dynamic interplay between haploid gametophyte and diploid sporophyte phases in the plant life cycle. Additionally, specialized reproductive strategies, such as those employed by carnivorous plants and epiphytic species, showcase nature's adaptability to diverse environmental niches. Lastly, human influence on plant reproduction, through selective breeding and genetic engineering, underscores the pivotal role of plants in agriculture and biotechnology. This chapter serves as а comprehensive guide to understanding the intricate processes and ecological significance of plant reproduction, offering insights into the wondrous world of botanical diversity and the transformative impact of human intervention in the natural order.

Keywords: Sexual reproduction, Asexual reproduction, Types of pollinations, Apomixis, Alteration of generations

INTRODUCTION:

Plant reproduction is a captivating and essential aspect of the natural world, playing a pivotal role in sustaining life on our planet. As we delve into the intricate mechanisms by which plants give rise to new generations, we embark on a journey that unveils the profound importance of these processes for the environment, ecosystems, and human civilization. The world of plant reproduction is a testament to nature's ingenuity, showcasing an array of strategies, adaptations, and evolutionary wonders that have allowed plants to thrive for millions of years.

Through the millennia, plants have developed a wide spectrum of reproductive strategies, each finely tuned to their unique ecological niches. Whether through the propagation of clones, the orchestration of elaborate pollination dances, or the formation of resilient seeds, plants have perfected their methods to ensure the continuity of their species in the ever-changing tapestry of the natural world. Understanding how plants reproduce is not merely a scientific endeavour but a profound exploration of the intricate web of life that sustains our existence.

In this chapter, we embark on a journey to unravel the mysteries of plant reproduction, from the simplest modes of asexual propagation to the intricate choreography of sexual reproduction. We will delve into the mechanisms of pollination, the formation of seeds, the significance of genetic diversity, and the remarkable adaptations that have allowed plants to conquer an astonishing array of habitats. From the microcosms of flower ecosystems to the macrocosm of global ecosystems, the chapter on plant reproduction offers a window into the astounding complexities of the natural world that surrounds us.

TYPES OF REPRODUCTION IN PLANTS: ASEXUAL REPRODUCTION IN PLANTS:

Asexual reproduction in plants is a method of propagation that does not involve the fusion of male and female gametes (sperm and egg cells), as is the case in sexual reproduction. Instead, it enables plants to create genetically identical offspring, often referred to as clones, directly from a single-parent plant. This process typically occurs through various mechanisms, including the division of plant cells, specialized reproductive structures, or the growth of vegetative parts. Asexual reproduction offers several advantages, such as rapid reproduction, preservation of desirable traits, and the ability to colonize new habitats efficiently. In contrast to sexual reproduction, asexual reproduction does not require the involvement of pollinators, the production of seeds, or the investment of energy in creating complex reproductive structures.

Advantages and Disadvantages of Asexual Reproduction in Plants:

Asexual reproduction offers several advantages to plants. Firstly, it allows for the rapid generation of new individuals, as there is no need to wait for the development of flowers, pollination, and seed production. This can be particularly advantageous in environments with short growing seasons or in the case of plants facing sudden disturbances or damage to their reproductive structures. Secondly, asexual reproduction preserves the genetic traits of the parent plant, ensuring that the offspring inherit the exact genetic makeup of the parent. This is beneficial when a plant has desirable characteristics, such as disease resistance, exceptional fruit quality, or other traits that are advantageous for survival. However, it's important to note that this lack of genetic diversity can also be a disadvantage. Without genetic variation, populations of asexually reproducing plants may be more susceptible to diseases, pests, or environmental changes that a sexually reproducing population with diverse genes could better withstand.

Asexual reproduction in plants is a valuable mechanism that enables rapid propagation and the preservation of desirable traits. While it offers advantages in certain situations, it also comes with the potential drawback of reduced genetic diversity, which can impact a plant population's resilience and adaptability to changing conditions. Understanding when and how plants employ asexual reproduction is essential for comprehending their reproductive strategies and ecological roles in various environments.

Various Methods of Vegetative Propagation

Vegetative propagation is a vital aspect of asexual reproduction in plants. It involves the formation of new plants from non-reproductive parts of the parent plant, such as stems, leaves, roots, or specialized storage structures. This process ensures the continuity of plant species and is employed by both wild and cultivated plants. The following methods are key techniques in vegetative propagation:

1. Rhizomes and Runners

Rhizomes are horizontal, underground stems that grow horizontally beneath the soil surface. These specialized structures enable plants to propagate by producing new shoots and roots at various points along their length. Rhizomes play a significant role in the vegetative propagation of numerous plant species.

Runners, on the other hand, are similar to rhizomes but grow above the ground. They are also known as stolons and typically originate from the base of the parent plant. Runners produce new plantlets at nodes, allowing for the efficient spread of the species. Rhizomes and runners are essential for the survival and expansion of many plants, particularly in environments where disturbances like grazing or fires occur. They allow plants to rapidly colonize new areas, forming dense, interconnected populations (Harper, 1977).

2. Tubers

Tubers are enlarged, fleshy, underground storage structures that store energy and nutrients. They serve as a reservoir for the plant, enabling it to survive harsh conditions and propagate new plants when conditions become favourable.

Tubers are vital for the vegetative propagation of numerous crop plants, including potatoes, sweet potatoes, and yams. By storing energy and nutrients, tubers provide a head start for the new plant's growth, giving it an advantage in establishing itself (Jackson & Crock, 1999).

3. Bulbs, Corms and Rhizomes

Bulbs, corms and rhizomes are underground storage organs that enable plants to reproduce vegetatively. They store nutrients and provide a resource reserve for new shoots to emerge, often when conditions are favourable.
These storage organs are prevalent among ornamental plants such as tulips (bulbs), crocuses (corms), and irises (rhizomes). They allow gardeners and horticulturists to propagate and cultivate desirable varieties with ease.

4. Cuttings

Cuttings involve the removal of a portion of a plant, typically a stem, leaf, or root, and encouraging it to develop roots and grow into a new plant. This method is widely used in horticulture for the propagation of numerous ornamental, fruit, and vegetable plants.

Cuttings allow growers to reproduce plants with desirable traits, such as specific flower colours or disease resistance, while retaining the genetic characteristics of the parent plant. It is a cost-effective and efficient method of propagation (Hartmann *et al.*, 2010).

5. Grafting

Grafting is a specialized form of vegetative propagation in which two different plant varieties are joined together to grow as one. The upper part, known as the scion, bears desirable traits such as fruit quality or flower colour, while the lower part, the rootstock, provides the root system and physical support. Grafting is commonly used in fruit tree cultivation and allows growers to combine the best features of different varieties into a single plant. It is also employed to improve the overall health and vigour of a plant (Janick, 2002).

These various methods of vegetative propagation are invaluable tools for gardeners, horticulturists, and farmers, allowing them to propagate plants efficiently, maintain desirable traits, and contribute to the diversity and resilience of cultivated plant species.

Apomixis as a Form of Asexual Reproduction

Apomixis is a fascinating and relatively uncommon form of asexual reproduction in plants. Unlike sexual reproduction, which involves the fusion of male and female gametes, apomixis allows plants to produce seeds without fertilization. In apomixis, the offspring are genetically identical to the parent plant, essentially creating clones. This mode of reproduction is of significant interest to botanists and has practical implications in agriculture and plant breeding. In this section, we will explore apomixis, its mechanisms, advantages, and examples of apomictic plants.

Mechanisms of Apomixis

Apomixis encompasses several mechanisms that bypass traditional fertilization and seed formation processes. It can occur in

various plant species and has evolved independently multiple times. There are three main types of apomixis:

1. Diplosporous Apomixis:

In this form of apomixis, the embryo develops directly from a diploid (2n) cell within the ovule, typically a megaspore mother cell. This cell undergoes mitosis to produce an embryo, bypassing meiosis and the need for fertilization. Diplosporous apomixis is commonly found in a number of grasses, including several species of Poa and Pennisetum (Koltunow *et al.*, 1995).

2. Aposporous Apomixis:

Aposporous apomixis involves the formation of one or more additional cells within the ovule, known as aposporous initials. These cells develop into embryos directly, without fertilization. Aposporous apomixis is found in various plant families, including Asteraceae and Rosaceae. It is well-documented in species like dandelions (Taraxacum) and blackberries (Rubus) (Nogler, 1984).

3. Adventitious Embryony:

In some cases, apomixis can occur when embryos form from somatic cells outside the ovule, often in tissues such as the seed coat or the placenta. Adventitious embryony is found in several economically important plants, including citrus (Citrus spp.) and mango (Mangifera indica) (Krishna and Reddy, 2014).

Advantages and Significance of Apomixis

Apomixis offers several advantages to plants and has ecological and practical implications:

- 1. Conservation of Desirable Traits: Apomixis allows plants to perpetuate favourable genetic characteristics without the risk of genetic recombination. This is particularly important for preserving traits such as disease resistance, fruit quality, or specific flower characteristics.
- 2. Reproduction in Isolation: Some plants live in isolated or harsh environments where pollinators are scarce. Apomixis ensures that these plants can reproduce successfully even without the presence of compatible mates.
- **3. Seed Production without Pollinators:** Apomixis is especially valuable for plants that do not rely on external pollinators, such as wind or insects, to transfer pollen. This makes it a reliable method of seed production.

Apomixis is a remarkable form of asexual reproduction in plants that has evolved through various mechanisms in different plant families. It offers advantages such as the conservation of desirable traits and seed production without fertilization. Studying apomictic plants not only contributes to our understanding of plant reproductive biology but also has practical applications in agriculture and horticulture.

SEXUAL REPRODUCTION IN PLANTS

Sexual reproduction is a complex and critical process in the life cycle of plants, allowing them to create genetically diverse offspring by the fusion of male and female reproductive cells (gametes). In this section, we will explore the fundamental aspects of sexual reproduction in plants, including the mechanisms involved, the significance of genetic diversity, and the essential role of flowers in this process.

Sexual reproduction in plants involves a series of intricate steps that culminate in the formation of seeds, which serve as the next generation of plants. Here is a simplified overview of this process:

1. Flower Formation: Sexual reproduction in plants typically begins with the development of specialized reproductive structures called flowers. Flowers are remarkable structures that house the reproductive organs of the plant. They can vary greatly in size, shape, colour, and scent, depending on the species and its pollinators.

- 2. Pollination: The first crucial step in sexual reproduction is **pollination**, the transfer of pollen from the male reproductive organ (anther) to the female reproductive organ (stigma) within the same flower or between different flowers of the same or different plants. Pollination can occur through various agents, including wind, water, insects, birds, and bats, depending on the plant species.
- 3. Fertilization: After successful pollination, pollen grains adhere to the stigma and form a pollen tube that grows down to the ovule, located in the ovary. Inside the ovule, a female gamete (egg cell) and two male gametes (sperm cells) are produced. Fertilization occurs when one of the sperm cells fuses with the egg cell, forming a zygote (2n).
- 4. **Seed Formation:** The zygote develops into an embryo, and the ovule matures into a seed. The seed is composed of the embryo, stored nutrients (endosperm), and a protective seed coat. This structure protects and nourishes the embryo during germination and early growth.
- 5. Seed Dispersal: Once the seed is mature, it is dispersed from the parent plant to new locations. This can occur through various mechanisms, including wind, animals, water, or explosive mechanisms, depending on the plant species.

Significance of Genetic Diversity in Sexual Reproduction

Genetic diversity is a fundamental outcome of sexual reproduction in plants and plays a pivotal role in their adaptation, evolution, and ecological success. Here are the key reasons why genetic diversity is crucial in plant populations:

- Adaptation to Environmental Changes: Genetic diversity allows plant populations to possess a range of genetic traits. When environmental conditions change, some individuals within the population may have advantageous traits that enable them to survive and reproduce, ensuring the continued existence of the species.
- 2. Resistance to Diseases and Pests: Diverse plant populations are less vulnerable to diseases and pest infestations. If all individuals in a population are genetically identical (as in asexual reproduction), a single disease or pest could potentially wipe out the entire population.
- 3. Enhanced Reproductive Success: Genetic diversity can lead to improved reproductive success. In some cases, cross-pollination between genetically different individuals can result in hybrid vigour, where the offspring exhibit increased fitness and growth rates.
- 4. Long-Term Survival: Genetic diversity provides the raw material for evolution. Over long periods, genetic diversity

enables plant species to adapt to changing environmental conditions, ensuring their long-term survival.

Structure and Importance of Flowers in Plants

Structure of Flowers:

Flowers are the reproductive organs of angiosperms, the most diverse group of plants on Earth. They exhibit remarkable structural diversity, but they typically consist of four main parts:

- 1. Sepals: Sepals are the outermost whorl of modified leaves that enclose and protect the developing flower bud.
- **2. Petals:** Petals are often brightly coloured and serve to attract pollinators. They are the second whorl of modified leaves.
- **3. Stamens:** Stamens are the male reproductive organs of the flower, consisting of an anther (where pollen is produced) and a filament (the stalk that supports the anther).
- 4. Carpels (Pistils): Carpels are the female reproductive organs, typically consisting of an ovary (which contains ovules), a style (a slender tube connecting the ovary to the stigma), and a stigma (the receptive surface for pollen).

Sexual reproduction in plants is a complex process involving the formation of flowers, pollination, fertilization, and seed production. The significance of this process lies in the generation of genetic diversity, which enables plants to adapt to changing environments and ensures their long-term survival. Flowers, with their diverse structures and functions, are the central players in this essential reproductive process and play a crucial role in plant biology and ecosystem functioning.

Types of Pollination in Plants

Pollination is a critical process in the life cycle of plants, facilitating the transfer of pollen from the male reproductive structures to the female reproductive structures of flowers. This process is essential for the fertilization of seeds and the production of fruit. Pollination can occur through various mechanisms, leading to different types of pollination. In this article, we will explore the main types of pollination in plants, including self-pollination, crosspollination, and the various agents involved in pollination.

A. Self-Pollination

Self-pollination, also known as autogamy, occurs when pollen from the same flower or the same plant is transferred to the stigma of that same flower or plant. This type of pollination is often associated with flowers that have structural adaptations to promote self-fertilization. Some common mechanisms of self-pollination include:

- 1. Cleistogamy: Cleistogamous flowers remain closed and selfpollinate before they open. This strategy is common in many grass species and some legumes.
- 2. Herkogamy: In some flowers, structural arrangements prevent self-pollination. For example, the stigma may be positioned above the anthers, making it difficult for pollen to fall directly onto the stigma.

Advantages of Self-Pollination:

Self-pollination is a reliable method of reproduction when pollinators are scarce or unreliable. It ensures that a plant can produce seeds even when no other compatible mates are nearby.

Disadvantages of Self-Pollination:

Self-pollination can result in reduced genetic diversity within a population, as it does not involve the mixing of genetic material from different individuals. This can make a population more susceptible to diseases and environmental changes.

B. Cross-Pollination

Cross-pollination, or allogamy, occurs when pollen from the male reproductive structures of one flower is transferred to the stigma of a different flower on the same or a different plant. Crosspollination is a more common and diverse form of pollination in plants and often relies on external agents, such as animals or wind, to facilitate the transfer of pollen.

Types of Cross-Pollination on the bases of pollination agents:

- 1. Entomophily: Entomophily refers to pollination by insects. Many plants have evolved floral adaptations to attract specific pollinators, such as bees, butterflies, moths, and beetles. These adaptations include colourful petals, sweet nectar, and scents that appeal to the pollinators.
- 2. Anemophily: Anemophily is pollination by wind. Plants that rely on wind pollination often have small, inconspicuous flowers with abundant pollen. Examples of wind-pollinated plants include grasses, many trees (e.g., oak and pine), and some agricultural crops (e.g., corn and wheat).
- **3. Hydrophily:** Hydrophily is pollination by water. This is a rare form of pollination, primarily observed in aquatic plants. Pollen is released onto the water's surface and transported to the female flowers by water currents. Examples of hydrophilous plants include water lilies and pondweeds.
- 4. Zoophily: Zoophily refers to pollination by animals other than insects. While insects are the most common animal pollinators, other animals, such as birds (ornithophily) and bats (chiropterophily), can also serve as pollinators. Plants that rely

on animal pollinators often have adaptations like brightly coloured flowers and copious nectar.

Self-Incompatibility and Outcrossing

While sexual reproduction allows for genetic diversity, some plant species have mechanisms in place to prevent self-fertilization. Self-fertilization can lead to offspring with reduced genetic diversity and an increased risk of inheriting harmful mutations. To avoid this, some plants exhibit self-incompatibility, a mechanism that prevents fertilization by their own pollen.

Self-incompatibility can be controlled by specific genes that regulate the acceptance or rejection of pollen based on genetic similarity. Plants with self-incompatibility systems typically require pollen from a genetically different individual to achieve successful fertilization. This promotes outcrossing, where pollen from one plant is transferred to another plant, increasing genetic diversity within the population.

One well-studied example of self-incompatibility is found in the Brassicaceae family, which includes plants like broccoli, cabbage, and mustard. These plants possess a self-incompatibility system that promotes outcrossing and genetic diversity.

Advantages of Cross-Pollination:

Cross-pollination promotes genetic diversity within plant populations, as it involves the mixing of genetic material from different individuals. This diversity allows for adaptation to changing environmental conditions and increased resistance to diseases and pests.

Disadvantages of Cross-Pollination:

Cross-pollination often relies on external agents such as animals or wind for pollen transfer. If these agents are scarce or disrupted, it can impact plant reproduction. It can be energetically costly for plants, as they must produce extra pollen and nectar to attract pollinators and ensure successful fertilization.

Pollination is a vital process in the reproduction of plants, involving various mechanisms and agents. Self-pollination and cross-pollination represent two fundamental strategies, each with its advantages and disadvantages. The diversity of pollinators, including insects, birds, bats, and wind, showcases the intricate relationships between plants and their pollination partners, highlighting the incredible adaptations that have evolved over millions of years to ensure the survival and propagation of plant species.

ALTERNATION OF GENERATIONS

In the life cycle of most plants, there is a phenomenon known as alternation of generations. This alternation involves two distinct multicellular phases: the haploid (n) gametophyte generation and the diploid (2n) sporophyte generation. These phases alternate in the plant's life cycle, with one generation giving rise to the other.

Gametophyte Generation

The gametophyte generation is haploid and produces gametes, which are the reproductive cells involved in fertilization. In most plants, the gametophyte is a relatively small and inconspicuous structure that develops from a spore. Gametophytes produce male and female gametes through mitosis. Male gametophytes produce sperm cells, while female gametophytes produce egg cells.

Sporophyte Generation

The sporophyte generation is diploid and is the dominant, more recognizable phase in the life cycle of many plants. It grows from the fertilized zygote and produces spores through meiosis. Spores are typically enclosed in protective structures, such as sporangia, and are released into the environment. Spores germinate and develop into new gametophytes, thus completing the cycle.

Examples of Alternation of Generations

One well-known example of alternation of generations is found in ferns. Ferns have a conspicuous sporophyte phase that consists of fronds and is responsible for photosynthesis and reproduction. On the underside of fern fronds, sporangia produce spores through meiosis. When these spores are released and germinate, they give rise to small, heart-shaped gametophytes that produce eggs and sperm. Fertilization occurs on the gametophyte, resulting in the formation of a new sporophyte.

In mosses, the gametophyte generation is the dominant phase, forming the familiar green carpet-like structures seen in moist environments. The sporophyte, in contrast, is a short-lived structure that develops atop the gametophyte and produces spores.

SPECIALIZED REPRODUCTIVE STRATEGIES

Some plants have evolved specialized reproductive strategies to adapt to unique environmental conditions or challenges. These strategies can be complex and often involve interactions with other organisms. Two notable examples are:

Carnivorous Plants

Carnivorous plants have adapted to nutrient-poor environments by supplementing their nutrition through the capture and digestion of small animals, such as insects. While they still engage in typical sexual reproduction, their unique adaptations for obtaining nutrients set them apart from non-carnivorous plants. Common examples of carnivorous plants include the Venus flytrap (Dionaea muscipula), pitcher plants (Nepenthes spp.), and sundews (Drosera spp.). These plants have specialized structures, such as sticky leaves or pitfall traps, to capture and digest prey. By doing so, they obtain essential nutrients like nitrogen and phosphorus, which are often scarce in their habitats.

Epiphytic Plants

Epiphytic plants are adapted to grow on the surfaces of other plants, typically trees, without harming their host. These plants have evolved unique strategies for obtaining water and nutrients from the air and rain. While they reproduce sexually, they often employ a variety of methods to access support, light, and moisture.

Orchids are a well-known example of epiphytic plants. They often attach themselves to trees in tropical rainforests and rely on rainwater, air humidity, and nutrients trapped in decaying organic matter for survival. Their seeds are tiny and equipped with specialized structures that aid in wind dispersal.

HUMAN INFLUENCE ON PLANT REPRODUCTION

Humans have long played a significant role in plant reproduction, both unintentionally through cultivation and intentionally through selective breeding. Agriculture, in particular, has relied on human intervention to improve crop yields, enhance desirable traits, and adapt plants to various environmental conditions.

Selective breeding, also known as artificial selection, involves deliberately choosing and breeding plants with desirable characteristics, such as disease resistance, flavor, or yield. Over generations, this process has led to the development of countless cultivated plant varieties, from modern wheat and rice to heirloom tomatoes and sweet corn.

Furthermore, humans have harnessed biotechnology to manipulate plant reproduction. Genetic engineering techniques, like the introduction of specific genes into plants, have allowed for the creation of genetically modified organisms (GMOs) with novel traits. These traits can include resistance to pests, tolerance to herbicides, and improved nutritional content.

CONCLUSION

Plant reproduction is a complex and diverse process that encompasses a wide range of mechanisms and adaptations. From asexual reproduction through vegetative propagation to the intricate world of sexual reproduction involving pollination, fertilization, and seed formation, plants have evolved an impressive array of strategies to ensure their survival and adapt to changing environments. The alternation of generations, self-incompatibility systems, and specialized reproductive strategies like carnivory and epiphytism all contribute to the remarkable diversity of the plant kingdom. Humans have played a significant role in shaping plant reproduction through agriculture and biotechnology, further highlighting the importance of understanding these processes for both scientific knowledge and practical applications.

REFERENCES:

- Fenster, C. B., Armbruster, W. S., Wilson, P., Dudash, M. R., & Thomson, J. D. (2004). "Pollination Syndromes and Floral Specialization." Annual Review of Ecology, Evolution, and Systematics, 35, 375-403.
- Harper, J. L. (1977). Population biology of plants. Academic Press.
- Hartmann, H. T., Kester, D. E., Davies Jr, F. T., & Geneve, R. L. (2010). Hartmann & Kester's Plant Propagation: Principles and Practices. Prentice Hall.
- Jackson, M. T., & Crock, J. E. (1999). The nature of the potato crop. In Advances in Potato Chemistry and Technology (pp. 27-53). Elsevier.
- Janick, J. (Ed.). (2002). Horticultural Reviews (Vol. 28). John Wiley & Sons.
- Koltunow, A. M., Johnson, S. D., & Bicknell, R. A. (1995). "Sexual and apomictic development in Hieracium." Sexual Plant Reproduction, 8(6), 275-286.
- Kozloff, E. N. (1974). "Plants and Animals: A Comparative Survey." McGraw-Hill.
- Krishna, H., & Reddy, M. S. (2014). "Adventitious embryony in recalcitrant seeds of mango (Mangifera indica L.)." Acta Physiologiae Plantarum, 36(8), 2111-2118.

- Lovell, J. T., Mullen, J. L., Lowry, D. B., Awole, K., Richards, J. H., Sen, S., ... & Juenger, T. E. (2013). "Exploiting differential gene expression and epistasis to discover candidate genes for drought-associated QTLs in Arabidopsis thaliana." The Plant Cell, 25(4), 1386-1401.
- Nogler, G. A. (1984). "Gametophytic Apomixis." In Embryology of Angiosperms (pp. 475-518). Springer.
- Proctor, M., Yeo, P., & Lack, A. (1996). "The Natural History of Pollination." Timber Press.
- Stearn, W. T. (2002). "Botanical Latin: History, Grammar, Syntax, Terminology, and Vocabulary." Timber Press.

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Plant Hormones and Their Roles Sanjay Kumar¹, Mahima Choudhary¹ and Revendra Kushwaha²

¹Department of Plant Pathology, Rajmata Vijayaraje Scindia Krishi Vishwa Vidyalaya, Gwalior, Madhya Pradesh, India ²Department of Plant Pathology, Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur, Madhya Pradesh, India (Corresponding Author E-mail: sparihar734.sp@gmail.com)

Abstract

Plant hormones are chemical substances that mediate various physiological growth and development processes as a result of several intricate chemical and physical responses in plants. Auxins, gibberellins, cytokinins, ethylene, and abscisic acid make up this class of "classical phytohormones". The development of plants can be regulated, and plant growth regulators can also encourage flowering, fruit ripening, breaking fruit dormancy, stimulating cutting roots, and inhibiting shoot growth. They can also interrupt fruit dormancy and hasten or delay fruit ripening. However, because of their origin, structural diversity, and inherent influence on biological processes, understanding these biochemical entities and their characteristics is also a potential substitute for agrochemicals. The aim of this study is to provide an overview of the theoretical bases, underlying processes, and possible applications of plant hormones in a wide range of technical disciplines, including agriculture, medicine, biotechnology and the food industry, as well as the many known biosynthetic techniques.

Keywords: Auxins, Abscisic acid, Cytokinins, Ethylene, Gibberellin

Introduction

In higher plants, chemical signals produced by the plant itself are essential for the efficient regulation, coordination, and growth of metabolism, growth, and morphogenesis. Many of our present ideas on intercellular communication in plants came from studies done on animals in the 19th century, which is why several words and ideas are analogous. Hormones are the chemical messengers that mediate intercellular communication in animals (the Greek word for hormone is hormaein, which means to stimulate), whereas plant hormones, also known as phytohormones, are the chemical messengers that mediate growth and other physiological processes in plants. Utilized to identify plant-produced compound compounds, excluding xenobiotic substances, that can be transmitted and employed in particular physiological communication pathways. However, the terms phytohormone and plant growth regulator are also used synonymously (Overbeek, 1944; Rademacher, 2015).

Phytohormones are produced by plants, and they control a variety of metabolic processes, including fruit ripening, flowering, leaf abscission, shoot growth, and root growth. The most important hormones used in plant vegetative reproduction are typically regarded as auxins (AUXs), cytokinins (CTKs), gibberellins (GBRs), abscisic acid (ABA), and ethylene (ETH). This hormone, among others, encourages stem elongation, cell division, and root growth. But as research and technology advanced, new chemical compounds with traits of plant hormones were recently found (Hsan *et al.*, 2018; Ganchaeva *et al.*, 2019). In addition to natural phytohormones, it is now feasible to acquire commercially available

synthetic and biotechnological items referred to as plant growth regulators that are well known for their regulatory actions (Andresen and Cedergreen, 2010). Naphthalene acetic acid, or ANA, which is a root development stimulant (Ortola *et al.*, 1991; Sanower and Urbi, 2016), is one example of a synthetic, biological, or biosynthetic chemical or system that can have an impact on plants. and bacterial inoculate capable of strengthening the rhizosphere and helping to fix nitrogen (Tsavkelova *et al.*, 2006; Garces *et al.*, 2017; Lerma *et al.*, 2018).

Phytohormones

Phytohormones can be produced anywhere since, unlike animal hormones, plants lack specific organs or sites for their biosynthesis. The AUXs, GBRs, CTKs, ETHs, and ABSAc classes of phytohormones-often referred the classical to as phytohormones-are now understood to control plant development (Davies, 2010). Despite the fact that NO is regarded by many scientists as a crucial signaling molecule and a mediator in metabolic processes, it is still not acknowledged as a phytohormone merely because it is an inorganic substance (Arc et al., 2013). Nevertheless, as knowledge grows and researchers are able to reach consensus on fresh data and an understanding of the chemical processes taking place within plants, the number of phytohormones and messenger molecules will undoubtedly keep rising.

Most phytohormones have been identified in sieve components, and it is thought that here is where long distance transport takes place. Chemical messengers that travel through phloem and xylem often alert one organ to the status of another; therefore, the passage of these messengers across various plant organs is essential for this reason as well. For instance, roots make CTKs, which are subsequently delivered to the buds by the xylem, but buds produce AUXs, which can be immediately transmitted to the roots via the phloem. Another route of transport involves the diffusion of several hormones through cell membranes when they are in their protonated form, which is a weak acid (Vanneste and Friml, 2009).

Each of the plant hormones that have been found so far and their primary uses are described sequentially in order to provide more detail.

A. Auxins (AUXs)

Charles Darwin and his son examined the occurrence of phototropism in plants and published their findings in the book "The Power of Movement in Plants" in 1881 (Darwin, 1880). The first AUX was discovered in 1926 by Frits Went, who continued his research on growth stimuli and discovered a chemical substance that promoted development and accumulated on the side opposite the lighted zone (Thimann, 1940). Since it was initially discovered in human urine and has the potential to regulate plant growth, indole-3-acetic acid (IAA) was regarded as the most significant AUX (Kögl *et al.*, 1934). And intriguingly, auxins have also been derived from substances other than plants, such as indole-3-butyric acid (IBA), indole-3-propionic acid (IPrA), and naphthalene acetic acid (NAA) (Enders and Strader, 2015).

Enders and Strader (2015) say that the term "AUX" refers to all natural and man-made substances that promote cell division, the elongation of cell segments, the formation of new roots in leaves and stems, the stretching of stem sections, and other developmental processes that are similar to those caused by IAA. Therefore, a single molecular structure cannot be utilized to describe these compounds chemically, as the term "AUX" is now accessible as a physiological definition describing a group of substances in terms of functioning for plants (Ferro *et al.*, 2007). Notably, IAA and IBA correspond to a heterocyclic structure distinguished by an indole ring with one N- alkyl carboxylic acid group on position 3, whereas phenyl acetic acid (PAA) contains the N-alkyl carboxylic acid group on the phenyl and naphthyl rings, respectively.

Roles of auxin

a) Cell division and elongation

The main physiological effects of auxin in the shoots are cell division and cell elongation. It is essential for xylem and phloem tissue differentiation as well as stem secondary development.

b) Apical dominance

Many plants will continue to prevent the lateral buds below the terminal bud from growing as long as they are healthy and developing. The lateral buds quickly expand after the removal of the apical bud. Apical dominance is a situation in which the apical bud dominates the lateral buds and prevents the lateral buds from growing.

Auxin, a substance that is created at the terminal bud and transferred to the lateral buds along the stem, where it limits growth, was suggested by Skoog and Thimmann (1934) as a



potential contributor to apical dominance. An agar block was placed in its place after the apical bud was removed. The lateral buds grew swiftly as a result. However, the lateral buds remained stationary and did not develop when the apical bud was swapped out for an auxincontaining agar block.

c) Root initiation

When applied to the cut end of a young stem in lanolin paste, a soft fat derived from wool that works well as a solvent for auxin, auxin encourages more lateral branch roots. In contrast, a higher auxin concentration inhibits root growth but considerably increases the number of lateral roots. This finding, which has been extensively used to encourage root growth in commercially beneficial plants that are propagated by cuttings, is of significant practical significance.

d) Prevention of abscission

Auxins found in nature inhibit the growth of the abscission layer, which would otherwise cause fruits, flowers, and leaves to fall.

e) Parthenocarpy

Auxin can cause parthenocarpic fruit production, which is fruit formation without pollination and fertilization. Auxin is more concentrated in the ovaries of parthenocarpic fruits than it is in the ovaries of plants that only produce fruit after fertilization. After pollination and fertilization, auxin concentrations in the ovaries rise in the later cases.

f) Respiration

Auxin-induced growth and the stimulation of respiration by auxin are connected. By raising the amount of ADP produced by the rapid ATP utilization in the growing cells, auxin may indirectly speed up respiration.

g) Callus formation

Auxin may be involved in cell division in addition to cell elongation. In certain tissue cultures where the callus growth is generally normal, the callus growth only continues with the addition of auxin.

h) Eradication of weeds

When used in higher concentrations, several synthetic auxins, particularly 2, 4-D and 2, 4, 5-T, are effective at eliminating weeds.

i) Flowering and sex expression

Auxins often prevent flowering; however, they encourage uniform

flowering in pine apples and lettuce.

B. Gibberellins (GBRs)

Tetracyclic and diterpenoid chemicals are examples of endogenous plant growth regulators, or GBRs. According to Salazar-Cerezo *et al.* (2018), the majority of GBRs are biochemically inert and serve primarily as precursors and intermediates in the production or degradation of bioactive GBRs. GBRs, often referred to as gibberellic acids, were first identified in the 1930s in Japan in connection with a fungal disease known as "bakanae" or "foolish seedling" disease in rice harvests. Bakanae is distinguished by plants that grow excessively long and waste their seed production. The phytopathogenic fungus *Gibberella fujikuroi* secretes a chemical known as gibberellin A, which is the cause of this rice disease.

In the future, three unique gibberellins (GA1, GA2, and GA3) that have been isolated and studied from gibberellin A will be found (Gupta and Chakrabarty, 2013). GBR is structurally a collection of four-isoprene-unit diterpenoid molecules that make up the kaurene cycle. Only a small portion of the 136 GBRs found in bacteria, fungi, and higher plants thus far are bioactive. Genetic testing has additionally demonstrated that GA3 is the most active GBR and is routinely produced at the industrial level for horticultural and agronomic reasons (Cowling *et al.*, 1998; MacMillan, 2011). The most remarkable discovery of GBR activity, however, was the adoption of semi-dwarf mutant rice and wheat types, which have a gene that alters GBR biosynthesis without disrupting other processes of plant development and greatly boost grain productivity in the absence of GBR (Spielmeyer *et al.*, 2002).

Environmental elements like light, temperature, water, nutritional status and abiotic and biotic pressures all have an impact on how GBRs function and can either increase or decrease the amount of GBRs in an organism (Toyomasu *et al.*, 1998).

Roles of gibberellins

a. Seed germination

Some light-sensitive seeds, like those for lettuce and tobacco, do not germinate well in low light. When exposed to light or red light, these seeds start to germinate very quickly. By administering gibberellic acid to the seeds when they are in the dark, it is possible to get around this requirement for light.

b. Dormancy of buds

In locations with extreme cold, the autumn-formed buds go dormant until the following spring. Treatments with gibberellin can remove the buds' dormancy. After harvest, potatoes similarly go dormant, but the administration of gibberellin causes the roots to sprout vigorously.

c. Root growth

Gibberellins have negligible to no impact on root development. There may be some root development suppression at higher concentrations. Gibberellins in isolated cuttings significantly slow down the development of roots.

d. Elongation of internodes

The elongation of the internodes is the effect of gibberellins on plant growth that is most noticeable. Therefore, gibberellins override genetic dwarfism in numerous plants, including dwarf peas, dwarf maize, etc.

e. Bolting and flowering

Early growth in many herbaceous plants exhibits a rosette habit with short stems and few leaves. While the rosette habit persists during short days, bolting takes place during long days, during which the stem elongates quickly and transforms into primordia bearing polar axes. Gibberellins can also be used to trigger bolting in such plants, even on non-inductive short days.

Gibberellin administration produces bolting and flowering

in *Hyoscyamus niger* (long-day plant) under non-inductive short days. While the use of gibberellin typically leads to early blooming in long-day plants. Its impact on short-day plants can be highly diverse. It could either have no effect, stop, or start blossoming.

f. Parthenocarpy

Gibberellins encourage pollen grains to germinate, and they can also promote fruit growth and the development of parthenocarpic fruits when applied to plants. Many times, such as with pome and stone fruits, where auxins have been unable to promote parthenocarpy, gibberellins have succeeded. Gibberellin treatments are used on a large scale to produce seedless, fleshy tomatoes and enormous seedless grapes.

g. Synthesis of the enzyme amylase

Gibberellins are needed for the amylase enzyme to form in the aleurone layer of the endosperm of cereal grains when they start to grow. Starch is digested by this enzyme into simple sugars, which are then given to developing embryos as a source of energy.

C. Cytokinins (CTKs)

Skoog and his associates identified a new phytohormone called CTKs in the 1950s. The basic characteristics of the leaves are affected by hormones that encourage growth, cell division, and reproduction, as well as by hormones that activate the development of pigments in the cells. Additionally, cell growth slows natural senescence, enhances the ability of organs and tissues to absorb various nutrients, and improves photosynthesis by encouraging chloroplast differentiation and stomal opening. The growth of the female gametophyte, the development of the vascular system, the formation of nitrogen-fixing nodules in plant roots, and the intake of essential soil components (such as N, P, S, and Fe) are all regulated by CTKs. According to Durmus and Kadiolu (2005), these phytohormones are essential for plants' defense against biotic

stressors, including bacteria, fungus, and viruses, as well as abiotic stressors like cold, drought, heat, and soil salinity. The hormonal effect of CTKs is influenced by their trans-Zeatin structure. It is the most well-known natural CTK due to its high affinity for all examined CTK receptor isoforms, or "protein variants."

The cytokinin activity of cis-Zeatin is substantially lower than that of the trans-isomer. Natural CTKs usually only have an effect on plants at nanomolar concentrations (1–50 pmol g–1), and the effects may even be negative at higher concentrations (Oshchepkov *et al.*, 2020). Furthermore, CTKs are not just limited to higher plants; they can also be present in lower-order plants, including bacteria, algae, parasitic insects, and nematodes, as well as in some genuine environmental components like soil and water (Stirk & van Staden, 2010).

Roles of Cytokinins

a. Cell division

The induction of cell division is kinetin's most important biological effect on plants, especially in tissues like the tobacco pith callus, carrot root tissue, soybean cotyledon, pea callus, etc.

b. Cell enlargement

Auxins and gibberellins, as well as kinetin, may promote cell proliferation. The cortical cells of tobacco roots, tobacco pith culture, pumpkin cotyledons, Phaseolus vulgaris leaves, and other samples have all demonstrated noticeable cell expansion.

c. Concentration of apical dominance

Cytokinin is a substance that is given topically to counteract the effects of apical dominance by encouraging the development of lateral buds.

d. Dormancy of seeds

Kinetin treatment can also break the dormancy of some light-sensitive seeds, such as lettuce and tobacco. This is similar to how gibberellins accomplish it.

e. Delay of senescence (Richmand-Lang effect)

Chlorophyll depletion and fast protein degradation are typically associated with the senescence of leaves. By enhancing RNA synthesis first, then protein synthesis, kinetin therapy can delay senescence by several days.

Working with detached *Xanthium* leaves, Richmand and Lang (1957) discovered that kinetin might delay senescence for a number of days.

f. Flower induction

It is possible to successfully use cytokinins to encourage flowering in short-day plants.

g. Morphogenesis

It has been shown that high auxin and low kinetin produced roots instead of the shoot buds that could be formed by high kinetin and low auxin.

h. Accumulation and translocation of solutes

With the aid of cytokinin, plants actively accumulate solutes and aid in solute translocation in the phloem.

i. Protein synthesis

Osborne (1962) showed that the translocation caused by kinetin therapy boosted the rate of protein synthesis.

j. Other effects

Some plants have cytokinins that give them tolerance to cold, heat, and pathogens. They substitute for the photoperiodic requirements to aid in flowering as well. In some situations, they promote the production of a number of photosynthesis-related enzymes.

k. Commercial applications

Fruits' shelf lives have been extended with cytokinins, which have also been used to speed up root induction, create effective root systems, and increase the yield and oil content of oil seeds like ground nuts.

D. Ethylene (ETH)

ETH, a low molecular weight unsaturated hydrocarbon (CH2=CH2, 28 g/mol) that can control plant development and senescence, is crucial to the various processes taking place when agricultural goods are ripening. All plants in nature contain ETH; however, the amount varies significantly throughout the course of a plant's life cycle depending on the climate and the stage of development at which the plant is at. One of the earliest substances to be recognized as influencing plant development and regulating natural growth was ETH. The first person to explain the pea seedlings' tripartite growth reaction by employing ETH was a Russian student named Neljubov in the early 19th century. These three reactions included a reduction in height, an expansion of the hypocotyl hook, and an inhibition of root growth. ETH was the first phytohormone to be chemically identified, but it took until the introduction of gas chromatography in the second half of the 20th century for ETH to be recognised as an endogenous hormone that controls plant growth and development. Due to the fact that ETH is a gas, it can be difficult to locate it in manufacturing facilities (Grierson, 2012; Jiang and Asami, 2018).

One of ETS's distinguishing qualities is its capacity to interact molecularly with other phytohormones. For instance, ETH interacts with AUXs and ABSAc to encourage the growth of leaves. Since S-adenosyl-methionine (SAM), an immediate substrate for ETH biosynthesis, is maintained by CTKs, they also play a significant role in leaf initiation (Iqbal *et al.*, 2017). This relationship is defined as the influence of AUX and ABSAc on ETH production.

Roles of Ethylene a. Fruit Ripening:

Ethylene is also referred to as the "fruit ripening hormone," since one of its most noticeable effects is the ripening of fruits. When ethylene is applied exogenously to different kinds of fruits, the reactions vary. In climacteric fruits like apples, bananas, tomatoes, etc., mature fruits exposed to ethylene cause climacteric respiration (a noticeable increase in respiration during the commencement of ripening), which is followed by further ethylene production, hastening the ripening process.

Autocatalytic ethylene production occurs as fruits ripen. However, ethylene treatment does not result in climacteric respiration or extra ethylene generation in non-climacteric fruits like grapes and citrus fruits, and the ripening process is unaffected.

For all varieties of fruits to ripen, a minimum threshold level of endogenous ethylene is necessary. This was shown in tests with transgenic tomatoes, where ACC synthase or ACC oxidase antisense completely stopped ethylene from being made.

These transgenic tomatoes' ripening process, which could only be restored by exogenous ethylene treatment, was thoroughly examined. The never-ripe mutant tomato doesn't ripen because of a change in the ethylene receptor. This change stops the receptor from binding to ethylene, which stops ethylene from having its hormonal effect.

b. Plumular Hook Formation:

The plumular tip, or shoot apex, of etiolated dicot seedlings is typically twisted in the shape of a hook. This hook-shaped structure helps seedlings penetrate the soil while preventing damage to the delicate apical growth point.

The establishment and maintenance of the plumular hook in an etiolated (darkly developing) seedling are caused by the production of ethylene in that region, which causes asymmetric or uneven growth on the two sides of the plumular tip. The outside side of the plumular tip lengthens from ethylene more quickly than the inner side. When the seedling is exposed to white light, its ethylene production decreases, and the inner side of the hook also elongates swiftly, bringing the growth of the two sides into balance and opening.

The red light is effective at releasing a plumular hook. This impact can be undone by exposing the seedling to far-red light. The red/far-red reversibility provides a clue as to the function of the pigment phytochrome in it.

The plumular hook does not open when etiolated seedlings are exposed to light while being treated with ethylene. On the other side, the plumular hook opens if seedlings are grown in the dark with an ethylene absorbent like KMnO4.

In etiolated dicot seedlings, asymmetric growth on opposite sides of the plumular tip that results in the production of hooks and their maintenance is thought to be caused by an ethylene-dependent auxin gradient, much like the one that forms during phototropic curvature.

c. Triple Response:

In etiolated seedlings, such as those of the pea, ethylene causes a "triple response" that comprises

(i) An inhibition of stem extension

(ii) stimulation of radial swelling of stems, and

(iii) An induction of stems that grow horizontally with respect to gravity (i.e., diageotropism).

(iv) The triple response characteristics of etiolated seedlings were related to the early discoveries of ethylene as a natural plant growth hormone.

d. Formation of Adventitious Roots and Root Hairs:

In plants, ethylene stimulates the development of adventitious roots from a variety of plant components, including the leaf, stem, peduncle, and even other roots. Ethylene treatment encourages the development of root hairs in many plants, including Arabidopsis.

e. Inhibition of Root Growth:

Dicotyledonous plants' roots have been shown to develop less linearly when exposed to ethylene.

f. Leaf Epinasty:

When the petiole's top (adaxial) side grows more swiftly than its lower (abaxial) side, the leaf curls downward. This is referred to as an epinasty. Ethylene causes the production of leaf epinasty in tomatoes and other dicot plants, including the potato, pea, and sunflower. Young leaves are more delicate than older leaves. Monocots, however, do not display this reaction.

Through increased ethylene synthesis, stress factors such as salt stress, water logging, and pathogen infection, as well as auxin concentrations that are higher, also cause leaf epinasty. Water logging causes anaerobic conditions surrounding the roots of tomatoes and other plants, which leads to an accumulation of ACC (the primary precursor to the synthesis of ethylene) in the roots. When oxygen is present, ACC is subsequently translocated to shoots along with the transpiration stream, where it is converted to ethylene and causes leaf epinasty.

g. Flowering:

It is well known that ethylene prevents plant blossoming. However, it promotes flowering in mango as well as in pineapple and its friends (Family Bromeliaceae). Commercially, ethylene is used to time the flowering and fruit set of pineapples. With the help of ethylene, *Plumbago indica* (a short-day plant) can be forced to flower even during long, non-inductive days.

h. Sex Expression:

In monoecious species (species with separate male and female flowers on the same plant), especially in a number of cucurbits, including cucumber, pumpkin, squash, and melon, ethylene significantly promotes the development of female flowers. This significantly reduces the number of male flowers.

i. Senescence:

Ethylene accelerates the senescence of plant leaves and flowers. It is now widely accepted that a balance between these two phytohormones regulates senescence because endogenous ethylene concentrations rise and cytokinin concentrations fall during senescence.

When freshly cut carnations are kept in water in a conical flask, the colour of their petals fades, and they wither (go through senescence) in a matter of days. However, clipped carnations can stay fresh for many weeks if they are kept in a conical flask filled with a silver thiosulfate solution. This is so that ethylene activity can be effectively inhibited by silver thiosulfate. Studies using transgenic plants have now confirmed the role of ethylene in enhancing senescence. The regulation of senescence has also been linked to ABA (abscisic acid). Its concentration also grows throughout the procedure.

j. Abscission of Leaves:

In plants, ethylene stimulates leaf abscission. Younger leaves are less sensitive than older leaves. The wild-type birch tree (*Betula pendula*) rapidly loses its leaves within a few days after being fumigated with 50 ppm ethylene. On the other hand, ethylene has no effect on a transgenic birch tree with a mutant form of the Arabidopsis ethylene receptor ETR1-1. This means that the tree does not lose its leaves when treated with ethylene.

The synthesis of ethylene, which promotes leaf abscission, is regulated by the relative concentration of auxin on the two sides of the abscission layer. Auxin concentration in the laminar zone drops at the time of abscission, but ethylene production rises at the same time. This also makes cells in the abscission zone more susceptible to the effects of ethylene since they are now producing enzymes that break down cell walls, like cellulose and pectinases.

These enzymes' activity causes cell wall thinning and cell separation, which ultimately cause leaves to abscise. ABA has also been linked to leaf abscission, in addition to auxin and ethylene. When the leaf abscises, its conc. rises.

Breaking the Dormancy of Seeds and Buds:

In barley and other cereals, ethylene is known to cause seeds to emerge from dormancy. Treatment with ethylene also helps strawberries, apples, and other plants overcome seed dormancy. Compared to dormant variants, seeds from non-dormant kinds release more ethylene.

Ethylene speeds up seed germination in many plants, and researchers have discovered a strong link between ethylene production and peanut (Arachis hypogaea) seed germination. The dormancy of buds can also be disrupted by ethylene treatment in various plants. In order for potato tubers to sprout their latent buds, ethylene may occasionally be applied.

E. Abscisic acid (ABSAc)

In addition to controlling flower induction, cell division, elongation, and dormancy, the signaling molecule ABSAc also controls how plants react to environmental stresses like drought, salinity, cold, pathogen attack, and UV radiation. It was initially termed "abscissine II" and was isolated from young cotton fruit in the early 1960s during studies connected to the identification of chemicals responsible for fruit abscission (Ohkuma *et al.*, 1963).
ABSAc is a terpenoidal molecule with a 15-carbon structure (C15). Additionally, the trans or cis isomers of ABSAc depend on the orientation of the carboxyl group at carbon 2. Additionally, a symmetric carbon atom in the ring's 1' position determines whether the compound is S (+) or R (-).

The commercially available form is an almost equal blend of the S and R forms, whereas the natural form is (S)-cis-ABSAc. The physiologically inactive forms of (S)-2-trans-ABSAc and (R)-2trans-ABSAc can be created by the isomerization of (S)-cis-ABSAc and (R)-cis-ABSAc by light, respectively. Both the (S)-cis- and (R)cis-forms of ABSAc are highly active in quick ABA responses like stomatal closure, whereas the (S)-cis-ABSAc form is more active in delayed ABSAc responses like seed maturation. (Xiong and Zhu, 2003). According to Jiang and Asami (2018), All plants, including green algae and terrestrial plants, certain phytopathogenic bacteria, metazoans from marine sponges, and humans all contain ABSAc.

Roles of Abscisic Acid

a. Stomatal regulation:

Abscisic acid has several effects, but its regulation of stomatal closure in plants under drought or water stress is the most significant and well-known. It prevents guard cells from absorbing K+ and encourages malic acid leakage. Because of the reduction in osmotically active solutes, the guard cells sag and the stomata close.

b. Seed and bud dormancy:

In a number of plants, ABA inhibits development and causes bud dormancy. According to research on tomatoes, it prevents the lateral expansion of buds. Generally speaking, it has an inhibitory chemical effect that interferes with bud growth as well as seed and bud dormancy. It prevents growth in temperate-region plant species, which contributes to bud and seed dormancy; nevertheless, growth restarts once it disappears from seeds or buds. As gibberellin level rise and ABA levels fall, other plants start to grow. Without ABA, seeds and buds would begin to grow during the mild winter months and would perish when the temperature fell below freezing.

Due to the slow rate at which ABA exits the tissues and the time it takes for other plant hormones to act to counteract it, physiological processes that provide some protection against premature development are delayed. It accumulates inside seeds as fruits grow and hinders germination outside of the fruit or before winter.

c. Seed development and germination:

It has been noted that ABA either accumulates entirely from scratch or is transported from leaves into the embryos of growing seeds. It prevents vivipary and the development of germinationrelated enzymes in the embryo. Most non-dormant seeds cannot germinate when exogenous ABA is supplied. As soon as it is removed by washing the seeds, germination can occur, which may be caused by enzymes that are involved in the germination process being inhibited or sprouting seeds being unable to absorb water, among other things.

d. Senescence and Abscission:

Many researchers hypothesized that ABA functions as an endogenous component in the senescence and abscission of leaves and other plant parts. Herbaceous plants and deciduous trees are just two examples of the species in which ABA can produce primary yellowing in the leaf tissues. The production of ABA increases in the senescing leave as soon as the photosynthetic activity of the leaves falls below the compensatory point.

e. Geotropism:

There is sufficient data to prove that ABA regulates roots' geotropic responses. The root tips of maize have been found to contain significant quantities of ABA. It would appear that light and

gravity are needed for the ABA to accumulate at the tip. It is made in the root cap, moves basipetally, and, by acting as an inhibitor, promotes a favorable geotropic response.

f. Flowering:

By negating the impact of gibberellins on flowering in longday plants, ABA functions as a flowering inhibitor. Conversely, ABA causes short-day plants to blossom.

g. Cambium activity:

As winter approaches, abscisic acid causes the vascular cambium to stop mitosis.

h. Role in water stress:

In times of drought and water stress, abscisic acid is crucial for plants. It is known as a "stress hormone" because it makes plants more resilient to many forms of stress. It has been noted that plants under such stressors have an increase in ABA concentration in their leaves. ABA contributes to stomata closure in plants experiencing water stress. After plants experience water stress and their roots become dehydrated, a signal travels up to the leaves, where it triggers the creation of ABA precursors. These precursors then travel to the roots. The roots release ABA, which is then carried to the leaves through the vascular system. ABA changes the way potassium and sodium are absorbed by the guard cells, which causes them to lose their elasticity.

i. Counteracts the effects of other hormones:

The stimulatory and inhibitory actions of other hormones are offset by abscisic acid.

i) IAA stimulates cell development, which ABA inhibits;

ii) ABA prevents seeds treated with gibberellins from producing amylase,

iii) ABA promotes chlorosis, which cytokinin inhibits.

This might be because ABA is a Ca++ antagonist, and because it interferes with Ca++ metabolism, it inhibits the stimulatory effects of cytokinin and IAA.

j. Some other effects are:

Prevents cell elongation and cell division; Rose's parthenocarpic development is induced, increasing plants' ability to withstand cold; During the germination of cereal grains, gibberellin is inhibited, which prevents the production of amylase. prevents the ripening of fruit, etc.

References

- Andresen M., Cedergreen N. (2010). Plant growth is stimulated by tea-seed extract: A new natural growth regulator? *HortScience* 45:1848-1853.
- Arc E., Sechet J., Corbineau F., Rajjou L., Marion-Poll L. (2013). ABA Crosstalk with Ethylene and Nitric Oxide in Seed Dormancy and Germination. *Frontiers in Plant Science* 4: 1-19.
- Cowling R., Kamiya Y., Seto H., Harberd P. (1998). Gibberellin Dose-Response Regulation of GA4 Gene Transcript Levels in Arabidopsis. *Plant Physiology*. 117: 1195– 1203.
- Darwin, C. (1880). General considerations on the movements and growth of seedling plants. In the Power of Movement in Plants (Cambridge Library Collection - Darwin, Evolution and Genetics. 67-128.
- Davies, P. J. (2010). Their nature, occurrence, and functions. Plant Hormones. Springer. 1–15.
- Durmuş, N., Kadioğlu A. (2005). Reduction of Paraquat Toxicity in Maize Leaves by Benzyladenine. Acta Biologica Hungarica. 56: 97–107.
- Enders T., Strader L. (2015). Auxin Activity: Past, Present, and Future. *American Journal of Botany* 102:180–96.
- Ferro N., Bultinck P., Gallegos A., Jacobsen H., Carbo-Dorca R. (2007). Unrevealed structural requirements for auxin-

like molecules by theoretical and experimental evidences. *Phytochemistry*. 68: 237–250.

- Gancheva S., Malovichko Y., Poliushkevich L., Dodueva I., Lutova L. (2019). Plant Peptide Hormones. *Russian Journal of Plant Physiology*. 66: 171-189.
- Garcés V., Palencia M., Combatt, E. (2017). Development of bacterial inoculums based on biodegradable hydrogels for agricultural applications. *Journal of Science with Technological Applications*. 2: 13-23.
- Grierson D. (2012). 100 Years of Ethylene A Personal View. In the plant hormone ethylene, Wiley-Blackwell. 44: 1–17.
- Gupta R., Chakrabarty S. (2013). Gibberellic Acid in Plant: Still a Mystery Unresolved. *Plant Signaling and Behavior*. 8: 255041-255045.
- Hasan M., Razvi S., Kuerban A., Balamash K., Al-Bishri W. (2018). Strigolactones-A Novel Class of Phytohormones as Anti-Cancer Agents. *Journal of Pesticide Science*. Pesticide Science Society of Japan. 43: 168-172.
- Iqbal N., Khan N., Ferrante A., Trivellini A., Francini A. (2017). Ethylene role in plant growth, development and senescence: interaction with other phytohormones. *Frontiers in plant science*. 8: 475-503.
- Jiang K., Asami T. (2018). Chemical regulators of plant hormones and their applications in basic research and agriculture. Bioscience, Biotechnology and Biochemistry. Japan Society for Bioscience Biotechnology and Agrochemistry. 82: 1265-1300.
- Kögl F., Haagen-Smit A., Erxleben H. (1934) Über Ein Neues Auxin ('Hetero-Auxin') Aus Harn. 11. Mitteilung Über Pflanzliche Wachstumsstoffe. Hoppe-Seyler's Zeitschrift Fur Physiologische Chemie. 228: 90–103.
- Lerma T., Combatt E., Palencia M. (2018). Soil-mimicking hybrid composites based on clay, polymers and nitrogen-fixing bacteria for the development of remediation systems of

degraded soil. *Journal of Science with Technological Applications*. 4 :17-27.

- MacMillan J. (2001). Occurrence of gibberellins in vascular plants, fungi, and bacteria. *Journal of Plant Growth Regulation.* 20 :387–442.
- Ohkuma K., Lyon J, Addicott F., Smith O. (1963). Abscisin II, an abscission-accelerating substance from young cotton fruit. *Science*. 142 :1592–1593.
- Ortolá, A., Monerri C., Guardiola JL. (1991). The use of naphthalene acetic acid as a fruit growth enhancer in Satsuma mandarin: a comparison with the fruit thinning effect. *Scientia Horticulturae*. 47 :15-25.
- Osbrone, D.J. (1962). Effect of kinetin on protein and nucleic acid metabolism in *Xanthium* leaves during senescence. *Plant Physiol.* 37:595-602.
- Oshchepkov M., Kalistratova A., Savelieva E., Romanov G., Bystrova N. (2020). Natural and synthetic cytokinins and their applications in biotechnology, agrochemistry and medicine. *Russian Chemical Reviews*. 89:787–810.
- Overbeek J. (1944). Growth-Regulating substances in plants. *Annual Review of Biochemistry*. 13 :631–66.
- Rademacher W. (2015). Plant growth regulators: backgrounds and uses in plant production. *Journal of Plant Growth Regulation*. 34 :845–72.
- Richmond. A.E. and Lang, A. (1957). Effect of kinetin on protein and survival of detached *Xanthium* leaves. *Science*. 125:650.
- Salazar-Cerezo S., Martínez-Montiel N., García-Sánchez J., Pérez-Terrón R., Martínez-Contreras R. (2018). Gibberellin biosynthesis and metabolism: a convergent route for plants, fungi and bacteria. *Microbiological Research*. 208 :85-98.
- Sanower M., Urbi Z. (2016). Effect of Naphthalene Acetic Acid on the adventitious rooting in shoot cuttings of Andrographis Paniculata (burm.f.) wall. ex nees: an

important therapeutical herb. International Journal of Agronomy. 1-6.

- Skoog, F., and Thimann, K.V. (1934). Further experiments on the inhibition of the development of lateral buds by growth hormone. *Proc. Natl. Acad. Sci.* 20:480.
- Spielmeyer W., Ellis M., Chandler P. (2002). Semidwarf (Sd-1), 'Green revolution' rice, contains a defective Gibberellin 20-Oxidase gene. Proceedings of the National Academy of Sciences of the United States of America. 99 :9043– 9048.
- Stirk W., van Staden J. (2010). Flow of Cytokinins through the environment. plant growth regulation. *Springer*. 62 :101-116.
- Thimann K. (1940). Growth hormones in plants. *Journal of the Franklin Institute*. 229: 337–346.
- Toyomasu T., Kawaide H., Mitsuhashi W., Inoue Y., Kamiya Y. (1998). Phytochrome regulates gibberellin biosynthesis during germination of photoblastic lettuce seeds. *Plant Physiology*. 118: 1517–23.
- Tsavkelova E., Klimova S., Cherdyntseva, T., Netrusov A. (2006). Microbial producers of plant growth stimulators and their practical use: A review. *Applied Biochemistry and Microbiology*. 42:117-126.
- Vanneste S., Friml J. (2009). Auxin: a trigger for change in plant development. *Cell*. 136:1005-1016.
- Xiong L., Zhu J. (2003). Regulation of Abscisic Acid biosynthesis. Plant Physiology. *American Society of Plant Biologists*. 133:29-36.

Plant Stress Physiology: Abiotic and Biotic Stress Responses ¹Vijay Kumar Choudhary and ²Priyanka Choudhary

¹Department of Genetics and Plant Breeding, DDUGU, Gorakhpur ²Department of Soil Science, Dr RPCAU, Pusa, Bihar

Introduction

6

Plant stress physiology is a branch of biology that focuses on understanding the responses of plants to various stressors in their environment. These stressors can be categorized into two main types: abiotic and biotic stress. Abiotic stress refers to nonliving, environmental factors that can negatively impact plant growth and development. Examples of abiotic stressors include extreme temperatures, drought, salinity, heavy metals, and pollution. When plants encounter these stressors, they activate various mechanisms to adapt and survive. These mechanisms involve physiological, biochemical, and molecular changes that help plants maintain cellular homeostasis, protect themselves from damage, and continue essential metabolic processes. Understanding the responses of plants to abiotic stress is crucial for developing strategies to improve crop productivity and mitigate the negative effects of environmental changes.

On the other hand, biotic stress involves living organisms that can cause harm to plants, such as pathogens (e.g., bacteria, fungi, viruses) and pests (e.g., insects, nematodes). When plants are attacked by these biotic stressors, they elicit complex defense mechanisms to resist or tolerate the damage caused. These defense responses often involve the synthesis of specialized chemicals, activation of specific genes, and morphological changes. Studying the interactions between plants and biotic stressors is essential for developing strategies to enhance plant defense mechanisms and protect crops from diseases and pests. Both abiotic and biotic stress can have detrimental effects on plant growth, development, and productivity. Therefore, understanding the physiological, biochemical, and molecular mechanisms underlying plant stress responses is of great importance. This knowledge can aid in the development of strategies for crop improvement, including the breeding of stress-tolerant varieties and the development of sustainable agricultural practices. Additionally, it can contribute to a deeper understanding of plant biology and the intricate interactions between plants and their environment.

Keywords- - Drought stress, Heat stress, Cold stress, Salt stress, Oxidative stress, Water deficit, Osmotic stress

1. Overview of plant stress physiology

Plant stress physiology is the study of how plants respond and adapt to various stress factors in their environment. Plants are constantly exposed to a range of stressors, including abiotic factors such as drought, extreme temperatures, salinity, and nutrient deficiencies, as well as biotic factors like pathogen infections and herbivory. When plants encounter stress, they undergo a series of physiological and biochemical changes to mitigate the damage caused and ensure survival. These responses can be categorized into two general strategies: avoidance and tolerance.

Avoidance mechanisms involve strategies to prevent or minimize stress impact. For example, plants in dry environments may have adaptations like deep root systems to access water sources, or a waxy layer on their leaves to reduce water loss through transpiration. Similarly, plants may undergo seasonal changes like shedding leaves in response to cold temperatures.

Importance of understanding abiotic and biotic stress responses in plants

Understanding abiotic and biotic stress responses in plants is of utmost importance for several reasons:

1. Crop productivity: Abiotic stresses such as drought, salinity, extreme temperatures, and nutrient deficiencies can significantly reduce crop productivity. By understanding how plants respond to these stresses at a molecular, physiological, and biochemical level, scientists and farmers can develop strategies to enhance plant tolerance and productivity.

2. Sustainable agriculture: Climate change and increasing global population pose immense challenges to food security. By studying how plants respond to biotic stresses such as pests, diseases, and weeds, researchers can develop pest and disease management strategies that minimize the use of chemical pesticides.

3. Crop improvement: Understanding stress responses in plants can contribute to crop improvement programs. By identifying key genes and molecular pathways involved in stress tolerance, scientists can develop genetically modified crops that are more resilient to abiotic and biotic stresses.

4. Ecosystem resilience: Plants play a crucial role in maintaining ecosystem health and resilience. By understanding how abiotic and biotic stressors impact plant growth and performance, scientists can predict and mitigate the potential negative impacts on ecosystems.

5. Human health: Many plants have medicinal properties, and their stress responses can influence the synthesis and accumulation of bioactive compounds with potential health benefits..

Section 1: Abiotic Stress Responses

Abiotic stress refers to the negative impact of non-living factors on plants and organisms. These factors include extreme temperatures, drought, salinity, heavy metals, and various other environmental conditions. In response to abiotic stress, organisms have developed various adaptive mechanisms to survive and mitigate the damage caused by these stressors. Here are some common abiotic stress responses: 1. Tolerance: Some organisms have developed the ability to tolerate or withstand certain abiotic stressors. For example, certain plants can tolerate extreme temperatures by adjusting their physiological processes and metabolic activities.

2. Avoidance: Some organisms have evolved mechanisms to avoid extreme conditions altogether. For instance, certain animals migrate to more favorable environments during periods of extreme temperatures or drought.

3. Morphological adaptations: Organisms may develop physical structures or adaptations to cope with abiotic stress. For example, plants growing in arid regions often have deep root systems to access water from deeper soil layers.

4. Physiological adaptations: Organisms can undergo physiological changes to cope with abiotic stress. For example, plants may close their stomata to reduce water loss during periods of drought or increase the production of certain protective compounds. 5. Biochemical responses: Organisms can produce specific biochemical compounds that help them respond to abiotic stress. For example, some plants produce Osmo protectants like proline or betaine to maintain cellular osmotic balance during drought or salinity.

6. Genetic adaptations: Organisms may possess genetic variations that confer resistance or tolerance to abiotic stress.

1. Definition and examples of abiotic stressors (drought, high/low temperatures, salinity, etc.)

Abiotic stressors refer to non-living factors in the environment that can negatively impact the growth, development, and productivity of organisms.Here are some examples of abiotic stressors:

1. Drought: This occurs when there is a prolonged period of insufficient water supply, resulting in water scarcity. Drought stress can lead to wilting, reduced growth, and even death of plants.

2. High temperatures: Extreme heat can cause thermal stress, especially when combined with low humidity. High temperatures can lead to heat stress in plants and animals, affecting their physiological processes and metabolism.

3. Low temperatures: Extreme cold conditions, such as frost and freezing temperatures, can damage cellular structures, disrupt metabolic processes, and lead to tissue damage in plants and animals.

4. Salinity: High levels of salts in the soil or water, known as salinity, can affect plant growth by reducing water uptake and the ability to absorb essential nutrients.

5. Flooding: Excessive water accumulation due to heavy rainfall or flooding can lead to oxygen deprivation, reduced nutrient availability, and increased susceptibility to diseases in plants. Flooding stress can also impact the respiration process in animals. 6. Nutrient deficiency/toxicity: Imbalances in essential nutrients like nitrogen, phosphorus, potassium, or micronutrients can limit plant growth and development. Conversely, excessive accumulation of certain nutrients can lead to toxicity, causing damage to plants.

7. UV radiation: Ultraviolet (UV) radiation, especially UV-B and UV-C rays, can cause DNA damage, cell membrane disruption, and impaired photosynthesis in plants. It can also lead to skin cancer and other health issues in animals and humans. 8. Pollution: Environmental pollutants, such as air pollutants,

heavy metals, and chemicals, can have detrimental effects on living organisms.

2. Physiological and biochemical responses to abiotic stress:

Physiological and biochemical responses are the various strategies that plants employ to cope with and adapt to these stressful conditions. Here are some common physiological and biochemical responses to abiotic stress:

1. Stomatal closure: Under abiotic stress, plants often close their stomata, which are small pores on the leaf surface, to reduce water loss through transpiration. Stomatal closure helps to maintain water balance and prevent dehydration.

2. Osmotic adjustment: Plants can accumulate compatible solutes, such as proline, sugars, and amino acids, to maintain cell turgor and osmotic potential under water-limited conditions.

3. Antioxidant production: Abiotic stress can lead to the overproduction of reactive oxygen species (ROS), which can cause oxidative damage to plant cells.

4. Heat shock proteins (HSPs): Heat stress triggers the synthesis of heat shock proteins, which act as molecular chaperones that help restore and maintain proper protein folding and stability under extreme temperatures. HSPs help protect cellular structures and maintain normal metabolic processes.

5. Accumulation of compatible solutes: Under salt stress, plants can accumulate compatible solutes like glycine betaine and proline to maintain cellular osmotic balance and minimize the toxic effects of salt ions.

6. Changes in hormone levels: Abiotic stress can alter the balance of plant hormones, such as abscisic acid (ABA), gibberellins (GA), and ethylene (ET). ABA increases under water deficit conditions, promoting stomatal closure and reducing water loss.

A. Water relations and osmotic adjustments

Water relations and osmotic adjustments play crucial roles in helping plants adjust to abiotic stress conditions, such as drought, salinity, and extreme temperatures. These processes help plants maintain proper water balance and cellular integrity, allowing them to survive and thrive in challenging environments. One important aspect of water relations is the regulation of water uptake, transport, and loss in plants. Under drought conditions, for example, plants need to limit water loss through transpiration by closing their stomata, small openings on the leaf surface.

. In freezing temperatures, for instance, plants accumulate cryoprotectants, such as sugars or antifreeze proteins, to lower the freezing point of their cellular fluids and prevent ice crystal formation. Similarly, in extreme heat, plants may accumulate osmolytes to protect cellular structures and maintain proper water balance.

B. Reactive oxygen species (ROS) and antioxidant defense systemsReactive oxygen species (ROS) are highly reactive molecules containing oxygen that are generated as byproducts of various metabolic processes in plants. They include molecules such as superoxide (O2•-), hydrogen peroxide (H2O2), and hydroxyl radical (•OH).

While ROS serve important signaling roles in plant growth and development, their accumulation can be detrimental to plants under abiotic stress conditions, such as drought, high temperature, salinity, and heavy metal exposure. Abiotic stresses can lead to an imbalance between ROS production and detoxification, causing oxidative stress.

C. Hormonal regulation (ABA, ethylene, etc.)

1. ABA (abscisic acid): ABA is a stress-responsive hormone that helps plants survive under adverse environmental conditions. During drought stress, ABA levels increase in plant tissues, triggering various physiological and molecular responses to reduce water loss and maintain cellular integrity. ABA promotes stomatal closure, reducing transpiration and conserving water.

2. Ethylene: Ethylene is a gaseous hormone involved in multiple aspects of abiotic stress responses. It is produced in response to various stresses, including drought, flooding, heat, and high salinity. Ethylene influences plant growth and development, as well as stress adaptation. During drought stress, ethylene inhibits shoot growth while promoting root growth, helping plants enhance water uptake efficiency. It also modulates leaf expansion, stomatal conductance, and seed germination under stress conditions.

D. Photosynthesis and carbon metabolism

Photosynthesis and carbon metabolism play critical roles in the response of plants to abiotic stress. Abiotic stresses such as drought, extreme temperatures, salinity, and nutrient deficiency can negatively affect the photosynthetic process and carbon metabolism in plants. During abiotic stress conditions, plants may experience reduced photosynthetic activity due to stomatal closure, reduced chlorophyll content, and damage to photosynthetic machinery. Stomatal closure is a common response to drought stress, which limits the entry of CO2 into the leaf and thereby reducing the carbon fixation during photosynthesis.

E. Nutrient uptake and transport

Nutrient uptake and transport play crucial roles in the abiotic stress response of plants. Abiotic stress factors such as, salinity, extreme temperatures, and nutrient deficiencies can significantly affect a plant's ability to take up essential nutrients from the soil and transport them to different parts of the plant. This, in turn, can lead to nutrient imbalances and reduced plant growth and productivity. Similarly, salinity stress can disrupt nutrient uptake and transport by affecting root hydraulic conductivity and ion balance. High salt concentrations in the soil can create osmotic stress, reducing water availability for plants and hindering nutrient uptake. Excessive accumulation of certain ions, such as sodium, can also interfere with the uptake of essential nutrients like potassium, calcium, and magnesium.

F. Growth and development

Abiotic stress refers to non-living or environmental factors that can negatively impact plant growth and development. These factors can include extreme temperatures, drought, salinity, and heavy metal toxicity, among others.Growth and development in abiotic stress response is a complex process that involves various physiological and biochemical mechanisms in plants. Here are some key points regarding the growth and development of plants under abiotic stress:

1. Morphological changes: Plants often exhibit morphological changes as a response to abiotic stress. These changes can include altered root architecture, reduced leaf size, and changes in plant height and structure. These adaptations help plants to optimize resource uptake and minimize water loss.

2. Cellular and molecular responses: Abiotic stress triggers various cellular and molecular responses in plants. These responses involve the activation of signaling pathways and the production of stress-related proteins. For example, plants may produce stress proteins called chaperones, which help in repairing damaged proteins and maintaining cellular homeostasis.

3. Metabolic adjustments: Under abiotic stress, plants undergo metabolic adjustments to maintain energy and resource balance. For example, they may alter their metabolic pathways to produce compatible solutes, such as proline and sugars, which act as osmoprotectants and help in maintaining cellular hydration.

4. Hormonal regulation: Phytohormones play crucial roles in coordinating growth and development in response to abiotic stress. For instance, the plant hormone abscisic acid (ABA) is known to regulate stomatal closure to reduce water loss during drought stress. Other hormones, such as auxins, cytokinins, and gibberellins, also play roles in modulating growth and development under abiotic stress conditions.

5. Epigenetic modifications: Abiotic stress can induce epigenetic modifications in plants. These modifications involve changes in DNA methylation, histone modifications, and small RNA expression patterns. Epigenetic modifications can lead to altered gene expression and help plants adapt to stress conditions. Overall, plants have evolved various strategies to cope with abiotic stress and maintain growth and development.

3. Molecular mechanisms underlying abiotic stress responses:

Abiotic stress refers to environmental factors such as temperature extremes, drought, salinity, and heavy metals, which can negatively impact plant growth and development. Understanding the molecular mechanisms underlying abiotic stress responses is crucial for developing strategies to enhance plant tolerance to these stressors.

One molecular mechanism involved in abiotic stress responses is the activation of stress signaling pathways. These pathways involve a series of molecular events that lead to the activation of specific genes involved in stress tolerance. For example, during drought stress, plants activate various signaling molecules such as abscisic acid (ABA) and reactive oxygen species (ROS), which trigger the expression of stress-responsive genes.

Moreover, abiotic stress responses also involve alterations in plant hormone levels. Hormones such as ABA, ethylene, jasmonic acid, and salicylic acid play important roles in coordinating stress responses. For example, ABA is involved in regulating stomatal closure during drought stress, while ethylene is involved in responses to flooding and hypoxia.Overall, the molecular mechanisms underlying abiotic stress responses are complex and involve various signaling pathways, transcriptional regulation, cellular processes, and hormone signaling. Understanding these mechanisms is essential for developing strategies to enhance plant tolerance to abiotic stress and ensure food security in the face of changing climatic conditions.

A. Gene expression regulation

In abiotic stress response, gene expression regulation is a complex process that involves various molecular mechanisms. Here are some key mechanisms involved in regulating gene expression during abiotic stress response:

- 1. Transcriptional regulation: Transcription factors (TFs) play a crucial role in regulating gene expression during abiotic stress. TFs can either activate or repress the transcription of target genes by binding to specific DNA regulatory elements. These regulatory elements are often located upstream of the gene's coding region, and their binding by TFs can either enhance or inhibit gene expression.
- 2. Epigenetic regulation: Abiotic stress can lead to changes in the epigenetic landscape, including DNA methylation and histone modifications. DNA methylation, the addition of methyl groups to DNA molecules, can influence gene expression by modulating the accessibility of DNA to transcriptional machinery.
- 3. Post-transcriptional regulation: Several mechanisms exist at the post-transcriptional level to regulate gene expression during abiotic stress. These mechanisms include alternative splicing, RNA editing, and RNA stability. Alternative splicing allows for the production of different protein isoforms from a single gene, which can enhance stress tolerance. RNA editing

involves the modification of RNA sequences, often resulting in changes in protein coding potential..

- 4. Post-translational modifications: After translation, proteins can undergo various modifications that can impact their activity and stability. For example, protein phosphorylation, acetylation, and ubiquitination are common post-translational modifications that regulate protein function.
- 5. Small RNAs: Small RNAs, such as microRNAs (miRNAs) and small interfering

RNAs (siRNAs), play a crucial role in post-transcriptional regulation during abiotic stress response. These small RNAs can bind to target mRNAs, leading to mRNA degradation or translational inhibition. By targeting specific transcripts, small RNAs can fine-tune gene expression during abiotic stress.

B. Signal transduction pathways

Signal transduction pathways of molecular abiotic stress response refer to the molecular mechanisms by which plants perceive and respond to various abiotic stresses such as drought, salinity, temperature extremes, and nutrient deficiencies. These pathways involve signal perception, signal transduction, and activation of downstream stress-responsive genes.

1. Perception of abiotic stress signals: Plants have specialized receptors, such as receptors for drought stress (e.g., membrane-localized receptor-like kinases) or receptors for salt stress (e.g., ion channels or transporters), which allow them to sense specific abiotic stress signals.

2. Signal transduction: Upon perception of the stress signal, a cascade of biochemical events is initiated to transmit the signal from the receptor to the nucleus, where stress-responsive genes are activated. Various signaling molecules, such as calcium ions (Ca2+), hormones (e.g., abscisic acid or ABA), reactive oxygen

species (ROS), and protein kinases, act as messengers in the transduction process.

3. Activation of downstream stress-responsive genes: In response to the stress signal, specific transcription factors (TFs) are activated or inhibited, which bind to the promoter regions of stress-responsive genes.

4. Gene expression and stress tolerance: The activation of stress-responsive genes leads to the synthesis of stress-related proteins and metabolites. These proteins and metabolites help plants to cope with the abiotic stress conditions by regulating various physiological and molecular processes, including osmotic adjustment, ion homeostasis, antioxidant defense, and repair of damaged cellular components.

C. Stress-responsive genes and proteins

In the field of plant biology, abiotic stresses refer to non-living factors in the environment that can negatively affect plant growth and development. These stresses include high or low temperatures, drought, salinity, and heavy metal toxicity, among others. When plants encounter these adverse conditions, they activate various mechanisms to adapt and survive.

One important aspect of the plant's response to abiotic stress is the activation of stress-responsive genes and synthesis of stressresponsive proteins. These genes and proteins play crucial roles in signaling and physiological pathways that enable plants to cope with the stress.

Stress-responsive genes are often classified based on their involvement in different stress response pathways. For example, stress-responsive transcription factors (TFs) regulate the expression of downstream stress-responsive genes. Examples of stress-responsive TFs include DREB/CBF (Dehydration-Responsive Element Binding/C-repeat binding factor) and AREB/ABF (Abscisic Acid Responsive Element Binding/Abscisic Acid Binding Factors). Other stress-responsive genes encode enzymes involved in the synthesis of protective compounds, such as antioxidants, osmoprotectants, and detoxifying enzymes. For instance, genes encoding superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPX) are upregulated under abiotic stress conditions to counteract oxidative damage caused by reactive oxygen species (ROS). Section 2: Biotic Stress Responses

Biotic stress responses refer to the ways in which plants and other organisms react to and defend against harmful biological factors, such as pathogens (disease-causing organisms) and herbivores (organisms that feed on plants). There are several different types of biotic stress responses that plants can exhibit:

1. Pathogen recognition and immune responses: Plants have mechanisms to detect the presence of pathogens and activate defense responses. These responses may include the production of antimicrobial compounds, reinforcement of cell walls, and the induction of a hypersensitive response, which involves localized cell death to prevent the spread of the pathogen.

2. Induced resistance: When plants are exposed to certain pathogens, they can develop a heightened resistance to future infections. This is known as induced resistance, and it involves the activation of various defense mechanisms, such as the production of defensive proteins or the accumulation of secondary metabolites.

3. Systemic acquired resistance: Systemic acquired resistance (SAR) is a type of induced resistance that is not limited to the site of infection.

4. Hormonal regulation: Plants use hormones, such as salicylic acid (SA), jasmonic acid (JA), and ethylene, to regulate their responses to biotic stress. Different hormones can be involved depending on the type of pathogen or herbivore. For

example, SA is often associated with defense against biotrophic pathogens (which require a living host to survive), while JA is commonly involved in defense against necrotrophic pathogens (which kill host tissues).

5. Physical barriers: Plants have physical barriers, such as waxy cuticles, trichomes (hair-like structures), and thorns, that can help deter or impede herbivores. Some plants also produce toxic or repellent substances to discourage feeding.

6. Interactions with beneficial organisms: Plants can form beneficial associations with certain organisms, such as symbiotic bacteria or fungi.

1. Definition and examples of biotic stressors (pathogens, pests, herbivory, etc.)

Biotic stressors refer to living organisms or their activities that negatively affect other organisms in an ecosystem. These stressors can include pathogens, pests, herbivory, and competition. Here are some examples:

1. Pathogens: Pathogens are microorganisms such as bacteria, viruses, and fungi that cause diseases in plants and animals. Examples include the bacteria causing citrus canker and the fungus causing wheat rust.

2. Pests: Pests are organisms that harm crops, livestock, or other organisms. They can include insects like aphids, caterpillars, and beetles, as well as rodents and birds that consume crops.

3. Herbivory: Herbivory refers to the consumption of plant material by herbivorous organisms. For example, grazing animals such as deer or cattle feeding on grass or insects feeding on leaves can cause significant damage to plants.

4. Competition: Competition occurs when organisms compete for limited resources, such as food, water, or territory. This competition can lead to stress on populations and individuals. For example, plants competing for limited sunlight or animals competing for nesting sites can experience biotic stress.

2. Physiological and biochemical responses to biotic stress:

Biotic stress refers to the negative impact that living organisms, such as pests, diseases, and weeds, have on plants. Plants have evolved various physiological and biochemical responses to counteract biotic stress and ensure their survival. Some of the key responses include:

1. Activation of defense genes: When plants detect the presence of biotic stress, they activate specific genes that initiate defense responses. These genes encode for various defense-related proteins and enzymes that help in fighting off the stress. 2. Production of defense compounds: Plants produce a wide range of secondary metabolites that act as defense compounds against biotic stress. These compounds include phytoalexins, alkaloids, flavonoids, and terpenoids. They inhibit the growth and reproduction of pests and pathogens.

3. Reinforcement of cell walls: Plants reinforce their cell walls as a defense mechanism against biotic stress. This reinforcement makes it difficult for pests and pathogens to penetrate the plant tissues.

4. Production of antimicrobial compounds: Plants produce antimicrobial compounds that inhibit the growth and colonization of pathogens. These compounds can directly kill the pathogens or prevent their growth by interfering with their metabolic processes.

5. Induction of systemic acquired resistance (SAR): SAR is a mechanism by which plants develop resistance against a wide range of pathogens following an initial infection.

6. Activation of hypersensitive response (HR): HR is a rapid defense response that leads to localized cell death at the site of infection.

7. Enhanced production of reactive oxygen species (ROS): ROS are highly reactive molecules that can damage DNA, proteins, and lipids in pests and pathogens.

8. Regulation of hormonal signaling: Plants regulate their hormonal signaling pathways in response to biotic stress. Hormones such as salicylic acid (SA), jasmonic acid (JA), and ethylene play crucial roles in activating defense responses and coordinating various defense mechanisms.

Overall, the physiological and biochemical responses to biotic stress are complex and interconnected. Through these responses, plants can limit the damage caused by pests and pathogens and enhance their chances of survival.

A. Defense mechanisms against pathogens:

Biotic stress responses are defense mechanisms employed by plants to protect themselves against pathogens, such as bacteria, viruses, and fungi. There are several ways plants can respond to biotic stress and prevent pathogen attack:

1. Physical barriers: Plants have physical barriers like the waxy cuticle on leaves, bark on stems, or thorns on stems and leaves that can act as physical barriers to prevent pathogen entry.

2. Cell wall reinforcement: When a plant detects the presence of a pathogen, it can strengthen its cell walls by depositing additional cellulose, lignin, and other compounds. This reinforcement helps prevent pathogen penetration and the subsequent spread of infection.

3. Phytochemicals production: Plants can produce a wide range of secondary metabolites, known as phytochemicals, which have antimicrobial properties. These compounds include alkaloids, phenolics, terpenoids, and flavonoids.

4. Hormonal responses: Plants can produce various hormones, such as salicylic acid (SA), jasmonic acid (JA), and ethylene, which play crucial roles in regulating defense responses against

pathogens. SA is primarily involved in defense against biotrophic pathogens, while JA and ethylene are mainly associated with defense against necrotrophic pathogens.

5. Pathogen recognition and activation of defense genes: Plants have specific receptors, known as pattern recognition receptors (PRRs), which can detect conserved molecular patterns present in pathogens.

6. Systemic acquired resistance (SAR): Once a plant has been infected by a pathogen, it can induce a systemic acquired resistance response, where it activates defense mechanisms throughout the entire plant. This response involves the production of defense proteins and the accumulation of phytochemicals that can hinder pathogen growth and spread.

B. Pathogen recognition and activation of defense responses

Biotic stress refers to the negative impact on plants caused by various living organisms such as pathogens, pests, and weeds. Pathogens, in particular, can cause severe damage to plants and crop yields. Here's a breakdown of the process:

1. Pathogen recognition: Plants possess a range of proteins called pattern recognition receptors (PRRs) on their cell surfaces. These PRRs are capable of recognizing specific molecular patterns, called pathogen-associated molecular patterns (PAMPs), which are unique to pathogens. PAMPs can be components of the pathogen's cell wall, cell surface molecules, or released by the pathogen during infection.

2. PAMP-triggered immunity (PTI): Upon recognition of PAMPs by PRRs, plants initiate a rapid immune response known as PAMP-triggered immunity. This response involves the activation and signaling of various defense-related genes, resulting in the production of antimicrobial compounds, reinforcement of the cell wall, and oxidative burst, which generates reactive oxygen species (ROS) to kill the pathogen.

C. Induced systemic resistance (ISR) and systemic acquired resistance (SAR) Induced systemic resistance (ISR) and systemic acquired resistance (SAR) are two important defense mechanisms employed by plants to protect themselves against biotic stress caused by pathogens.

ISR is a localized defense response that occurs when a plant is exposed to a beneficial microorganism, such as certain types of bacteria or fungi.

On the other hand, SAR is a more wide-ranging defense response that occurs in the entire plant after it has been infected by a pathogen. When a plant is attacked by a pathogen, it recognizes the presence of the pathogen and triggers a systemic response throughout its tissues.

Both ISR and SAR are effective mechanisms for protecting plants against pathogen attacks. By activating these defense responses, plants can enhance their resistance to biotic stress and minimize the damage caused by pathogens.

D. Defense mechanisms against pests and herbivory:

1. By Physical barriers and structural defenses: Plants have evolved various physical barriers and structural defenses to protect themselves against pests and herbivores. These include thorns, spines, and prickles, which deter herbivores from feeding on the plant. Other physical barriers include tough and waxy outer layers, such as bark or cuticles, which make it difficult for pests to penetrate the plant's tissues. Some plants also have specialized structures like trichomes (hairs) or tough leaves that can physically impede pests and make it harder for them to consume plant tissues. ii. By Induced defenses and chemical signaling in biotic stress: Plants have the ability to produce a wide range of secondary metabolites, such as alkaloids, terpenoids, and phenolics, which can act as chemical defenses against pests and herbivores. These compounds can be toxic or repellent to pests, making the plant less attractive or harmful to them.

3. Molecular mechanisms underlying biotic stress responses:

A. Pattern recognition receptors (PRRs) and pathogenassociated molecular patterns (PAMPs): PRRs are proteins present on the surface of plant cells that can recognize specific molecules, called PAMPs, which are commonly found in microbial pathogens.

B. Effector-triggered immunity (ETI): ETI is another layer of the plant's immune response. During infection, some plant pathogens deliver specific effector proteins directly into the plant cells to suppress host defenses and promote infection.

C. Plant defense-related genes and signaling pathways: Plants have evolved various defense-related genes and signaling pathways to combat biotic stress. These genes encode for proteins involved in the synthesis of antimicrobial compounds, such as phytoalexins, or proteins that directly inhibit pathogen growth, such as chitinases or glucanases.

Section 3: Crosstalk between Abiotic and Biotic Stress Responses

Crosstalk between abiotic and biotic stress responses refers to the interaction and intersection of the molecular and physiological pathways involved in plants' ability to respond to both abiotic (non-living) and biotic (living) stress conditions. Plants are constantly exposed to a variety of stress factors, including drought, temperature extremes, salinity, nutrient deficiencies, and various pathogens such as bacteria, fungi, viruses, and insects. The ability of plants to cope with and adapt to these stressors is crucial for their survival and productivity. One example of crosstalk between abiotic and biotic stress responses is the involvement of the phytohormone jasmonic acid (JA). JA is well-known for its role in plant defense against chewing insects and necrotrophic pathogens. However, it has also been shown to play a role in response to abiotic stresses such as drought, salinity, and extreme temperatures. This suggests that JA signaling pathway components are shared between abiotic and biotic stress responses.

Biotic stresses, caused by pathogens or pests, can trigger the activation of specific hormonal signaling pathways. For example, SA is commonly associated with defense responses against biotrophic pathogens, while JA and ethylene are involved in defense against necrotrophic pathogens and herbivores.

Interestingly, these hormonal pathways not only regulate defense against biotic stress but also participate in responses to abiotic stress. ABA, in particular, plays a pivotal role in mediating responses to drought, salinity, and cold stress. It helps regulate stomatal closure, promotes seed dormancy, and activates various stress-responsive genes.

The crosstalk between hormonal pathways is complex and involves both synergistic and antagonistic interactions. For instance, SA and JA signaling pathways often exhibit antagonistic interactions, where high levels of SA can suppress JA-mediated defense responses and vice versa.

Understanding the intricate crosstalk between hormonal signaling pathways is crucial for developing strategies to enhance plant resilience to both biotic and abiotic stresses. Manipulating these pathways can potentially lead to the development of stress-tolerant crops with improved yield and sustainability.

Strategies for improving stress tolerance in plants:

a. Genetic engineering approaches

b. Breeding for stress tolerance

c. Enhanced stress management and crop management practices

a. Genetic engineering approaches: Genetic engineering involves manipulating the genes of plants to enhance their stress tolerance. This can be done by introducing genes from other organisms that confer stress tolerance traits, such as genes for drought resistance or cold tolerance. Genetic engineering can also be used to enhance the expression of existing genes related to stress tolerance in plants.

b. Breeding for stress tolerance: Breeding programs can be implemented to select and breed plants with improved stress tolerance traits. This involves cross-breeding plants that exhibit natural stress tolerance traits or identifying and selecting plants that perform well under stressed conditions.

c. Enhanced stress management and crop management practices: Implementing stress management techniques and crop management practices can help improve stress tolerance in plants.. By optimizing growing conditions and minimizing stress factors, plants can better tolerate stress and maintain their productivity.

Conclusion

In conclusion, this chapter has provided an overview of plant stress physiology, focusing on the response of plants to various stressors such as drought, high temperatures, and nutrient deficiencies. We have discussed the mechanisms by which plants cope with stress, including physiological and biochemical adaptations. Some key concepts covered in this chapter include the role of hormones in regulating plant stress responses, the importance of osmotic adjustment in maintaining cellular water balance, and the role of antioxidants in protecting plants from oxidative damage. It is clear from this discussion that further research is needed in the field of plant stress physiology. As climate change continues to pose challenges to crop production, understanding how plants respond and adapt to stress will be crucial for developing sustainable agricultural practices. By uncovering the underlying mechanisms of stress tolerance in plants, researchers can identify genes and traits that can be targeted for crop improvement through breeding or genetic engineering.

The implications of this research for sustainable agriculture are significant. By developing crops that are more resilient to stressors like drought and high temperatures, we can reduce yield losses and ensure food security in the face of climate change. Additionally, improving our understanding of plant stress physiology can also inform the development of more efficient irrigation and nutrient management strategies, reducing water and fertilizer use and minimizing environmental impacts.

In conclusion, plant stress physiology is a complex and important field of research, with implications for both basic science and practical applications in agriculture. Continued research in this area will be essential for addressing the challenges posed by climate change and ensuring sustainable crop production for future generations.

References

Chaves, M.M., Maroco, J.P., Pereira, J.S. (2003). Understanding plant responses to drought — from genes to the whole plant. Functional Plant Biology, 30, 239-264.

Mittler, R. (2006). Abiotic stress, the field environment and stress combination.

Trends in Plant Science, 11(1), 15-19.

Verslues, P.E., Agarwal, M., Katiyar-Agarwal, S., Zhu, J. and Zhu, J.K. (2006). Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status. The Plant Journal, 45(4), 523-539.

Rizhsky, L., Liang, H. and Mittler, R. (2002). The combined effect of drought stress and heat shock on gene expression in tobacco. Plant Physiology, 130(3), 1143-1151. Munns, R. and Tester, M. (2008). Mechanisms of salinity tolerance. Annual Review of Plant Biology, 59, 651-681.

Foyer, C.H. and Noctor, G. (2005). Redox homeostasis and antioxidant signaling: a metabolic interface between stress perception and physiological responses. The Plant Cell, 17(7), 1866-1875.

Xu, Y., Burgess, P., Zhang, X., Huang, B. (2014). Enhancing plant tolerance to abiotic stress by gene transfer of a novel aldehyde dehydrogenase gene isolated from the resurrection grass Sporobolus stapfianus. Plant Cell Reports, 33(12), 207-223. Mittler, R. (2002). Oxidative stress, antioxidants and stress tolerance. Trends in Plant Science, 7(9), 405-410.

Suzuki, N., Rivero, R., Shulaev, V., Blumwald, E., Mittler, R. (2014). Abiotic and biotic stress combinations. New Phytologist, 203(1), 32-43.

Chaves, M. M., Flexas, J., & Pinheiro, C. (2009). Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. Annals of Botany, 103(4), 551-560.

Munns, R., & Tester, M. (2008). Mechanisms of salinity tolerance. Annual Review of Plant Biology, 59, 651-681.

Mittler, R. (2006). Abiotic stress, the field environment and stress combination.

Trends in Plant Science, 11(1), 15-19.

Shinozaki, K., & Yamaguchi-Shinozaki, K. (2007). Gene networks involved in drought stress response and tolerance. Journal of Experimental Botany, 58(2), 221-227.

Krasensky, J., & Jonak, C. (2012). Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. Journal of Experimental Botany, 63(4), 1593-1608.

Tuteja, N. (2007). Mechanisms of high salinity tolerance in plants. Methods in Enzymology, 428, 419-438.

Verslues, P. E., & Juenger, T. E. (2011). Drought, metabolites, and Arabidopsis natural variation: a promising combination for understanding adaptation to water-limited environments. Current Opinion in Plant Biology, 14(3), 240-245.

Aliferis, K. A., & Jabaji, S. (2010). Metabolomics – a robust bioanalytical approach for the discovery of the modes-of-action underlying the phenotypic variations of plant-pathogen interactions. Journal of Plant Pathology, 92(1), 37-47.

Anderson, J. P., Badruzsaufari, E., Schenk, P. M., Manners, J. M., Desmond, O. J., Ehlert, C., ... & Kazan, K. (2004). Antagonistic interaction between abscisic acid and jasmonateethylene signaling pathways modulates defense gene expression and disease resistance in Arabidopsis. The Plant Cell, 16(12), 3460-3479. Bari, R., &

Jones, J. D. (2009). Role of plant hormones in plant defence responses. Plant Molecular Biology, 69(4), 473-488.

Glazebrook, J. (2005). Contrasting mechanisms of defense against biotrophic and necrotrophic pathogens. Annual Review of Phytopathology, 43, 205-227.

Pieterse, C. M. J., Van der Does, D., Zamioudis, C., Leon-Reyes, A., & Van Wees, S.

C. M. (2012). Hormonal modulation of plant immunity. Annual Review of Cell and Developmental Biology, 28, 489-521.

Robert-Seilaniantz, A., Grant, M., & Jones, J. D. (2011). Hormone crosstalk in plant disease and defense: more than just jasmonate-salicylate antagonism. Annual Review of Phytopathology, 49, 317-343.

Vlot, A. C., Dempsey, D. M., & Klessig, D. F. (2009). Salicylic acid, a multifaceted hormone to combat disease. Annual Review of Phytopathology, 47, 177-206. Wu, J., & Baldwin, I. T. (2010). Herbivory-induced signalling in plants: perception and action. Plant, Cell & Environment.



Vishnu^{*1}, Siddharth²

¹Sam Higginbottom University of Agriculture Technology and Sciences [SHUATS] Prayagraj - 211007, India ²University of Queensland, St. Lucia, Brisbane, QLD - 4072, Australia. Correspondence author: <u>siddharthbamnia@gmail.com</u>

Abstract

During a procedure referred to as photosynthesis, the phototrophs convert light energy into chemical energy, which is subsequently utilised to power cellular functions. Sugars act as a kind of storage medium for chemical energy and are made from water as well as carbon dioxide.

Increasing photosynthetic activity is essential since arable land is becoming scarcer and there are more people living on the planet. In addition to giving humans food, fiber, and a variety of other critical elements, oxygenic photosynthesis is an important activity because it affects almost every aspect of life on Earth. Water and carbon dioxide are changed (H₂O) into organic molecules like glucose (C₆H₁₂O₆), sugars, and starches using solar energy, plants, algae, and cyanobacteria also release molecular oxygen into the atmosphere.

Introduction

A rapid method that uses solar radiation to quickly prepare plant meals. By utilising photosynthetic pigments like carotene, xanthophyll, chlorophyll a, and b, the process of photosynthesis is said to only take place in chloroplasts. Photosynthesis is a process that all green plants employ to convert carbon dioxide, water, and sunlight into food. A result of the photosynthetic process is oxygen. Let's examine the activity, consequences, and procedure of photosynthesis in more detail.An additional revolution in agriculture centred around plant genetic engineering was expected to increase plant output and, as a result, significantly lessen famine.In order to increase the plant's resistance to diseases and drought, biological scientists have been able to change the genetic code (DNA) of the plant from the 1970s, its capacity for producing more and better-quality products, its capacity for withstanding winter, and other desired traits. Due to the inherent complexity of these features, genetic engineering has proven to be more challenging than initially thought. Such genetic engineering may improve photosynthesis in the future, but as of the first few years of the twenty-first century, it has not yet demonstrated considerable increase agricultural output.

The discovery that some organisms make chemical energy by converting light energy has sparked interest in yet another exciting area of photosynthesis study. For instance, the emerald green sea slug (Elysia chlorotica) can only create a small amount of chlorophyll because it consumes the alga Vaucheria litorea, which provides it with genes and chloroplasts. Once enough chloroplasts have been ingested, the slug may cease eating. The generation of carotenoid pigments by the pea aphid (Acyrthosiphon pisum) has been linked with aphid's capacity to utilise light to make the energizing compound adenosine triphosphate (ATP).

The Greek terms phs, meaning "light," and synthesis, meaning "combining together," are the origin of the name "photosynthesis." This signifies "**combining with the aid of light**."

In addition to plants, other creatures can also engage in photosynthesis. Numerous prokaryotes, including cyanobacteria, purple bacteria, and green sulfur bacteria, are among them. The photosynthesis of these organisms is comparable to that of green plants. The numerous biological processes are then powered by the glucose created during photosynthesis. Oxygen is a by-product of this process.



History

The discoveries of the English scientist and cleric Joseph Priestley in 1771 marked the beginning of the study of photosynthesis. Priestley inserted a lit candle inside the tightly closed container and allowed it to burn until the air inside could ceased to support combustion. After a few days, he discovered the mint plant had created a molecule that would later be known as oxygen, allowing the compressed air to sustain burning once more. He then filled the container with a mint sprig. The Dutch physician Jan Ingenhousz improved on Priestley's findings in 1779 by demonstrating that plants needed exposure to light to replenish the combustible component (oxygen). Additionally, he offered proof that the plant's green tissues are crucial for this.

The gas that fuels combustion (oxygen) was demonstrated to have been produced in 1782 through the extraction of a different gas, or "fixed air," which was previously recognised as carbon dioxide the year before.

In 1804 a plant grown in an exact weighted pot gained weight through the absorption of carbon, it was totally through the ingesting water and carbon dioxide, which was absorbed by roots of plants and oxygen that was left over was released back into the atmosphere. Before it was discovered in 1845 that the energy from sunlight could be preserved as a chemical energy within the leftovers of photosynthesis, the concept of chemical energy hadn't sufficiently matured.

Importance

- 1) Photosynthesis is another way that algae convert solar energy into chemical energy. As a result of photosynthesis, which is assumed to be initiated by light, oxygen is produced.
- 2) Plants use energy from light throughout photosynthesis to change water and carbon dioxide into glucose & oxygen. The chloroplasts are little biological entities found in leaves.
- 3) Chlorophyll, a pigment with a green colour, is present in every chloroplast. The molecules of chlorophyll capture the energy of light, whereas carbon dioxide and oxygen penetrate via stomata found in the outermost layer of leaves.
- 4) Sugars like fructose and glucose are another by-product of photosynthesis.

These sugars are then delivered to the roots, stems, leaves, fruits, flowers, & seeds. In the simplest terms, the sugars give plants an alternative source of energy, allowing them to grow and thrive. More complicated carbohydrates like cellulose and starch are created whenever these sugar molecules mix. It is believed that the structural element of cell walls in plants is cellulose.

Site of the process

In both vegetation and blue-green algae, photosynthesis takes place in chloroplasts. Chloroplasts, or plastids with a green tint, are found in all the green parts of a plant, including the green stems, leaves, and sepals. Cell organelles such as these are specific to plant cells and are found in the mesophyll cells of leaves.

The green plants' photosynthetic organelles are chloroplasts.The chloroplasts are the only site where photosynthesis occurs in plants. Robert Hill, a British biochemist, conducted extensive studies on the
function of these organelles. Hill observed that green particles produced by damaged cells may produce oxygen in the presence of light and a substance like ferric oxalate from water, which can serve as an electron acceptor. This was around the year 1940. The Hill reaction is the name given to this mechanism. In the 1950s, American biochemists Daniel Arnon and others created plant cell fragments that underwent both the Hill reaction and the creation of the energy-storing molecule ATP. The non-physiological electron acceptors utilised by Hill were replaced with the coenzyme NADP as the ultimate electron acceptor. His methods were improved so that the Hill reaction could be carried out by tiny, isolated lamellae, or chloroplast membrane fragments. Then, these tiny fragments of lamellae were broken up into smaller particles that could only carry out the light reactions required for photosynthetic activity.

Furthermore, the complete photosynthetic process, including light absorption, oxygen creation, carbon dioxide reduction, and the production of glucose and other products, may now be performed by the entire chloroplast when it is isolated.

Factors that influence the rate of photosynthesis

The rate of photosynthesis is calculated by divide the amount of oxygen production either per unit mass within green plant tissues or per unit weight of all chlorophyll. The environmental parameters that have the biggest impact on how quickly terrestrial plants synthesize oxygen and carbon dioxide are light, carbon dioxide availability, temperature, water availability, and mineral availability. The physiological state of the plant, including its health, maturity, and whether it is in bloom, as well as its kind, affect the rate of photosynthesis.

Temperature and intensity of the light

The two distinct phases of the complex procedure known as photosynthesis—the photochemical or light-harvesting phase, and

second enzymatic or carbon-assimilating phase-which includes reactions-have previously been established. chemical To determine these stages, examine the rate of photosynthesis with different temperatures and light intensity levels. The process of photosynthesis increases with the amount of light and is not significantly impacted by temperature across an array of moderate temperatures and low to medium-level light intensities, in contrast to the typical spectrum of plant species. Light saturation is reached at a particular light intensity depending on the species and growing environment; however, as the light intensity rises, the rate becomes saturated. As a result, dependency is the phase of light that precedes saturation. The rate at which photochemical processes influences the rate of photosynthesis. The rate of various chemical reactions in the dark stage is slowed down by high light intensities. Photorespiration, a common practice among land plants, has an escalating impact on photosynthesis as temperatures rise. More specifically, photorespiration hinders future improvements in the rate of photosynthesis because it outcompetes it, particularly when water is scarce.

Carbon dioxide

a few among the restricting procedures during the dark phase stage of photosynthesis are chemical interactions which generate organic compounds utilising carbon dioxide as a source of carbon.By boosting the quantities of carbon dioxide, these processes can be somewhat accelerated. Since the middle of the 19th century, a rise in the level of carbon dioxide in the atmosphere has been attributed to land use shifts carried on by deforestation, extensive use of fossil fuels, and cement production.Better measurements initially started in 1958, and by 2020 the atmospheric carbon dioxide content had increased from about 0.028 percent to 0.032 percent and subsequently to .042 % . This rise in carbon dioxide has a direct impact on varying levels of photosynthesis, depending on the species and the plant's physiological status. According to the majority of scientists, rising atmospheric carbon dioxide levels have an impact on the climate by changing rainfall patterns and raising global temperatures. The rate of photosynthetic production will also be affected by these changes.

Water

When it comes to photosynthesis and plant growth, water availability might be a limiting element for terrestrial plants. Leaves drain an enormous quantity of water or instead, water evaporates through the foliage to the environment through the stomata, in spite of the tiny amount of water required for the true photosynthetic process. The leaf epidermis, or outer skin, contains small pores called stomata, which compelably allow water vapour to escape. Water vapor can enter through stomata as well as leave. The leaf's stomata are going to open and then close in response to needs. In order to conserve water in hot, dry climates, stomata can close; however, limits dioxide this carbon uptake and slows photosynthesis.

Reduced transpiration causes the outside temperature of the plant's leaves to increase, which reduces cooling. The greater leaf temperatures and lesser carbon dioxide amounts stimulate the ineffective photorespiration pathway. Whenever the quantity of carbon dioxide in the environment increases, greater quantities of carbon dioxide can reach the plant's stomata via a narrower hole. There may be greater photosynthesis with the same amount of water as a result.

Minerals

For healthy plant development and good photosynthesis rates, several minerals are important. Significant quantities of nitrogen, sulphate, phosphate, iron, magnesium, calcium, and potassium must be present for the production of amino acids, proteins, coenzymes, deoxyribonucleic acid (DNA) and ribonucleic acid (RNA), chlorophyll and other pigments, and a number of other essential components of plants. Less manganese, copper & chloride are required for photosynthesis to take place. Further trace substances are needed for many different kinds of plant functions that are not photosynthetic.

Internal factors

Each kind of plant has evolved over time to accommodate varied environmental variables. Within this typical range of environmental conditions, intricate regulatory systems found within the plant's cell control the function of enzymes or organic catalysts. In accordance with the requirements of the entire plant, these modifications control and maintain the equilibrium of the entire photosynthetic process. It is possible that doubling the carbon dioxide concentration will result in a temporary boost in photosynthesis that is almost twice as fast for a particular plant species. Since the plant couldn't consume all of the sucrose produced during photosynthesis, the rate may drop a few hours or days later and return to its former level. A different plant species, on the other hand, would continue to photosynthesize and develop more quickly over the majority of its life cycle and Considering that it possessed more carbon-demanding organs, it could possibly be capable to make use of higher sucrose.

Photosynthesis's reaction or process

At the cellular level, chloroplasts are the cell organelles where photosynthesis takes place. These organelles contain the chlorophyll, which gives the leaves their unique green color. As was already noted, photosynthesis takes place in leaves, and the chloroplast is the specialized cell organelle in charge of this process. A leaf's petiole, epidermis, and lamina are its structural constituents. The lamina absorbs sunlight and carbon dioxide during photosynthesis. An oxidation-reduction reaction that is light-accelerated is photosynthesis. The process or reaction involved in photosynthesis Chloroplasts are the cell organelles that carry out photosynthesis at the cellular level. The chlorophyll, a green pigment that gives plants their distinct green color, is found in these organelles. As has already been mentioned, the chloroplast is the specialized cell organelle in charge of photosynthesis in leaves. The petiole, epidermis, and lamina of a leaf make up its structural elements. During photosynthesis, the lamina absorbs carbon dioxide and sunlight. Photosynthesis is an oxidation-reduction reaction catalyzed by light. (A molecule obtains electrons through the reduction process; a molecule loses electrons through the oxidation process.)

Light

Equation - $CO_2 + 2H_2O \longrightarrow (CH_2O) + O_2 + H_2O$

Since there are numerous reactions involved in photosynthesis which are accelerated through organic catalysts named enzymes, the above equation offers a synopsis. The "light" phase of such reactions, whose consists of photochemical reactions (or light-capturing) responses, or the "dark" stage, that consists of chemical reactions controlled by enzymes, were the two phases that comprise these processes. The energy from light which is absorbed throughout the first stage of photosynthesis powers a series of electrons transfers that result in ATP or the electrons-donor-reduced nicotinic adenine dinucleotide phosphate (NADPH). By the dark phase, carbon dioxide is converted to organic carbon molecules utilising the ATP and NADPH produced through the light-capturing systems. Carbon fixation is the process of assimilating carbon that is inorganic into organic compounds. In the course of the twentieth century, studies comparing the photosynthetic processes of plant life and certain photosynthetic sulphur bacteria revealed significant knowledge regarding the mechanism. Sulfur-producing bacteria use hydrogen sulphide (H2S) as an alternative source of hydrogen atoms

throughout photosynthesis in order to produce sulphur instead of oxygen.

Light

The equation : $CO_2 + 2H_2S \longrightarrow (CH_2O) + S_2 + H_2O$

Sulfur bacteria

The significant finding that both types of photosynthetic organisms use carbon dioxide to make organic chemicals was established in the 1930s by Dutch researcher Cornelis van Niel. He suggested that hydrogen was transferred from a known acceptor (named A), which was then reduced to H2A, to either water or hydrogen sulfide, both of which are abundant in bacteria and green plants. He went on to say that there are also variations in the light-dependent stage and the types of chemicals employed as hydrogen atom suppliers. The reduced acceptor (H2A), which may be found in bacteria and green plants, Through the dark procedures mixed with carbon dioxide (CO2) to produce carbohydrate (CH2O), it oxidised the unknown acceptor to A.

This equation can be shown as:

Light

 $CO_2 + 2H_2A$ $(CO_2 + 2H_2A + H_2O)$

According to a widely accepted (but incorrect) idea, carbon dioxide had its oxygen removed and after that, carbon and water united to produce carbohydrates. This is why Van Niel's suggestion was significant.

Application of Photosynthesis Reaction

Every time we cook or bake, we use chemical reactions. After that, we combine the components (the reactants), put them in an appropriate environment (typically heat), and then take pleasure in the end result (the products). The photosynthesis reaction is shown below:

 $6CO_2 + 6H_2O + Light C_6H_{12}O_6 + 6O_2$

The equation demonstrates that carbon dioxide, light energy, and water are the necessary components for the photosynthetic process. Light from the sun, CO_2 from the environment, water molecules from their surroundings, and light from the algae are all used by algae, plants, and photosynthetic microorganisms to produce glucose.

Of course, light, water, and carbon dioxide combine in the atmosphere even in the absence of plants. However, they do not undergo chemical change in order to produce food unless very stringent requirements are met. These are also exclusive to the cells of creatures that use photosynthetic mechanisms. Enzymes, proteins that hasten chemical reactions, are a prerequisite for this.

Chlorophyll is a pigment found in plant cells that absorbs light.

c)Chloroplasts are organelles with membranes that contain enzymes, chlorophyll, and auxiliary pigments in arrangements that maximise photosynthesis.

Chloroplasts are structures found inside plant or algal cells that organise the enzymes, chlorophyll, and other pigment molecules required for photosynthesis.

Photosynthesis Steps

1. Water is drawn off the ground through its root hairs and transferred onto the leaves via the vessels in the xylem throughout photosynthesis. Stomata allow carbon dioxide to pass through the circulatory system. By receiving solar energy, chlorophyll converts the molecules of water into hydrogen and oxygen.

2. Whenever glucose is ingested, carbon dioxide in the air combines with hydrogen via water molecules. Additionally, oxygen is expelled onto the atmosphere via the leaves being a waste product.

3. Glucose is a nutrient plants usage to promote their development as well as their growth. Any surplus glucose accumulates in the roots, leaves, and fruits of plants for future use.

4. Another biological element involved in photosynthesis is that is extremely important is pigment. In addition to reflecting light that was not absorbed while absorbing light of a specific wavelength, these molecules are responsible for color. The main elements of every green plant is chlorophylls a, b, and carotenoids, that are located in the thylakoids of the chloroplasts. Its primary function is to absorb the energy of light. Chlorophyll is the main colour.

Following are the two phases of photosynthesis:

- 1. The light response, often known as the light-dependent response
- 2. Dark reaction or reaction not requiring light

The light response, often known as the light-dependent response

- The light response, which can only occur in the presence of sunshine throughout the day, is the first step in photosynthesis. The light-dependent process occurs in plants' thylakoid membranes, which are found on the chloroplasts.
- The Grana, which are membrane-bound sac-like structures found inside thylakoids and are home to the photosystems, function by capturing light.

- The pigment and protein components required for photosynthesis are found in significant complexes in the plant cells that house these photosystems.
- In the light-dependent activities, which are necessary for the second phase of photosynthesis, light energy is transformed into ATP and NADPH. The photosystem I and the photosystem II constitute two different types of photosystems.
- A pair of electron transport chains rely on ATP, NADPH, oxygen, and water to carry out the light reactions, which result in these byproducts.

Dark reaction or reaction not requiring light

- The reaction of carbon-fixation, commonly referred to as the dark reaction, is a light-independent process that converts carbon dioxide and water molecules into sugar molecules.
- The ATP and NADPH intermediates of the light-induced reaction are utilised via the dark reaction, that occurs in the stroma of the chloroplast.
- Plants begin the process known as the Calvin cycle of photosynthesis after they take in carbon dioxide via their stomata, and they utilise the ATP and NADPH generated after a light reaction to carry out the reaction and convert six separate molecules of carbon dioxide create one sugar molecule.

Light reactions: $2H_2O + light \rightarrow O_2 + 4H_+ + 4e_- \quad G^\circ = +317 \text{ kJ/mol}$ Dark reactions: $CO_2 + 4H_+ + 4e_- \rightarrow CH_2O + H_2O \quad G^\circ = +162 \text{ kJ/mol}$ Overall: $H_2O + light + CO_2 \rightarrow CH_2O + O_2 \quad G^\circ = +479 \text{ kJ/mol}$

Artificial photosynthesis

In order to create a useful, affordable, and a successful process for turning sunlight into fuels that can be stored, such as hydrogen, artificial photosynthesis makes an effort to emulate this natural process. This is usually accomplished by creating photoelectrochemical cells that harness light to divide water into hydrogen and oxygen, according to Wang, X. et al. (2019); Kalyanasundaram, K. et al. (2010). As of right now, there isn't a technology that can effectively and affordably catalyze these processes, which presents difficulties for this project.

Comparatively, even though both artificial and natural photosynthesis utilise sunlight, their methods of execution and efficacy differ greatly.

Natural photosynthesis has evolved over billions of years to be very effective, converting sunlight into chemical energy at a rate of about 3-6%. Contrarily, it is currently challenging for artificial systems to achieve equivalent efficiencies at a comparable cost (Hong, Y.H et al., 2022). Artificial photosynthesis has the potential to produce fuels that are energy-dense, in contrast to natural photosynthesis, which mostly results in the production of glucose. Therefore, artificial photosynthesis may prove to serve as a long-term solution for our energy needs, even though it is still in the early phases of research, assisting in the fight against climate change and ensuring the security of our energy supply (Shen, L. et al., 2021; Sukhova, E. et al., 2020).

Benefits

1. One benefit of artificial photosynthesis is that it can offer a clean, renewable energy source that doesn't use fossil fuels or emit greenhouse gases.

2. It can utilise cheap, plentiful resources like sunshine, water, and carbon dioxide, which are nearly always present on Earth.

3. It can create a range of fuels or chemicals that are simple to store and transport, like ethylene, methane, hydrogen, and ethanol.

4. It might be more cost-effective and efficient than other solar energy sources, including photovoltaics or solar thermal.

Disadvantage

1. Finding effective and durable catalysts or materials that can carry out the water-splitting and carbon dioxide reduction reactions under sunlight is one of the many obstacles that artificial photosynthesis, which is still in its infancy, must overcome.

2. Creating and improving tools or systems that can integrate the many steps and elements of synthetic photosynthesis.

3. Taking the technology from the lab to the industrial level while maintaining its safety and minimal impact on the environment

The development of artificial photosynthesis holds great promise for the future of energy production and consumption. By lowering human activity's carbon footprint, it might also aid in lessening the effects of climate change. To get over the present constraints and difficulties, additional research and innovation are necessary.

Sr.	Factors	Natural	Artificial
No.			
1	Reaction place	Chlorophyll in photo-system	Photo- electrochemical
		1 5	cells
2	Storage of energy	Glucose	Hydrogen or other solar fuels
3	Carbon fixation	Yes	Potentially
4	Effect	3–5%	Variable, still under development
5	Utiliztion of product	Mainly through food and biomass	Only fuels for energy and industry
6	Catalysts	Enzymes	Man-made catalysts
7	Reaction rate	Relatively slow	Potentially faster
8	Process operating conditions	Surroundings temperature and pressure	Variable, can be optimized for reaction
9	Evolution	Billions of years of natural selection	Still under development
10	Water dependency	High, water is electron donor	High, water often used for proton/electron source

Table 1 : Difference between artificial and naturalphotosynthesis

Products of photosynthesis

Carbohydrates are the primary direct organic product of photosynthesis in the majority of green plants, as was already mentioned. The creation of glucose, a simple carbohydrate, is signaled chemically.

Sunlight

Equation - $6 \text{ CO}_2 + 12\text{H}_2\text{O} \longrightarrow \text{C}_6\text{H}_{12}\text{O}_6 + 6\text{O}_2 + 6\text{H}_2\text{O}$

Carbon dioxide Water Glucose oxygen water

Plants don't produce much free glucose; instead, molecules of glucose are linked to create starch. Contrary to popular belief, photosynthesis also results in the formation of the proteins, lipids, pigments, and other organic materials found in green tissues. The components needed to create these compounds, including nitrogen, phosphorus, and sulfur, are found in minerals. While the chemical compounds among oxygen ,carbon , hydrogen , nitrogen , and sulphur break down and fresh chemical bonds develop, products like gas oxygen (O2) and molecules of organic matter are produced. It requires a greater amount of energy instead of is produced whenever fresh chains develop in the byproducts in order to break the chains which bind oxygen with other elements (such those in water, nitrate, and sulphate). A significant amount of the energy generated by light is retained as chemical energy in the organic compounds made throughout photosynthesis due to the disparity in bond energy. Converting simple molecules to complex ones requires more energy.

ATP production from light energy

ATP and NADPH must be synthesized in order for the dark reactions to take place, and the energy for these reactions comes from the electron transfers of the light reactions. The result is a decrease in NADP+ to NADP is the noncyclic electron transport discussed in the section preceding. This section explains the process that creates the potent chemical ATP.

Adenosine Diphosphate (ADP) is converted into ATP by adding a phosphate group, or, to put it chemically, by phosphorylating ADP. The link connecting the additional phosphate group to ADP is where most of the energy needed for this reaction is stored. The electrontransport chain within the mitochondrion is not known as phosphorylation by oxidation, this process in the chloroplasts during photosynthesis is called photophosphorylation because light energy drives it.

When photophosphorylating ADP, both cyclic and noncyclic electron transport are involved, unlike when making NADPH. In fact, because photophosphorylation doesn't require a exact electron transport to reducing agents, researchers speculate that photophosphorylation might be the only reason for cyclic electron flow. A modification in the mix of cyclic and noncyclic flow could be caused by an alteration in the physiological requirements for ATP, reduced ferredoxin, and NADPH in chloroplasts.

Even though light reactions one and two can use membrane fragments for electron transfer, photophosphorylation necessitates the presence of entire thylakoids. The unique characteristics of the system connecting photophosphorylation and electron transport in the lamellae necessitate the necessity for this.

English biochemist Peter Dennis Mitchell, recipient of the 1978 Nobel Prize in Chemistry, was the first to propose the link between ATP synthesis and both mitochondrial and chloroplastic membranes both experience electron flow, the organelles in charge of generating ATP during cellular respiration. The chemiosmotic theory has been slightly altered in order to account for more recent experimental findings. Now, many people are familiar with the traits. The development of an electrical charge and a gradient of hydrogen ion (proton) concentration across entire lamellae is a critical characteristic.

It is possible to see inside the thylakoid the manganese-protein complex connected to light reaction II. In light reaction two, hydrogen ions are released enter the inside thylakoid region as a consequence of the oxidation of water. The fact that plastoquinone molecules require protons from the outside of the thylakoid in order to decrease means that it is also likely that photoreaction two involves the passage of electrons through the lamella towards its outer face. Protons are liberated inside the thylakoid as a result of the decreased plastoquinone molecules' oxidation and subsequent transfer of their cytochrome-iron-sulfur complex with electrons.

While the lamella is impervious to neutrons, protons escape as an outcome of the oxidising of water and plastoquinone within the thylakoid, resulting in a greater number of protons within the thylakoid than outside. Alternatively said, the lamella produces a proton gradient. Protons have a positive charge, therefore whenever they travel through the thylakoid lamella during a light reaction, they leave an electrical charge behind.

Reference

Matthew P. Johnson, *photosynthesis Essays in Biochemistry* (2016) 60 255–273 DOI: 10.1042/EBC20160016

Machín, A.; Fontánez, K.; Arango, J.C.; Ortiz, D.; De León, J.; Pinilla, S.; Nicolosi, V.; Petrescu, F.I.; Morant, C.; Márquez, F. One-Dimensional (1D) Nanostructured Materials for Energy Applications. *Materials* **2021**, *14*, 2609.

Wang, X.; Wu, Q.; Ma, H.; Ma, C.; Yu, Z.; Fu, Y.; Dong, X. Fabrication of PbO2 Tipped Co3O4 Nanowires for Efficient Photoelectrochemical Decolorization of Dye (Reactive Brilliant Blue KN-R) Wastewater. *Sol. Energy Mater. Sol. Cells* **2019**, *191*, 381–388.

Kalyanasundaram, K.; Graetzel, M. Artificial Photosynthesis: Biomimetic Approaches to Solar Energy Conversion and Storage. *Curr. Opin. Biotechnol.* **2010**, *21*, 298–310.

Hong, Y.H.; Lee, Y.-M.; Nam, W.; Fukuzumi, S. Molecular Photocatalytic Water Splitting by Mimicking Photosystems I and II. *J. Am. Chem. Soc.* **2022**, *144*, 695–700.

Shen, L.; Yang, X.; An, J.; Zhang, L.; Yang, K.; Deng, Z. Effect of Different Site Trifluoromethylbenzoic Acid Organic Photosensitizer for Dye-sensitized Solar Cells. *ChemistrySelect* **2021**, *6*, 4645–4650.

Sukhova, E.; Sukhov, V. Relation of Photochemical Reflectance Indices Based on Different Wavelengths to the Parameters of Light Reactions in Photosystems I and II in Pea Plants. *Remote Sens.* **2020**, *12*, 1312.

Machín, A.; Cotto, M.; Ducongé, J.; Márquez, F. Artificial Photosynthesis: Current

Advancements and Future Prospects. *Biomimetics* **2023**, *8*, 298. https://doi.org/10.3390/biomimetics8030298



Photosynthesis: Mechanism and Significance

Mahesha K. N^{1*}, Apoorva Guddaraddi², Sakriya, S. G³ and Gamit Upmakumari, C⁴

 ¹Ph. D Research scholar, Department of Vegetable Science, Navsari Agricultural University, Navsari.
²Ph. D Research scholar, Department of Floriculture and Landscape Architecture, Dr. P.D.K.V-Akola.
³Assistant Professor, School of agriculture, P. P. Savani University, Surat, Gujarat
⁴Ph. D Research scholar, Department of Vegetable Science, Navsari Agricultural University, Navsari. Email: maheshkn123456@gmail.com

Abstract

Photosynthesis is a vital process where plants and certain organisms use light to turn it into energy. Green plants transform water, carbon dioxide, and minerals into oxygen and energy-packed compounds. Solar energy becomes glucose through light-dependent reactions and the Calvin cycle. These processes take place in plant cell chloroplasts. Commencing with the assimilation of photons by pigments like chlorophyll, the light-dependent reactions prompt the division of water molecules into oxygen, protons, and electrons. This dual role not only generates oxygen but also replenishes the electron reserves of chlorophyll. Electrons, infused with energy, navigate a protein-laden path along the thylakoid membrane. This journey triggers proton transfer into the thylakoid space, forging a proton gradient. The gradient, a driving force, kindles the synthesis of indispensable energy carriers - ATP and NADPH - priming subsequent stages. Within the chloroplast stroma, the Calvin cycle, or light-independent reactions, transpires. Carbon dioxide initiates the cycle by amalgamating with a five-carbon sugar molecule to create an evanescent compound, swiftly fragmenting into two threecarbon molecules. These molecules, termed 3-phosphoglycerate, undergo reduction via the ATP and NADPH produced during the light-dependent reactions. This transformation vields glyceraldehyde-3-phosphate, a foundational constituent for glucose and other carbohydrates. Some molecules are allocated to regenerate the original carbon dioxide receptor, ribulose-1,5-bisphosphate, unbroken cycle progression. magnitude ensuring The of photosynthesis extends profoundly. First and foremost, it engenders a substantial share of Earth's atmospheric oxygen, nurturing the respiration of multitudinous life forms. Second, as the bedrock of the food chain, photosynthesis orchestrated by plants and algae manufactures organic compounds that confer energy upon consumption. Furthermore, this process holds a pivotal role in energy storage, allowing plants to amass surplus glucose during periods of diminished sunlight. Beyond carbon dioxide reduction and organic matter synthesis, it undertakes the critical responsibility of regulating maintaining carbon and ecosystem vitality. **Besides** its environmental impact, photosynthesis greatly influences human life. It provides oxygen, food, and energy sources like those for farming and biofuels. It demonstrates nature's balance and the collaboration of sunlight and chemicals that sustain life on Earth.

Key words: Calvin Cycle, Chloroplasts, Light-dependent reactions, Oxygen production and Photosynthesis.

Introduction: In the presence of light, green plants have the ability to produce their own sustenance in the form of carbohydrates

utilising carbon dioxide and water. This process is known as photosynthesis. Such mode of nutrition is termed as autotrophic nutrition. Carbohydrates provide energy to the plant and the ones not used are stored in the form of starch. Starch is the reserve food of plants whereas reserve food in case of animals in glycogen. The process of creating organic food in the form of carbohydrates from CO2 and water in the presence of light is known as photosynthesis. The constant photosynthetic performance is regarded to be crucial for healthy plant growth and development, even if numerous physiological, biochemical, and molecular processes together characterise plant productivity (Nguyen et al., 2018; Sharma et al., 2020). A amazing mechanism known as photosynthesis can be found in microorganisms, young growing embryos of aquatic and land plants, and the leaves of green plants (Pan et al., 2012). By starting the process of turning light energy into chemical energy, photosynthesis powers a number of metabolic activities (Chen et al., 2018). Photosynthesis is the life force that keeps our world alive. This is demonstrated by the planet's spectral signature, which is coloured by pigments that capture sunlight (Kiang, 2007). Another indication that life is present on the entire planet is an atmosphere with high oxygen concentrations and enormous numbers of complex organic compounds. Over billions of years, photosynthetic organisms have transformed our planet and the life on it (Cavalier-Smith, 2006). The interdependence of photosynthesis and the development of our planet and the life it harbors make the study of the evolutionary development of photosynthesis an exciting endeavour that connects experimental data and theoretical concepts across scientific disciplines.

Historical background: The historical background is crucial to understanding the journey of discovery that has led to our current understanding of this intricate process. This historical exploration sheds light on the contributions of key scientists who have paved the way for our comprehension of photosynthesis.

The story of photosynthesis begins with the early observations and inquiries of scientists who sought to understand how plants produce energy. In the late 17th century, Jan Baptista van Helmont conducted a famous experiment involving willow trees, realizing that their growth could not be solely attributed to the soil they were planted in. This marked an early recognition of the role of water in plant growth. Joseph Priestley and the Discovery of Oxygen in the mid-18th century, Joseph Priestley conducted groundbreaking experiments that uncovered the vital role of plants in releasing a substance that could support combustion - oxygen. He observed that a mint plant could restore the air that had been "injured" by burning a candle, a remarkable insight into the relationship between plants and the atmosphere. Around the same time, Jan Ingenhousz expanded upon Priestley's findings by demonstrating that light was essential for the process of "injuring" or restoring air in plants. He discerned that the effect of light on plants was pivotal to their role in purifying the air. In the late 19th century, Julius von Sachs contributed significantly to our understanding of photosynthesis. He confirmed that carbon dioxide was necessary for the process and that chlorophyll played a central role in capturing light energy. The mid-20th century witnessed the unravelling of the Calvin cycle, thanks to the pioneering work of Melvin Calvin and his colleagues. This discovery illuminated the intricacies of how carbon dioxide is transformed into carbohydrates, providing a deeper understanding of the mechanics of photosynthesis.

Photosynthetic organisms: More than 180 billion tonnes of organic matter are produced annually by photosynthetic organisms like plants, algae, and some bacteria. The most prevalent organic molecule on the earth, cellulose, which makes up half of this

biomass, is a biopolymer. The majority of plants' cell walls are mostly made up of this carbohydrate macromolecule (Serge and Samain, 2010).

Photosynthetic pigments: Photosynthetic organisms employ pigments as molecular tools to harness light energy during photosynthesis. The eminent pigment in this ensemble is chlorophyll, recognized for imparting plants their characteristic green hue. Chlorophyll holds a remarkable adeptness for capturing sunlight and orchestrating its conversion into energy. Nonetheless, the cast of pigments is not limited to chlorophyll alone. Greenish pigments called chlorophylls have a porphyrin ring in them. It is a molecule with a stable ring structure that allows electrons to freely move all around it. Because electrons may move freely, the ring has the ability to gain or lose electrons quickly, which gives it the potential to give other molecules energised electrons. This is the basic mechanism by which chlorophyll "captures" solar energy. There are various types of chlorophyll, with chlorophyll "a" being the most significant. Through the transfer of its energised electrons to molecules that will produce sugars, this molecule enables photosynthesis. Chlorophyll "a" is present in all plants, algae, and cyanobacteria that photosynthesize. Chlorophyll "b," a different type of chlorophyll, is only found in plants and "green algae." Chlorophyll "C" is the name of a third, more prevalent form of chlorophyll that is only present in dinoflagellates and photosynthetic members of the Chromista. One of the earliest hints that these major groupings were not as closely linked as previously believed came from the discrepancies in the chlorophylls of these groups. These pigments have a wide spectrum of hues and are essential for expanding the sorts of sunlight that can be caught. Each auxiliary pigment excels in absorbing a particular wavelength of sunlight, expanding the range of light energy that the plant may use. These pigments combine their

talents in a symbiotic way to transform sunlight into the tasty energy currency that powers the plant's development and nourishment. This interaction of the colours highlights the synergy that underlies the photosynthetic process and is an example of nature's brilliant craftsmanship.

The site of photosynthesis in plants: In land plants, the leaves are the main organs that produce oxygen through photosynthesis (Figure 1A). In order to expose as much green tissue as possible to light, leaves have evolved. Stomata, which are microscopic apertures in the lower epidermis, control the entry of CO2 into the leaf (Figure 1B). Because the size of the stomatal apertures can alter and is controlled by a pair of guard cells that respond to the turgor pressure (water content) of the leaf, the stomata can open to let CO2 in when the leaf is hydrated. On the other hand, the guard cells reduce turgor pressure and close when water is limited, stopping water from the leaf from evaporating by transpiration. The tiny organelles responsible for photosynthesis are called chloroplasts, and each cell in the green mesophyll tissue of a leaf has around 100 of them. A fourth membrane known as the thylakoid, which in turn encloses a single continuous aqueous area known as the lumen, surrounds the stroma, an aqueous space. Figure 1C, D shows the intricate anatomy of the chloroplast, which contains two colourless exterior membranes (the envelope) that are not involved in photosynthesis.



Fig. 1: Location of the photosynthetic machinery

(A) The Arabidopsis thaliana model plant. (B) A cross-section of a leaf showing the basic structure. Within the cells, chloroplasts are seen as green dots. (C) An image of an Arabidopsis chloroplast taken using an electron microscope. The layered arrangement of the thylakoid membrane is shown in (D), a close-up area of the chloroplast.

Types of Photosynthesis

As classified by Cornelius Van Neil, there exist two primary forms of photosynthesis:

1. Oxygenic Photosynthesis: This process uses water (H_2O) as the electron source and produces oxygen (O_{2}) as a byproduct. It is primarily seen in green plants, algae, and cyanobacteria.

2. Non Oxygenic Photosynthesis: Found in some bacteria, such as Rhodospirillum, Chlorobium, and Chromatium, this kind uses hydrogen sulphide (H_2S) as the electron source without releasing oxygen (O_2) .

Mechanisms of Photosynthesis:

Photosynthesis stands as the primary wellspring of nourishment across our planet, vital not only for sustaining life but also for liberating oxygen into the atmosphere. The portion of the spectrum used in this transforming process, properly known as Photosynthetically Active Radiation (PAR), ranges in wavelength from 400 to 700 nanometers. In the green sections of plants, such as leaves and stems, photosynthesis occurs. Chlorophyll pigments, a specialist light-absorbing green pigment, are found in large quantities in the chloroplasts of the mesophyll cells of leaves. An organelle with two membranes surrounding it is a chloroplast. A challenging redox process is photosynthesis. Water is converted to oxygen during this process, while carbon in carbon dioxide is converted to glucose.

There are two phases to the photosynthetic process:

- 1. Light-dependent or photochemical reaction or Hill reaction
- 2. Light-independent phase or biosynthetic phase or dark reaction

1. Light-dependent or Photochemical Reaction or Hill reaction

The chloroplast's grana is where the light reaction occurs. Photosynthetic pigments in the quantasomes of grana are responsible for capturing light.

(i) The 'light-dependent or photochemical' phase comprises the absorption of light, water splitting, oxygen release, and production of the high-energy chemical intermediates ATP and NADPH.

(ii) Several protein complexes and pigments are found in two different photochemical light-harvesting complexes, LHC inside the photosystem-I (PS-I) and photosystem-II (PS-II).

(iii) Protein- and pigment-associated antenna pigments make up the light-harvesting system, also called the antennae. Chlorophyll-b, xanthophylls, and carotenoids make up the majority of the molecules in the antenna pigments, which absorb light of various wavelengths and transfer their energy to the reaction centre.

(iv) The energy is retained in the reaction centre, which is formed by a single chlorophyll molecule. P700 is the result of the reaction center's absorption peak at 700nm in PS-I, while P680 is the result of the reaction center's absorption peak at 600nm in PS-II.





The transport of electron:

(i) PS-II's chlorophyll absorbs light at a wavelength of 680 nm (red light), becomes stimulated, and triggers an electron leap.

(ii) Through oxidation-reduction or redox reactions, these electrons are captured by a primary electron acceptor and sent downstream to an electron transport system made up of cytochromes.

iii) The electrons are then transmitted to the PS-I pigments, which are then activated by red light with a wavelength of 700 nm and carried downhill once more to convert NADP to NADPH and H.

(iv) Due to its unusual shape, the entire electron transfer scheme that Bendall and Hill developed is referred to as a Z-scheme.



Fig.3: Chloroplast Electron Transport Chain Water splitting or water photolysis

(i) The splitting of water by PS-II, which is found on the inner side of the thylakoid membrane, results in a continual supply of electrons. (ii) The splitting of water into $2H^+$, [O] and electrons. One of the net products of photosynthesis, oxygen, is released as a result. $2H_2O \rightarrow 4H^{++}O_2 + 4e^{-2}$

$2H_2O \rightarrow 4H + +O_2 + 4e^{-2}$

Cyclic and noncyclic photo-phosphorylation

(i) Energy is extracted from oxidizable substances by living things and stored as ATP.

(ii) The process by which cell organelles like the mitochondria and chloroplast produce ATP from ADP and Pi (inorganic phosphate) is known as phosphorylation.

(iii) Photophosphorylation is the process of creating ATP in the presence of light from ADP and inorganic phosphate.

(iv) In noncyclic photophosphorylation, the two photosystems are linked by an electron transport chain to create ATP, NADPH, and H^+ .

(v) In PS-I, the electron is transported across the photosystem and phosphorylation takes place in a cyclic fashion in the stromal lamellae, only synthesising ATP and not NADPH and H^+ .



Fig.4: Cyclic and Noncyclic Photo-Phosphorylation

Chemiosmotic Hypothesis: This explains how the chloroplast produces ATP.

i)A proton gradient forms across the thylakoid membrane, causing ATP production to take place.

ii) The membrane is where the proton or hydrogen ion buildup takes place. Due to the flow of electrons through the photosystems, protons are carried across the membrane from the stroma and divided water molecules are split in the lumen. iii) Protons are taken up by the NADP reductase from the stroma. The NADP reductase converts NADP⁺ to NADPH and H⁺ using electrons from PS-I's acceptor of electrons.

As a result, protons in the stroma of the chloroplast drop in number while those in the lumen (which has a lower pH) accumulate, forming a proton gradient across the thylakoid membrane.

(v) This gradient causes the transport of protons through the membrane to the stroma, which in turn triggers the creation of ATP. (vi) Facilitated proton transport from the lumen to the stroma occurs through the transmembrane channel of the CF_0 of the ATP synthase. (vii) The gradient breakdown causes the CF_1 to face the stroma and change conformation, which releases energy for ATP production.







2. Dark Reaction, Light-Independent Reaction, or Biosynthetic Phase

i) This process is light-independent, meaning that it doesn't require direct sunlight but instead relies on ATP and NADPH as byproducts of the light reaction in addition to CO_2 and H_2O .

ii) Sugars are created when CO2 and H2O are mixed.

iii) Melvin Calvin found that the initial CO_2 fixation product in C_3 plants is a 3-carbon organic acid called 3-phosphoglyceric acid (PGA), using radioactive 14C in algal photosynthesis. The Calvin cycle is a biochemical route that Calvin also described in detail.

(iv) There is a different class of plants (C_4 plants) in which the first stable byproduct of CO2 fixation is oxaloacetic acid, also known as OAA. Ribulose bisphosphate (RuBP), a 5-carbon ketose sugar, is the first CO₂ acceptor molecule.

The Calvin Cycle (C₃ Pathway)

i) Calvin and his coworkers described how the regeneration of RuBP caused the pathway to function in a cyclic fashion.

ii) The Calvin Cycle has three phases: carboxylation, reduction, and regeneration.

iii) The most important step is carboxylation. Fixing CO_2 into an organic intermediate that is stable.

(iv) The enzyme RuBP carboxylase-oxygenase, also known as RuBisCO, uses CO_2 to carboxylate RuBP, culminating in the creation of two 3-PGA.

v) The chain of events known as reduction results in the creation of glucose. involves two ATP molecules for phosphorylation and two NADPH molecules for reduction for each fixed CO_2 molecule. One molecule of glucose is produced from the route after the fixation of six CO_2 molecules and six cycle turns.

(vi) During regeneration, to carry on the cycle, the CO_2 acceptor molecule RuBP is renewed. One ATP is needed for each of these steps in order to phosphorylate RuBP.

(vii) Three ATP and two NADPH molecules are needed for every CO_2 molecule that enters the Calvin cycle.

viii) It takes six cycles of the cycle, 18 ATP, and 12 NADPH to produce one glucose

molecule.



Fig.6: Calvin Cycle

Regulation and Factors Affecting Photosynthesis: Photosynthesis, the intricate energy-making process in plants, is intricately regulated by various factors that influence its pace and efficiency. These factors include:

- 1. **Light Intensity:** The amount of sunlight available directly impacts photosynthesis. A certain level of light is optimal for energy production, but excessive light can actually damage the plant's photosynthetic machinery. This is why plants have evolved mechanisms to protect themselves from excessive light, such as adjusting the opening and closing of their stomata.
- 2. **Temperature:** Photosynthesis is sensitive to temperature variations. As temperature increases, the rate of photosynthesis generally rises. However, extreme temperatures can hinder enzyme activity and disrupt the delicate balance of reactions. Plants have evolved to thrive within specific temperature ranges that optimize their photosynthetic performance.
- 3. **Carbon Dioxide Concentration:** Carbon dioxide is a vital ingredient in photosynthesis. Higher levels of CO2 can accelerate the process, but only up to a point. Beyond a certain threshold, further increases in CO2 concentration don't

significantly enhance photosynthesis. This balance is essential for maintaining efficient carbon fixation.

- 4. Water Availability: Adequate water supply is crucial for the transport of nutrients and the maintenance of turgor pressure within plant cells. Water scarcity can lead to the closure of stomata, reducing the intake of carbon dioxide, and consequently, hampering photosynthesis.
- 5. **Nutrient Availability:** Essential nutrients like nitrogen, phosphorus, and potassium play roles in various stages of photosynthesis. A deficiency in any of these nutrients can limit the plant's ability to carry out photosynthesis effectively.
- 6. **Stomatal Regulation:** Stomata are tiny openings on leaves that facilitate gas exchange. Plants can regulate the opening and closing of stomata to control the influx of carbon dioxide and the release of oxygen and water vapor. This adaptation helps plants optimize photosynthesis while conserving water.
- 7. **Environmental Stressors:** External stressors like pollutants, pathogens, and extreme weather conditions can impact photosynthesis. Plants may respond by adjusting their photosynthetic machinery or producing protective compounds.

Understanding these regulatory factors provides insights into how plants optimize their energy production under varying conditions. Just like a conductor orchestrating a symphony, plants delicately balance these factors to ensure efficient and sustainable photosynthesis, contributing to their growth, survival, and the overall health of ecosystems.

Significance of photosynthesis:

1. **Oxygen Production:** We begin by unraveling the vital role of photosynthesis in oxygen generation. This process not only provides the oxygen we breathe but also shapes the composition of Earth's atmosphere, influencing the very air we depend on.

- 2. Energy Storage and Biomass Production: Moving forward, we explore how photosynthesis serves as the ultimate energy storage mechanism. We delve into the intricate process by which plants convert light into carbohydrates, effectively storing energy in the form of biomass a foundation for plant growth and ecosystem vitality.
- 3. **Role in Food Chains:** Venturing deeper, we uncover how photosynthesis forms the bedrock of food chains in both land and water ecosystems. By producing the organic compounds that sustain life across trophic levels, photosynthesis intricately supports the delicate balance of entire ecosystems.
- 4. **Carbon Dioxide Reduction and Climate Regulation:** We then dive into the crucial connection between photosynthesis and climate regulation. Through the absorption of carbon dioxide, photosynthesis plays a pivotal role in reducing greenhouse gases, thereby aiding in mitigating climate change's adverse effects.
- 5. **Human Applications and Industries:** Our journey extends to the practical applications of photosynthesis in human endeavors. We explore how this process powers agriculture, serves as a source for biofuel production, and contributes to various industries, showcasing its far-reaching influence.
- 6. **Evolution and Earth's History:** Our exploration culminates in understanding how photosynthesis has sculpted Earth's evolutionary trajectory. We delve into its transformative impact on our planet's atmosphere over eons, driving the development of life forms that shape the world we inhabit today.

Future Prospects of Photosynthesis: The exciting possibilities and potential pathways that lie ahead in the realm of photosynthesis, envisioning how this fundamental process could shape our world in the coming years.

- 1. Enhancing Agricultural Sustainability: Our journey begins with a focus on agriculture's future. We delve into the realm of precision farming and genetic advancements, discussing how a deeper understanding of photosynthesis can lead to crops that are not only more resilient but also produce higher yields with reduced resource inputs.
- 2. **Solar Fuel Revolution:** Moving forward, we venture into the realm of energy. We unravel the promising potential of photosynthesis in producing clean, renewable energy. We explore innovative technologies that harness sunlight to create fuels, potentially revolutionizing our energy landscape and reducing our carbon footprint.
- 3. **Climate Change Mitigation:** The chapter takes a crucial turn as we examine how photosynthesis can serve as a powerful tool against climate change. We discuss the potential of large-scale reforestation efforts and the role of photosynthetic organisms in capturing carbon dioxide, offering hope in the fight against global warming.
- 4. **Biotechnological Breakthroughs:** Our exploration extends to biotechnology's future, where the genetic manipulation of photosynthetic pathways holds promise. We discuss the potential to engineer organisms for enhanced photosynthesis, creating organisms that can thrive in extreme conditions or even support life in space.
- 5. **Synthetic Biology and Beyond:** We conclude by delving into the realm of synthetic biology. We explore the realm of creating entirely new photosynthetic organisms with tailored capabilities, envisioning a future where we can engineer biological systems for specific applications, from environmental remediation to food production.

Conclusion: Significant and intricately woven into the web of life on Earth is photosynthesis. It is more than just a biological process; it is a crucial component that supports ecosystems, produces oxygen, feeds the food chain, and controls our climate. Every living thing, from the smallest bacterium to the tallest trees, is affected by its effects. However, when we delve more deeply into the principles underlying photosynthesis, we see opportunities for advancement via cutting-edge techniques. One of the methods that has the potential to increase photosynthetic efficiency is biotechnology. Other tools include genetic engineering and precision breeding. We can improve important processes by fiddling with plant genetics, which will raise food yields and improve resource utilisation. Utilising computational models and artificial intelligence, we can forecast and alter photosynthetic processes to achieve the best results. Additionally, studying natural variances and adaptations in plants can reveal characteristics that can be incorporated to support photosynthesis under various circumstances. The voyage through the significance of photosynthesis and its enhancement techniques, in conclusion, is a monument to human ingenuity and nature's tenacity. It is a tale of environmental and scientific cooperation motivated by the search for long-term fixes to a world that is changing quickly. The advances we make in perfecting photosynthesis will not only transform agriculture and address issues of global food security, but also help us fight climate change. We hold the power to sculpt a more resilient and peaceful future for our planet and all of its people as we stand at the confluence of science and nature.

References:

- Cavalier-Smith, T. (2006). Cell evolution and Earth history: stasis and revolution. *Philosophical Transactions of the Royal Society of London*, 36(1):969–1006.
- Chen, Y., Zhou, B., Li, J., Tang, H., Tang, J., and Yang, Z. (2018). Formation and change of chloroplast-located plant metabolites in

response to light conditions. *International Journal of Molecular Sciences*, 19:654.

- Kiang, N., Siefert, J., Govindjee and Blankenship, R.E. (2007). Spectral signatures of photosynthesis. Review of earth organisms. *Astrobiology*, 7:222–251.
- Lovelock, J.E. (1965). A physical basis for life detection experiments. *Nature*, 207:568–570.
- Nguyen, H. C., Lin, K. H., Hsiung, T. C., Huang, M. Y., Yang, C. M., Weng, J. H., *et al.* (2018). Biochemical and physiological characteristics of photosynthesis in plants of two calathea species. *International Journal of Molecular Sciences*, 19:704.
- Pan, J., Lin, S., and Woodbury, N. W. (2012). Bacteriochlorophyll excitedstate quenching pathways in bacterial reaction centers with the primary donor oxidized. *Journal of Physical Chemistry*, 116: 2014–2022.
- Serge, P. and Samain, D. (2010) Structure and engineering of celluloses. Advances Carbohydrate Chemistry Biochemistry, 64: 25-116.
- Sharma, A., Kumar, V., Shahzad, B., Ramakrishnan, M., Singh Sidhu, G. P., Bali, A. S., et al. (2020). Photosynthetic response of plants under different abiotic stresses: a review. Journal of Plant Growth Regulation, 39: 509–531.



Anatomy of Seed C. M. Godhani and T. H. Borkhatariya

Ph.D. Scholar (Agri.), Department of Genetics and Plant Breeding, College of Agriculture, Junagadh Agricultural University, Junagadh-362 001 (Gujarat), India. Mail: godhanichirag13@gmail.com

Abstract

A seed's anatomy is an elaborate and complicated structure that represents the possibility of new life. The basic building blocks of plant reproduction, seeds contain developing plants with the nutrients they need to grow. For developing plants, seeds act as storage containers for energy and as protective coverings. The embryo, endosperm, along seed coat are the three fundamental components of a seed. The cotyledons, which are the embryonic leaves, the shoot meristem, the root meristem, and other early plant components are all found in the embryo. The endosperm functions as a nutritional tissue, supplying the growing embryo with vital proteins, and carbohydrates nutrients like lipids, during germination. The seed coat serves as a barrier around these essential parts, preserving the internal components from physical harm, infections, and stresses from the environment. The testa, or the outermost layer of the seed, facilitates gas and water exchange while protecting its insides from outside influences. The anatomy of seeds reveals the processes by which plants spread out, establish, and reproduce themselves throughout a range of ecosystems, which is crucial for plant research as well as agricultural practices, restoration of the environment, and food security. Enhancing agricultural yields, resistance to diseases, and stress tolerance are

additional benefits of modifying seed characteristics. Additionally, research on seed morphology provides insight into plant evolution, dispersal methods, and adaption techniques in a variety of habitats. As a whole, the anatomy of a seed reflects a remarkable interaction of parts that support the survival of plants. The seed has directions for growth and survival, from its complex internal parts to its outside protective coats. Our understanding of plant biology is improved by uncovering the mysteries of seed anatomy, which also opens up interesting opportunities for sustainable agriculture as well as environmental protection.

Keywords: Seed, Embryo, Endosperm, Germination and Cotyledon **Introduction**:

Seeds are cryptic reservoirs of potential that represent the potential of new beginnings in the complex world of plant life. The anatomy of a seed reveals a fascinating story about life's strength and creativity that is hidden behind its simple material. This chapter sets off on a journey of exploration into these amazing structures' inner workings, removing the layers and revealing the complex configurations that guide how they develop and grow. The anatomy of a seed is an appreciation of nature's production, from the protective covering that shields against difficulty to the primordial structures that contain the blueprints for future growth. We go deeply into the principles of germination and the change of a dormant creature into a healthy living thing by examining the cellular construction, the storage systems that maintain life throughout dormancy, and the complex interplay of hormones, nutrients, and genetic information. This chapter considers the broader consequences of seed morphology beyond its biological value, touching on issues like seed banking, farming methods, and ecological preservation.

Understanding the complex makeup of seeds provides us with insights into both the workings of life and the interconnection of the natural world. We are encouraged to think about the similarities between these miniature representations of life and the promise that lives within each of us as we solve the mysteries of a
seed's anatomy. The hidden potential we carry shapes our own routes, just as a seed has the capacity to create forests and landscapes. Let's start our exploration journey by breaking down the components of a seed's anatomy to better understand the complex mechanics underlying the never-ending cycle of growth, change, and renewal.

What is Seed?

A seed is a crucial and complex reproductive component of plants that contains an embryonic plant, as well as a supply of nutrients, all enclosed in a seed coat for protection. Many different plant species depend on seeds for reproduction and survival, which allows them to continue existing and adapt to changing environments (Loewer, 2005).

Significance of seeds in the plant life cycle:

In the process of a plant's life cycle, seeds play a crucial role in the survival, spread, and reproduction of the organism. Beyond basic reproduction, their importance includes ecological balance, genetic variety, and human nutrition. The following are the primary aspects of their significance:

- Genetic Continuity: The genetic code of the ancestor plant is carried by the seed. Genetic material comprising both the male and female ancestors is incorporated into the seed through sexual reproduction. This ensures the passing down of advantageous characteristics, adaptations, as well as evolutionary changes, helping in the long-term survival and adaption of species.
- Propagation: Most plants reproduce primarily through the development of seeds. They comprise a young, concealed plant embryo that, given suitable conditions, can grow into a mature plant. Because of their susceptibility to dormancy, plants are better able to adapt the conditions of their growth, increasing the probability that they will survive.
- Nutrient Reserve: Essential nutrients needed for the early phases of growth are stored in seeds. Until the developing embryo can develop its own photosynthetic ability and

absorb nutrients from the environment, these stores supply nutrition. Depending on stored nutrients allows seedlings throughout their early growth phases to adjust to unfavourable environments.

- Dispersal: For plants to spread, seeds are essential. Seeds are dispersed over wider areas by a number of methods, including wind, water, living things, and even selfpropulsion. By reducing competition between individuals who are closely related and increasing genetic diversity, this dispersal technique helps plants establish themselves in new environments.
- Adaptation: Plants depend on seeds to survive in unfavourable situations like drought or extremely high temperatures. Some plants develop seeds that can withstand these kinds of stresses effectively, enabling the seeds to survive difficult times and then germinate when the situation becomes more favourable. This form of adaptation increases the possibility of the species surviving over a long period of time.
- Genetic Variation: In order to sustain genetic diversity among plant populations, seeds are essential. Genetic recombination caused by sexual reproduction during seed development produces unique trait combinations. The species must have this diversity in order to adapt to the changing environment, diseases, and other difficulties.
- Ecological Importance: For many species, seeds are an essential source of food, which helps the ecosystem maintain. Seeds contribute to the restoration of plant communities from difficulties, which promotes the stability of ecosystems. They make sure that vegetation regenerates, minimising soil erosion, promoting nutrient cycling, and supporting the complex interactions that occur within ecosystems. They draw herbivores, which promote pollination and the spread of seeds. Additionally, seed

banks can act as a storehouse for protecting plant species, particularly those in danger of extinction.

Overview of Seed Diversity Across Plant Species:

In the kingdom of plants, seeds are extraordinary carriers that hold the potential for new life. The size, shape, construction, and adaption methods of seeds vary remarkably among different plant species. Because of their diversity, plants have been able to survive in various kinds of habitats around the planet for millions of years.

- ✓ Seed Size and Shape: diverse plant species may have significantly diverse seed sizes and shapes. Certain kinds of seeds, particularly the seeds of orchids, are very small they only have a diameter of a few micrometres. On the opposite end, coconut palm seeds are among the biggest, growing to a diameter of up to 12 inches. Complex and specialized architecture adapted to varied ways of dissemination, may vary from the common circular and oval forms of seeds.
- ✓ Seed Dispersal Mechanisms: Plants have developed complex mechanisms for dispersing their seeds and maintaining their survival in a wide range of conditions. As observed in dandelions as well as maple trees, some seeds have features like wings, hooks, and hairs that facilitate wind dispersion. Others, such as burdock, spread more readily because their hooked spines cling to the fur of passing animals. Water lily seeds have air pockets that make them float, enabling water currents to carry them.
- ✓ Adaptations for Germination: The variety of seeds includes germination-supportive modifications. Some seeds need particular conditions in order to emerge from dormancy and begin to grow, such as changes in temperature or exposure to fire. For instance, several pine tree species contain serotinous cones that only open to release seeds under a forest fire's high temperatures. On the

other hand, arid plants such the Joshua tree are growing seeds that only grow when there is sufficient rainfall.

- ✓ Nutrient Storage: The developing embryo can store nutrients in seeds. Various species of plants have developed many kinds of mechanisms for storing nutrients in seeds. For instance, the protein-rich cotyledons of legumes like beans and peas are used to store nutrition. In contrast, the endosperm tissue enclosing the embryo in endospermic seeds, such as those found in cereals like wheat and maize, stores nutrition.
- ✓ Seed Coat Diversity: The Testa, or seed coat, which is the outer layer of a seed, is remarkably diverse. The seed coat can be thick and stiff in some situations, providing a defense against infections or physical harm. Other seeds have thin, porous coverings that allow them to absorb water for germination more quickly. Certain seed coats also include chemicals that keep infections and herbivores away, increasing the likelihood that the seed will survive.

Types of Seeds

- Based on Source:
 - (a) Vegetable Seeds: These seeds are utilized for establishing new plants and come from a variety of vegetables. Cucumber seeds, tomato seeds and carrot seeds are a few examples.
 - (b) Fruit Seeds: These seeds can be utilized to generate new fruit-bearing plants; they are discovered inside fruits. Apple seeds, watermelon seeds, and avocado seeds are a few examples.
 - (c) Flower Seeds: New flowers are propagated using flower seeds, which are produced by a number of flowering plants. Sunflower, rose, and marigold seeds are a few examples.
- Based on Cotyledon:

- (a) Monocotyledonous Seeds: In the embryo of these seeds, there is only one cotyledon (seed leaf). Examples include corn (maize), rice, and wheat.
- (b) **Dicotyledonous Seeds**: The embryo of these seeds has two cotyledons. Examples include beans, peas, and sunflowers. The difference between monocot and dicot seeds is given in Table 1.
- **Based on Storage Food**:
 - (a) Endospermic Seeds: These seeds contain endosperm, a tissue rich in nutrients that feeds the growing embryo. Examples include corn (maize) and wheat.
 - (b) Non-Endospermic Seeds: These seeds contain endosperm, a tissue rich in nutrients that feeds the growing embryo. Examples include beans, peas, and sunflowers. The difference between endospermic seeds and non-endospermic seeds is given in Table 2.
 - (c) **Perispermic Seeds**: The perisperm, a specialized tissue distinct from the embryo, is the place where these seeds store food. Examples include black pepper and coffee.
- **Based on Dispersal Mechanism:**
 - (a) Wind-Dispersed Seeds: These seeds are dispersed through wind from one place to new areas. Examples include dandelion seeds and maple tree seeds.
 - (b) Animal-Dispersed Seeds: These seeds have features that allow animals to transport them to other places. Examples include burdock seeds and acorns.
 - (c) Water-Dispersed Seeds: Because of their ability to float on water, these seeds can be transferred from one place to new locations. Examples include coconut seeds and water lily seeds.

Based on Nutritional Value:

- (a) Edible Seeds: These seeds are healthy to eat and tend to have a lot of nutrition. Examples include chia seeds, flaxseeds, and sunflower seeds.
- (b) Inedible Seeds: These seeds contain poisons or other elements that make them unfit for human consumption. Examples include castor beans (a source of ricin) and apple seeds (contain cyanide in small amounts).
- (c) Oil Seeds: These seeds are frequently utilized for oil extraction because they contain a lot of oil. Examples include sesame seeds, soybeans, and canola seeds.

> Based on Germination Requirements:

- (a) Light-Dependent Seeds: For these germinating seeds, light is necessary. Examples include lettuce seeds and petunia seeds.
- (b) **Dark-Dependent Seeds**: For these seeds to germinate, they must be in darkness. Examples include radish seeds and most beans.
- (c) Cold-Stratification Seeds: Before these seeds may germinate, they require a period of freezing temperatures. Examples include many perennial flowers and certain tree species.

External Seed Structure:

External seed structures are visible to the normal eye features and physical properties of seeds. These structures are essential for the germination, protection, and spread of seeds. There are several essential components in the exterior seed structure (Shown in Figure 1):

Seed Coat (Testa): The seed's outermost layer of protection is known as the seed coat. It acts as a defense against infections, mechanical harm, and water loss. The seed coat can have different textures, colours, and patterns and normally consists of a few layers.

- Hilum: The seed coat has an indentation called a hilum that marks the spot where the seed became attached to its fruit or reproductive wall. It facilitates nutrient transport during seed development and acts as the point of connection for the seed inside the fruit.
- Micropyle: The micropyle is a tiny hole near the hilum in the seed coat. It facilitates the passage of the growing radicle (embryonic root) and permits water to get into the seed during germination.
- **Raphe**: The raphe is a rib or protrusion found along the entire length of some seeds, especially those of some flowering plants. It demonstrates how the seed coat and the growing embryo are connected.
- Seed Shape and Size: Seeds come in a variety of sizes and forms that can be specific to various dissemination methods. For instance, seeds that are disseminated by wind frequently have small sizes and wind-conveying features like wings or bristles. Larger seeds might be made to be dispersed by animals.
- Appendages: Specific appendages on some seeds facilitate their dissemination. Burdock seeds, for instance, contain hooked bristles that cling to animal fur and facilitate transportation to new places.
- Aril: An aril is an extra seed coat that emerges from the funiculus or seed stalk and often has a fleshy or colourful appearance. Animals that aid in seed dispersal are drawn to it. One of the best examples is the yew seed's aril.
- Seed Colour and Texture: The seed coat can have a wide range of colours and textures. Certain seeds may act as messages to potential dispersers of seeds because of their unusual patterns or colours.
- Pappus: Numerous species belonging to the Asteraceae (daisy) family have pappus structures in their seeds. It has bristles, hairs, or scaling that help spread the wind.

Elaiosome: A fatty, protein-rich protrusion called an elaiosome is found in the seeds of some plants, and it frequently attracts ants. The ants transport the seeds to their nests where they eat the elaiosome before allowing the seed to sprout in a protected area.

Internal Seed Structure:

The reproductive organ that gymnosperms and angiosperms (flowering plants) produce is called a seed. It acts as a container for the plant embryo, shielding it and giving it as necessary nutrients it needs to germinate and begin its early growth. Although the interior makeup of seeds differs depending on the species of plant, most seeds share some elements and layers (Shown in Figure 1):

- Endosperm: The endosperm is a nutritional tissue that envelops the embryo in many plant species. During germination, it supplies vital nutrients to the growing embryo. There is little or no endosperm inside the mature seeds of some species because the endosperm is taken in by the embryo throughout seed development.
- Embryo: The young, immature plant inside the seed is called the embryo. It has several important components:
 - **i. Radicle**: The developing root that, after germination, will produce the main root.
 - **ii. Plumule**: The developing embryonic stalk from which the plant's above-ground components will emerge.
- Cotyledons: These are the nutrient-storing seed leaves, which occasionally act as the developing plant's earliest structures for photosynthetic growth. Different plant species can have different numbers and sizes of cotyledons, which can be used to categorise plants as monocots or dicots.
- Seed Axis: These are the nutrient-storing seed leaves, which occasionally act as the developing plant's earliest structures for photosynthetic growth. Different plant species can have different numbers and sizes of cotyledons,

which can be used to categorise plants as monocots or dicots.

- Cotyledonary Node: The embryo is now joined to the seed coat at this location. It is situated where the cotyledons and embryonic axis meet.
- Cotyledonary Bud: Some seeds develop a little bud in the cotyledon axil. When the seed sprouts and grows, this bud can produce new branches and leaves.



Figure 1: Shows the structure of seeds from both dicot and monocot seeds. Dicots have two cotyledons (see left). The scutellum, a single cotyledon seen in monocots like maize (right), transports nutrients to the developing embryo. In both monocot and dicot embryos, the leaves, stem, and roots are formed by the plumule, hypocotyl, and radicle, respectively. The cotyledon(s), but not the plumule or the radicle, are not part of the embryonic axis. (Source: Lumen Learning, 2023)

Germination of Seeds:

The mechanism by which an embryo sprouts and develops into an embryonic plant is called germination. It entails the stimulation of the embryo inside the seed, resulting in the radicle's (the embryo's root) emergence and the development of the shoot system afterward. There are various germination processes, each adapted to the conditions under which the vegetative species developed. The following list of germination kinds includes appropriate examples:

- (a) Epigeal Germination: The cotyledons (seed leaves) are propelled above the soil surface in seedling emergence in this kind of germination. The cotyledons extend together with the epicotyl, which is the area of the embryo above them. Sunflowers (*Helianthus annuus*) and beans (*Phaseolus vulgaris*) are two examples of plants that germinate epigeal type of germination.
- (b) Hypogeal Germination: The cotyledons stay below the soil's surface during hypogeal germination. The cotyledons, which act as reserves of energy for the developing seedling, remain inside the seed coat as the epicotyl climbs upward. Chickpeas (*Cicer arietinum*) and peas (*Pisum sativum*) are two examples of plants that germinate in the hypogeal type of germination.
- (c) Vivipary: When seeds grow while still connected to the parent plant, vivipary results. In mangroves and some aquatic plants, this is typical. For instance, the red mangrove (*Rhizophora spp.*) releases seeds that sprout while still on the tree, and once the young seedlings are adequately developed, they fall into the water.
- (d) Intermediate Germination: There can sometimes be a combination of epigeal with hypogeal germination. While some of the cotyledons remain below the soil's surface, others partially rise above it. An example of a plant with intermediate germination is maize (*Zea mays*).
- (e) **Cryptobiosis**: During cryptobiotic germination, seeds remain unconscious in the soil for a long time before specific environmental factors induce germination. Desert plants, such as the "resurrection plant" (*Selaginella lepidophylla*), provide an example. These plants may survive for many years without water till sprouting when water is available.
- (f) Aerial Germination: When the process of germination occurs above the ground, it is called aerial germination. This is typical of several epiphytic plants, which grow on different plants

without consuming their nutrition. For instance, airborne germination is common in *orchids*.

- (g) Epiphytic Germination: Seeds can germinate epiphytically, or on the outside of other plants. Such germination is common in epiphytic orchids like the vanilla orchid (*Vanilla planifolia*).
- (h) Seedling Growth Response: Specific environmental stimuli can trigger specialized germination responses in some plants. For example, the seeds of fire-dependent plants, such as certain kinds of *eucalyptus*, need to be exposed to smoke or heat in order to germinate.

Factors Affecting Seed Germination:

The successful development of an embryo from seed is the result of a complex process called seed germination, which is regulated by a number of variables that interact. Some of the main elements influencing seed germination are provided below:

✓ Water Availability: Since it starts metabolic processes inside the seed, enough water intake is necessary for seed germination. Some seeds need a certain amount of water to start germination and break out of dormancy.

<u>Example</u>: *Cacti* and other desert plants have seeds that need a lot of water to break their dormancy and germinate. This behaviour is common in the saguaro cactus (*Carnegiea gigantea*).

✓ Temperature: For seeds to germinate, a certain temperature must be attained. Temperatures that are too warm or too low may prevent or delay germination.

Example: Tomato seeds (*Solanum lycopersicum*) require temperatures between 20-30°C for optimal germination.

✓ Light: Some seeds, especially those that are sensitive to light, may be affected by light and their ability to germinate. Depending on the species, light can either encourage or prevent germination.

<u>Example</u>: Lettuce seeds (*Lactuca sativa*) are known to have light-dependent germination. They germinate better under light conditions.

✓ Oxygen Availability: For aerobic respiration to start during germination, oxygen is necessary. Low oxygen concentrations can cause poor sprouting or seedling mortality.

<u>Example</u>: Rice (*Oryza sativa*) seeds need well-aerated soil for successful germination. Waterlogged conditions can reduce oxygen availability and lead to poor germination.

✓ Seed Coat Permeability: Germination may be affected by the seed coat's porosity to gases and water. Some seeds need scarification (mechanical and chemical treatments) to help water penetrate their tough or impenetrable seed coverings.

Example: Lupine seeds (*Lupinus spp.*) have hard seed coats that can be scarified by abrasion or acid treatment to increase germination.

- ✓ Dormancy: Under unfavourable conditions, seeds cannot germinate due to dormancy mechanisms. Different types of dormancies, including physiological dormancy, embryonic dormancy, and seed coat hibernation, might affect germination.
- ✓ <u>Example</u>: Apple seeds (*Malus domestica*) have embryo dormancy and require a period of cold stratification to break the dormancy and germinate.

Characteristic	Monocot Seeds	Dicot Seeds		
Seed Leaves	Usually one cotyledon	Usually two		
(Cotyledons)		cotyledons		
Vein Pattern in	Parallel veins	Branched veins		
Cotyledons				
Embryo	Embryo with single	Embryo with		
	cotyledon (scutellum)	two cotyledons		
Endosperm	Endosperm often present	Endosperm		
		usually absent		
Seed Coat	Single-layered seed coat	Double-layered		
	(testa)	seed coat (testa		
		and tegmen)		

Root Anatomy	Adventitious root system	Taproot system		
	often seen	usually present		
Hypocotyl	Short hypocotyl	Elongated		
		hypocotyl		
Cotyledon	Stores minimal nutrients	Stores		
Nutrient		significant		
Storage		nutrients		
Germination	Usually hypogeal	Can be hypogeal		
	(cotyledon remains	or epigeal		
	below ground)	(cotyledons		
		emerge)		

 Table 1: Difference between monocot seeds and dicot seeds

Table 2: Difference between	endospermic	seeds an	ıd non-
endospermic seeds			

Chanastanistia	Endospermic	Non-Endospermic	
Characteristic	Seeds	Seeds	
Seed Structure	Contains three	Contains two	
	distinct parts:	distinct parts:	
	embryo,	embryo and seed	
	endosperm, and	coat	
	seed coat		
Nutrient Storage	Nutrients stored in	Nutrients stored in	
	endosperm,	cotyledons	
	providing	(embryonic leaves)	
	nourishment to		
	embryo		
Embryo	Embryo develops	Embryo primarily	
Development	while being	relies on cotyledon	
	nourished by the	reserves for growth	
	endosperm		
Examples	Maize (corn),	Bean, pea,	
	coconut, barley	sunflower, tomato	

Conclusion:

In conclusion, studying the complex architecture of seeds offers a profound comprehension of the amazing features that aid in plant survival and reproduction. Researchers learn more about the adaption tactics that have developed through thousands of years by thoroughly examining the various parts of seeds, such as the outer layer of the seed, embryo, and endosperm. The sensitive internal structures are protected from potential injury and external pressure by the seed coat, which serves as a barrier. The embryo, which consists of the embryonic root and shoot (plumule and radicle), holds the potential for further development and growth that will be sparked by the appropriate environmental stimuli. Additionally, the developing plant is fed by the endosperm, a tissue rich in nutrients, during its initial phases of growth and until it can generate roots. The importance of these seed elements in relation to ecological interactions, farming methods, and biodiversity preservation. Crop breeding can benefit from research on seed anatomy by developing seed quality, rate of germination, and overall plant productivity. Understanding the differences in seed structure among various plant species also helps us better comprehend ecological tactics and evolutionary trends, revealing how plants have evolved to deal with a wide range of habitats and environmental constraints. The architecture of seeds continues to be a key topic in botanical research as we make strides in understanding both the historical and contemporary mechanics of plant reproduction as well as the promise for future developments in horticulture, agriculture, and environmental restoration. Scientists and professionals can use this knowledge to ensure the production of nutritious food, the wellness of ecosystems, and the continued survival of the natural environment by figuring out the precise features of seed anatomy.

References:

- Bailey-Serres, J. and Voesenek, L. (2008). Flooding stress: acclimations and genetic diversity. *Annual Review of Plant Biology*, 59: 313-339.
- Bewley, J. D. and Black, M. (2013). *Seeds: physiology of development and germination*. Springer Science & Business Media.

- Bewley, J. D.; Bradford, K. and Hilhorst, H. (2012). *Seeds: physiology of development, germination, and dormancy*. Springer Science & Business Media.
- Bewley, J. D.; Bradford, K. and Hilhorst, H. (2012). Seeds: physiology of development, germination and dormancy. Springer Science & Business Media.
- Esau, K. (1965). Plant anatomy. *Plant Anatomy.*, (2nd Edition).
- Fahn, A. and Werker, E. (1972). Anatomical mechanisms of seed dispersal. Seed biology: importance, development, and germination, 1: 151-221.
- Fenner, M. (2000). Seeds: the ecology of regeneration in plant communities. CABI publishing.
- Lang, G. A.; Early, J. D.; Martin, G. C. and Darnell, R. L. (1987). Endo-, para-, and ecodormancy: physiological terminology and classification for dormancy research. *HortScience*, 22(3): 371-377.
- Lim, T. K. (2012). *Edible medicinal and non-medicinal plants* (Vol. 1, pp. 656-687). Dordrecht, The Netherlands: Springer.
- Loewer, P. (2005). *Seeds: the definitive guide to growing, history and lore.* Timber Press.
- Lumen Learning (2023) Biology for Majors II. Sourse: <u>https://courses.lumenlearning.com/wm-</u>biology2/chapter/development-seeds-and-fruit/.
- Morgan, P. W. (1979). A Preliminary Research Plan for Development of a Photosynthetic Link in a Closed Ecological Life Support System (No. NASA-CR-160399).
- Pearson, T.; Burslem, D. and Swaine, M. (1999). Seeds: Ecology, biogeography, and evolution of dormancy and germination. *Journal of Tropical Ecology*, 15(4): 543-544.
- Street, H. E. and Öpik, H. (1984). *The physiology of flowering plants: their growth and development*. Edward Arnold Ltd.

10

Plant Defense Mechanisms in Plant Mahima Choudhary¹ and Sanjay Kumar¹ ¹Department of Plant Pathology, Rajmata Vijayaraje Scindia

Krishi Vishwa Vidyalaya, Gwalior, Madhya Pradesh, India E-mail: <u>mahimachoudhary408@gmail.com</u>

Abstract

Various morphological, biochemical, and molecular defense mechanisms are used by plants to safeguard against biotic and abiotic stressors. Numerous, highly dynamic biochemical systems of self-defense against plant diseases involve both direct and indirect responses. Different protective layers are formed, natural apertures are altered, secondary metabolites are produced, and occasionally primary metabolites are produced. These physical, morphological, and biochemical processes are known to provide some defense against pathogens and other biotic and abiotic pressures. There are two types of morphological and biochemical processes: those that are already present, that is specific morphological and biochemical characteristics in plants, as well as those that form afterwards as a result of disease attacks. Two types of resistance-systemic acquired resistance and induced systemic resistance-play a critical role in plants' capacity to protect themselves against numerous pathogenic assaults, In addition to secondary metabolite synthesis and other defense mechanisms.

Key word: physical, morphological, biochemical, molecular defense mechanisms

Introduction

Because biotic and abiotic forces differ enough for plants, they respond by developing sophisticated defensive systems. The constant attack of insects and microbial diseases on plants is one example of this type of stress. It is believed that plant pathogenic bacteria formerly drastically decreased the number of trees and plants. In such circumstances, the defense system might be engaged or it might produce damaging secondary metabolites. Plants detect abiotic stress conditions, which then activate the transcriptional or regulatory machinery to provide the necessary response. One of these techniques is the generation of secondary metabolites, which takes place in plant cells through metabolic pathways descended from the primary metabolic pathways. Plants frequently produce these metabolites in stressful (abiotic and/or biotic) conditions, with various signaling molecules acting as the primary mediators (Hussein et al., 2019). The plant kingdom has about 100,000 secondary metabolites that are connected to various chemical groups [Li et al., 2020]. Based on how they are made, these substances can be put into three main groups: phenolic compounds (such as flavonoids and phenyl propanoids), nitrogen-containing substances (such as amines, cyanogenic glycosides, alkaloids, and glucosinolates), and terpenes (such as volatile components, essential oils, and isoprenoids). Along with chemical defenses, plants also have other recognized defenses against pathogenic onslaughts. These are a combination of induced and acquired resistances. Both involve resistance to various signaling mechanisms.

In this chapter, we give a thorough overview of the various defenses employed by plant systems to fend off pathogenic attacks and give a brief description of how these defenses work.

Mechanisms of plant defense

Plants are the most abundant and prominent class of autotrophic creatures on Earth. The copious organic substance of these plants serves as food for all heterotrophic creatures, including animals, insects, and microbes. Pathogens of all kinds are exposed to plants. Through their aggressive actions, plant pathogens attack their hosts and harm them [Kombrink *et al.*, 1995]. Plants have developed

defense systems over time, adapted from different phyla, to protect themselves and stop prospective diseases from penetrating and colonizing, following the pathogens' evolution. Two components of a plant's active immune system operate as the primary regulators of perception and defense. The plant disease resistance gene (R) produces the R proteins. By identifying the presence of avirulence proteins or effectors produced by the pathogen, this protein controls plant defense. According to Flor (1971; Dodds et al., 2010), the effector or avr protein can bind to a target either directly or indirectly by altering the target [Flor, 1971; Dodds et al., 2010]. A new model (Mukhtar et al., 2011) suggests that the R proteins may be able to defend critical cellular hubs from many diseases, which might make them shared targets for effectors. Next on the list of the immune system's functional components are transmembrane pattern recognition receptors, which may nevertheless recognize the pathogen's molecular patterns (MAMPs) and respond to the invader. The majority of pathogens in a class required for disease persistence have MAMPs, including flagellin in bacteria and chitin in fungi (Voigt, 2014). A few of the defense mechanisms that are activated in plants include the production of phytoalexins with antibacterial potential, alterations to plant cell walls, especially the papillae deposition laden with the cell wall polymer (1,3)-glucan, callose, and the creation of enzymes with the capacity to disintegrate pathogen cell walls. According to research, thickening of cell walls acts as a physical barrier to prevent pathogens from entering the area where microbes are attacking them [Voigt, 2014]. The two types of induced resistance that plants exhibit is induced systemic resistance (ISR) and systemic acquired resistance (SAR). Combining the two strengthens plants' resilience to disease. If an antagonist is discovered at the exposure site in the case of ISR, the biological control agent may be able to manufacture an antibiotic chemical that's transported and ultimately directly inhibits the pathogen throughout the plant. SAR induces a condition in tissues distance from the trigger location that are generated at the induction site and spread throughout the plant. It consistently provides protection from a variety of bacteria (Prasannath, 2017). A second form of defense employed by plants is referred to as an oxidative burst. The oxidative burst has to do with the release of local and global signals that start gene expression and the oxidative cross-linking of the parts of the host cell wall. *In vitro*, the source of the infection creates a large enough buildup of reactive oxygen species to kill bacteria. It is possible that the reducing oxidative bursts in the lab contribute to the induction of later defense responses [Guest and Brown, 1997].

In order to protect themselves from pathogens, plants typically use two different types of defenses: biochemical processes that occur in tissues and cells that either render the pathogen toxic or produce conditions that prohibit the plant, as well as morphological or structural traits that act as physical barriers.

Structural defense mechanisms: These may be pre-existing, which means that they exist in the plant before the pathogen even contacts it, or induced, which means that even after the pathogen has gotten past the defenses that have already been set up, in order to safeguard the plant from additional disease invasion, one or more types of structures are created.

A) Pre-existing structural defense mechanisms

These include the size, position, and types of stomata and lenticels, two naturally occurring holes that stop the spread of infections, as well as the quantity and quality of the cuticle and wax that protect the epidermal cells.

i) Waxes: Using wax to coat the surfaces of leaves and fruit to make them hydrophobic or water-repellent to stop the growth of bacteria and fungi.

ii) **Epidermal cells and Cuticle**: The thick cuticle and strong outer wall of the epidermal cells may increase resistance to infection when the pathogen only indirectly penetrates the host.

For example, disease resistance has been linked to hard outer epidermal cells and a thick cuticle in Barbery species infected with *Puccinia graminis tritici*. The cuticle of linseed serves as a defense against *Melampsora lini*.

Pyricularia oryzae and Streptomyces scabies cannot get into paddy and potatoes because the epidermal cells harden into silica and the cell walls harden into lignin.

iii) Sclerenchyma cells: Sclerenchyma cells in leaf veins and stems efficiently block some bacterial and fungal diseases that result in angular leaf patches.

iv) Structure of natural openings: All types of natural apertures act as the initial line of protection against infections. Hydathodes, for example, are openings in leaves' corners that naturally permit the flow of excess water from the core. The filaments of the leaf repel pathogens. Because of the dense hairlines on the leaves and stems, Ascochyta rabei cannot live on chickpeas. Stronger cuticles, palisade coats, and epidermis can be found on the surfaces of groundnut species' leaves which withstand Cercospora leaf. They also have fewer trichomes and stomata. In a manner similar to this, lenticels develop in the outer walls and take part in respiration. Unless the cork cells below them are suberized, these are defenseweak zones. Lenticles seem to be more resistant to microbial penetration after suberization and periderm development (Melotto et al., 2008). Twigs with horns have evolved as a result of adaptations to defend plants from livestock. The barrel cactus has spines, which are modified leaves with traits comparable to thorns. Guard cells are a separate type that are dispersed among the numerous undifferentiated epidermal cells and monitor breathing through microscopic openings known as stomata. Plants can alter the size of their stomata holes and seal their guard cells to help them fight themselves against MAMPs (Jones and Dangl, 2006). Immune plant cells known as Idioblasts, or "crazy cells", are extremely specialized. They assist in protecting crops against herbivory because they contain poisonous substances or sharp crystals, typically calcium oxalate, that harm insects' and mammals' powerful jaws as they graze. Idioblasts can be colorful cells, sclereids, crystalliferous cells, or silica cells, among other cell types. Parenchymal plant components usually contain tannins, which give them an unpleasant flavor and prevent them from being used as feed ingredients [Doughari, 2015]. Pears (*Pyrus* spp.) have a rough exterior that can rattle grazing animals' teeth due to the abundance of sclereid stone cells. Cells known as sclerosis have a haphazard arrangement and thick secondary walls that are challenging to bite through. Another kind of plant that offers chemical and physical resistance to insect predators is the trichome. *Senecio cineraria*, sometimes referred to as dusty miller, has a velvety feel due to the countless tiny trichomes that cover its surface. Trichomes on the soybean's (*Glycine max*) surface hinder insect eggs from penetrating the epidermis.

B) Post infectional structural defense mechanisms or induced structural barriers: These can be viewed as cellular defensive structures (hyphal sheathing) as well as histological defense barriers (cork layer, abscission layers, and tyloses).

i) Histological defense structures

a) Cork layer: when nematodes, bacteria, viruses, or fungus infect plants, many layers of cork cells form far from the infection. By doing this, the pathogen is prevented from causing additional damage to the plant after the initial lesion. Additionally, cork layers prevent nutrients and water from moving from the healthy area to the sick area, starving the illness.

Ex: leaves of *Prunus domestica* attacked by *Coccomyces pruniphorae*; potato tubers infected with Rhizoctonia

b) Abscission layers: A gap between the infected and healthy cells of a leaf forms around the infection site as the central lamella of parenchymatous tissue splits. The "abscission layer" is the name of this aperture. The infection moves with the diseased region as it gradually shrivels, dies, and sheds. Stone fruits with fungal, bacterial, or viral contamination develop abscission layers on their young, active leaves.

For instance, peach leaves with *Xanthomonas pruni* and *Closterosporium carpophylum*

c) **Tyloses:** Tyloses are protoplast overgrowths from living parenchymatous cells close by. They stick out of xylem vessels through pits. Tyloses have cellulose-based walls.

and grow quickly before an infection. In resistant varieties, they may completely block the xylem channels, stopping the disease from spreading. Prior to pathogen invasion, sensitive cultivars produce little to no tyloses.

Ex: When the majority of vascular wilt pathogens attack such plants, tyloses grow in the xylem arteries of those plants.

ii) Cellular defense structures

Hyphal sheathing: The extension of the cell wall produces a cellulose sheath (callose) that is loaded with phenolic compounds to stop the pathogen from spreading further. When the hyphae enter the cell wall and transform into the cell lumen, they are encased in this sheath.

For instance, *Fusarium oxysporum* f. sp. *lini*-infected flax exhibits hyphal sheathing.

II) Biochemical defense mechanisms: Chemical barriers are one of the plants' passive defense tactics against disease. Exudates on plant surfaces or other substances present in cells may promote or prevent the development of infections. Pathogens can become resistant to infection when plants do not give them the nutrients they require. As they organically grow, plants frequently release compounds that stop illnesses in their tracks. Phytoanticipins, which are naturally occurring antimicrobial chemicals in plants, are unique in that they are created during normal growth even in the absence of a pathogen or disease attack. They could be accumulated in disintegrating tissue, released into the atmosphere, or maintained inactive in vacuoles. Ouinones, catechol, and protocatechuic acid, which are all found in the dead cells of brown onion skins, stop the spores of the diseases Botrytis cinerea and Colletotrichum circinans, which cause neck rot and smudging, from growing. Phytoanticipins and other plant glycosides in the saponins category bind sterols in pathogen cell membranes, impairing membrane function and integrity. Living creatures are killed by saponins because their membranes contain sterols. The hydrolase enzyme is released after injury or infection, and it appears that healthy plant cells have molecules of inactive saponin precursors in their

vacuoles. Later, the hydrolase enzyme transforms these compounds into their active, antimicrobial form. A group of plant peptides known as plant defensins inhibits the development of bacteria, viruses, fungi, and insects. These compounds function as proteinases, lectins, ribosomal inhibitors, and polygalacturonases. The defensins stop the pathogens from growing, maturing, and needing nutrition, which allows them to resist infection. Up to 10% of the total protein is made up of defensins in legumes, solanaceous seeds, and cereals. Defensins' antifeedant properties offer protection against viruses spread by insects [Guest and Brown, 1997].

These fall under the categories of both innate and acquired biochemical defenses.

1) **Pre-existing chemical defense:** Plants release substances that either shield their surroundings from pathogen infection or that were already present in the plant cells prior to infection as a component of the pre-existing biochemical defense. These chemicals prevent growth and promote disease resistance. These substances and the resultant metabolic conditions may negatively affect the invader either directly or indirectly by promoting antagonistic plant surface microorganisms. While wound antibiotics are created in response to wounds, constitutive compounds are antibiotics that are already present in plants. More details on these chemical mechanisms of action are given.

a) The plant's environment-released inhibitors:

Some plant species contain antifungal substances, which are typically phenolic chemicals. These phenolic substances that plants release into the environment have frequently been linked to preventing disease spores from germinating before infection. For instance, red-scale onion cultivars are resistant to *Colletotrichum circinans'* onion smudge, in contrast to white onion types. The phenolic chemicals catechol and protocatechuic acid, which inhibit the germination of fungal spores, are what give the red-scale type of onion its resistance [Jones *et al.*, 1946]. Similar to this, it is generally known that vulnerable types of grams are less likely to produce

malic acid than blight-resistant cultivars [Singh *et al.*, 1998; Cagirgan *et al.*, 2011].

Ex1: The exudates released by tomato leaves kill *Botrytis cinerea*.

Ex2: Apple powdery mildew-resistant varieties create waxes on the surface of their leaves, which prevent *Podosphaera leucotricha* from germination.

Ex3: HCN is put into the roots of linseed that is resistant to the pathogen *Fusarium oxysporum* f. sp. *lini* that causes linseed wilt.

Ex4: Worms find the terthinyl present in marigold's root exudates to be inhibitory.

Ex5: The chlorogenic acid in sweet potatoes, potatoes, and carrots inhibits the growth of *Ceratocystis fimbriata*. Similarly, apples and sweet potatoes both contain phloretin and caffeic acid.

b) Inhibitors present in plant cells before infection: Some plants have the ability to produce inhibitors, which stop the growth of pathogens. For instance, the tuber peel of potatoes contains chlorogenic acids, which are phenolic compounds that provide resistance to the scabies brought on by Streptomyces scabies [Singhai *et al.*, 2011]. A glucoside in oat leaves and roots called avenasin keeps *Ophiobolus graminis* var. *avenae* from spreading diseases like take-all disease to the roots (Turk *et al.*, 2005). Arbutin, a phenolic glucoside, shields pears from the fire blight that *Erwinia amylovora* causes [Smale and Keil, 1966].

Unsaturated lactones, cyanogenic glycosides, sulfurcontaining compounds, phenols, phenolic glycosides, and saponins are only a few of the chemicals that are already present in plant cells and have antibacterial properties.

- Young fruits, leaves, and seeds contain significant amounts of phenolic compounds, tannins, and different fatty acid-like compounds, including dienes, which are responsible for the young tissues' resistance to Botrytis. Numerous hydrolytic enzymes are efficiently blocked by these substances.
- Ex: The chlorogenic acid in potatoes inhibits both the pathogen *Verticillium alboatrum* and the common scab bacteria *Streptomyces scabies*.

- The antifungal membranolytic effect of saponins cannot treat fungus infections that lack saponinases. Example: tomatine in tomatoes and avenacin in oats.
- Similar to lectins, which inhibit fungal growth and cause lysis, lectins are proteins that specifically bind to specific sugars and are abundant in a variety of seeds.
- Plant surface cells also contain varying amounts of hydrolytic enzymes such glucanases and chitinases, which can cause the disintegration of pathogen cell walls.

2) Post infectional or induces defense mechanism: Post-infection metabolic defense is assumed to be the final line of defense against pathogen infection. Currently, plants are showing resistance to diseases that produce toxins such as phytoalexins and phenol. The pathogen's growth is inhibited or stopped by these poisons, which also activate the plant's defense mechanisms.

a) **Phytoalexins** (*phyton* = plant; *alexin* = to ward off)

Muller and Borger (1940) coined the term "phytoalexins" to refer to fungistatic substances produced by plants in response to injury (mechanical or chemical) or infection.

Only in response to certain bacteria's stimulation that causes plant disease or in response to chemical and physical damage do plants which phytoalexins. are detrimental antibacterial create components. These so-called post-inhibitors are low-molecularweight secondary metabolites made by stressed plants that have antibacterial activities [Ahuja et al., 2012]. Because they gather near the infectious source, phytoalexins inhibit bacterial and fungal growth in a laboratory setting. Theoretically, as a result of this [Cruickshank and Perrin, 1960], it might be a plant defense molecule against bacterial and fungal infections. These prevent the spread of the fungus infection in the host cell by preventing the mycelium from spreading to neighboring cells [Noman et al., 2020]. Mycelial radial growth, which restricts microbe tube extension and raises mycelial dry weight, is a sign of phytoalexin fungitoxicity. Phytoalexins have the potential to have a considerable impact on the cytological, morphological, and physiological properties of fungal cells.

Characteristics of phytoalexin

- At low doses, it is bacteriostatic and fungitoxic.
- Produced in response to elicitors, host plants, or stimuli, and metabolic byproducts.
- Missing in thriving plants.
- Be produced in amounts that correspond to the size of the inoculum.
- Remain close to the point of infection.
- Created in response to diseases rather than less severe or nonpathogenic infections.
- Generated between 12 and 14 hours following the vaccine, reaching its peak 24 hours later.
- The pathogen rather than the host.

It has been shown that phytoalexin synthesis and accumulation occur in a number of families, including Leguminosae, Solanaceae, Malvaceae, Chenopodiaceae, Convolvulaceae, Compositae, and Graminaceae.

Table 1: Phytoalexin and their effect on plant disease management.

S N o.	Phytoalex in	Chemical group	Host	Pathogen	Effect on plants	Refere nce
1	Phaseollin	Isoflavan oid	French bean	C. lindemuthi anum	Reduced (74– 92%) the pathogen's radial development when studied <i>in</i> <i>vitro</i>	Pozza et al., 2021
2	Ipomeama rone	Sesquiter pene	Sweet potato	Ceratocysti s fimbriata	Induced fungal infection on plants	Lewth waite <i>et</i> <i>al.,</i> 2011

3	Rishithin	Terpenoid	Tomato	B. cinera	Increased	Charles
		s			resistance	<i>et al.,</i>
					against	2008
			_		patnogen	
4	Pisatin	-	Pea	<i>F</i> .	Inhibition of	Bani et
				oxysporum	spore	al.,
				f. sp. pisi	germination of	2018
5	Classedlin		C 1	Destas	the lungus	Talaan oo
3	Gryceonin	-	Soybean	P. sojae	ragistanaa ta	Janan ei
					fungue	2020
					Tuligus	2020
6	Isocoumar	-	Kiwi	P. syringae	Inhibit bacterial	Chen et
	in			pv.	activities/antiba	al.,
				actinidiae	cterial activity	2022
7			D 11	<i>T</i>	X 1 1 2 C 1	
1	Medicarpi	-	Dalbergi	Trametes	Inhibit fungal	Martine
	n		a	versicolor	activities/	Z- Sotros
			loro		antinungai	sources
			Dittlar		activities	2012
			tree			2012
0	Callatate	Allralaid	Chilli	C. comaici	Incurrent	Chitam
ð	Lonetotric	Aikaloid	Chili	C. capsici	increased	Cnitarr
	numme A				nathogen	<i>a ei ul.,</i> 2020
					panlogen	2020
1		1				

 Table 1 lists the phytoalexins, which are essential for preventing plant diseases.

b) Hypersensitive response (HR)

Stakman (1915) coined the phrase "hypersensitivity" to describe wheat that had the *Puccinia graminis* rust fungus infected it.

- A localized induced cell death occurs in the host plant at the site of infection as part of the hypersensitive response, which prevents the pathogen from spreading. In the portion of the plant that is damaged, HR is evident as sizable water-soaked areas that have since turned necrotic and collapsed.
- Only incompatible host-pathogen pairings can result in HR. HR can occur when avirulent pathogens are injected into susceptible

cultivars as well as when virulent pathogens are injected into non-host plants or resistant strains.

• Identification of specific signal molecules produced by infections, known as elicitors, causes HR to be activated. Cellular activity is altered when the host detects the elicitors, which results in the production of molecules related to defense.

The following are the most common innovative cell processes and substances:

- Immediately after oxidative reactions
- Increased ion transport, especially of K+ and H+ via the cell membrane

Creating pathogenesis-related proteins (like chitinases) and antimicrobial substances like phytoalexins. Membrane rupture and the loss of cell compartmentalization Strengthening of plant cell walls and the cross-linking of phenolics with cell wall constituents. **cellular reactions to HR**

- In some host-pathogen interactions, the nucleus leaves the cell as soon as it comes into contact with the pathogen and immediately disintegrates.
- At the pathogen entry point first, then spreading throughout the cytoplasm, brown resin-like granules begin to form. The cytoplasm keeps turning browner, and the invading hypha begins to degrade, which prevents further invasion.

c) Plantibodies: By using genetic engineering, transgenic plants with the ability to express foreign genes and incorporate those genes into their genomes, i.e., mouse genes that manufacture antibodies for particular plant diseases, have been created. These antibodies, also referred to as plantibodies, are produced by and found in plants yet are genetically encoded by animals. Transgenic plants, for instance, develop antibodies against viral coat proteins like the artichoke mottle crinkle virus.

d) **Pathogenesis related protein:** In response to a pathogenic invasion, plants create proteins referred to as pathogenesis-related (PR) proteins. These proteins hunt down compounds in bacterial or fungal cell walls due to their antimicrobial capabilities. The majority

of PRs primarily have an antifungal effect. Several PRs are also antiviral, antibacterial, or insecticidal, according to Dodds *et al.* (2006).

Conclusion and prospectus for the future: Due to the expanding human population, the rising use of chemical pesticides to control plant diseases, and awareness of these chemicals' negative effects on crop quality and yield, there have been requests for alternatives to chemical pesticides. The findings of this study imply that this is possible by more carefully examining plant pathogenic relationships and various plant defense mechanisms. The ability to change and use a plant's natural defenses, like making it resistant on purpose or using possible secondary plant metabolites to protect crops, could be a major step forward in the field of plant protection. Both a detailed analysis of the defense mechanisms used by plants in real life and the development of methodical strategies to use these defense mechanisms are required.

References

- Ahuja, I., Kissen, R. and Bones, AM. (2012). Phytoalexins in defense against pathogens *Trends in Plant Science*, 17(2):73–90.
- Bani, M., Cimmino, A., Evidente, A., Rubiales, D. and Rispail, N. (2018). Pisatin involvement in the variation of inhibition of *Fusarium oxysporum* f. sp. *pisi* spore germination by root exudates of *Pisum spp*. germplasm. *Plant Pathology*. 67(5), 1046–1054.
- Cagirgan, MI., Toker C., Karhan, M., Mehmet, AK., Ulger, S. and Canci, H. (2011). Assessment of endogenous organic acid levels in Ascochyta blight [Ascochyta rabiei (pass.) Labr.] susceptible and resistant chickpeas (Cicer arietinum. Turkish Journal of Field Crops. 16(2), 121-124.
- Charles, MT., Mercier, J., Makhlouf, J. and Arul, J. (2008). Physiological basis of UV-C-induced resistance to *Botrytis cinerea* in tomato fruit: I. Role of pre-and post-challenge accumulation of the phytoalexin-rishitin. *Postharvest Biology and Technology*. 47(1), 10–20.
- Chen, Q., Yu, JJ., He, J., Feng, T. and Liu, JK. (2022). Isobenzofuranones and isocoumarins from kiwi endophytic fungus *Paraphaeosphaeria sporulosa* and their antibacterial activity against *Pseudomonas syringae* pv. *actinidiae*. *Phytochemistry*. 195, 113050.
- Chitara, MK., Keswani, C. and Varnava, KG. (2020). Impact of the alkaloid colletotrichumine A on the pathogenicity of *Colletotrichum capsici* in *Capsicum annum* L. Rhizosphere. 16, 100247.

- Cruickshank, IA. and Perrin, DR. (1960). Isolation of a phytoalexin from *Pisum* sativum L. Nature. 187 (4739):799–800.
- Dodds, PN., Lawrence, GJ. and Catanzariti, AM. (2006). Direct protein interaction underlies gene-for-gene specificity and coevolution of the flax resistance genes and flax rust avirulence genes. *Proceedings of the National Academy of Sciences*. 103(23):8888–8893.
- Dodds, PN. and Rathjen, JP. (2010). Plant immunity: Towards an integrated view of plant-pathogen interactions. *Nature Reviews Genetics*. 11(8), 539–548.
- Doughari, J. (2015). An overview of plant immunity. *Journal of Plant Pathology* and Microbiology. 6(11), 10–4172.
- Fang, X., Yang, C., Wei, Y., Ma, Q., Yang, L. and Chen, X. (2011). Genomics grand for diversified plant secondary metabolites. *Plant Diversity* and Resources. 33(1), 53–64.
- Flor, HH. (1971). Current status of the gene-for-gene concept. Annual Review of *Phytopathology*. 9(1), 275–296.
- Guest, D. and Brown, J. (1997). Plant defenses against pathogens. *Plant Pathogens and Plant Diseases*. 263-286.
- Hussein, RA. and El-Anssary, AA. (2019). Plants secondary metabolites: The key drivers of the pharmacological actions of medicinal plants. *Herbal Medicine*, 1:13.
- Jahan, MA., Harris, B., Lowery, M., Infante, A., Percifield, RJ. and Kovinich, N. (2020). Glyceollin transcription factor GmMYB29A2 regulates soybean resistance to *Phytophthora sojae*. *Plant Physiology*. 183(2), 530–546.
- Jones, HA., Walker, JC., Little, TM. and Larson, RH. (1946). Relation of colorinhibiting factor to smudge resistance in onion. *Journal of Agricultural Research*. 72:259–264.
- Jones, JD. and Dangl, JL. (2006). The plant immune system. *Nature*. 444(7117), 323–329.
- Kombrink, E. and Somssich, IE. (1995). Defense responses of plants to pathogens. Advances in Botanical Research. 21:1–34.
- Lewthwaite, SL., Wright, PJ. and Triggs, CM. (2011). Sweet potato cultivar susceptibility to infection by *Ceratocystis fimbriata*. *New Zealand Plant Protection*. 64:1–6.
- Li, Y., Kong, D., Fu, Y., Sussman, M. and Wu, H. (2020). The effect of developmental and environmental factors on secondary metabolites in medicinal plants. *Plant Physiology and Biochemistry*. 148:80– 89.
- Martínez-Sotres, C., López-Albarrán, P. and Cruz-de-león, J. (2012). Medicarpin, an antifungal compound identified in hexane extract of *Dalbergia congestiflora* Pittier heartwood. *International Biodeterioration and Biodegradation*. 69, 38–40.

- Melotto, M., Underwood, W. and He, SY. (2008). Role of stomata in plant innate immunity and foliar bacterial diseases. *Annual Review of Phytopathology*. 46:101–122.
- Mukhtar, MS., Carvunis, AR. and Dreze, M. (2011). Independently evolved virulence effectors converge onto hubs in a plant immune system network. *Science*. 333(6042), 596–601.
- Noman, A., Aqeel, M. and Qari, SH. (2020). Plant hypersensitive response vs pathogen ingression: Death of few gives life to others. *Microbial Pathogenesis*. 145:104224.
- Pozza Junior, MC., Pandini, JA. and Hein, DP. (2021). Phaseolin induction on common-bean cultivars and biological control of *Colletotrichum lindemuthianum* 89 race by *Baccharis trimera* (Less.) Dc. *Brazilian Archives of Biology and Technology*, 64.
- Prasannath, K. (2017). Plant defense-related enzymes against pathogens: A review. AGRIEST *journal of Agricultural Sciences*. 11(1), 38–48.
- Singh, PJ., PAL, M. and Devakumar, C. (1998). Role of malic acid in pycnidiospore germination of *Ascochyta rabiei* and chickpea blight resistance. *Indian Phytopathology*. 51(3), 254–257.
- Singhai, PK., Sarma, BK. and Srivastava, JS. (2011). Phenolic acid content in potato peel determines natural infection of common scab caused by *Streptomyces spp.* World *Journal of Microbiology and Biotechnology*. 27(7), 1559–1567.
- Smale, BC. and Keil, HL. (1966). A biochemical study of the intervarietal resistance of Pyrus communis to fire blight. *Phytochemistry*. 5(6):1113–1120.
- Türk, FM., Egesel, CÖ. and Gül, MK. (2005). Avenacin A-1 content of some local oat genotypes and the in vitro effect of avenacins on several soilborne fungal pathogens of cereals. *Turkish Journal of Agriculture* and Forestry. 29(3), 157–164.
- Voigt, CA. (2014). Callose-mediated resistance to pathogenic intruders in plant defense-related papillae. *Frontiers in Plant Science*. 5:168.

Fascinating World of Fungi

Ritika¹ and Dolly²

Ph.D Scholar, Department of Microbiology, Chaudhary Charan Singh Haryana Agricultural University, Hisar

Email : jangraritika79@gmail.com

Introduction

Fungi are the only group of organisms on Earth that differ from one other in terms of morphological, ecophysiological, and phylogenetic characteristics, and they are the second most abundant group after insects. Fungi are almost universally present and thrive in a variety of terrestrial and aquatic settings. They are expected to have between 2.2 million and 3.8 million different species. (Hawksworth & Lücking, 2017)

They also contain organelles that are membrane-bound, such mitochondria. Glucans and chitin make up the cell wall of fungi. As heterotrophic organisms, fungi often feed on decomposing material as their "default" nutritional strategy. Many fungi grow as hyphae, which are cylindrical thread-like structures with a diameter of 2 to 10 micrometres. Some fungus exist as single-celled creatures, commonly known as yeasts. The hyphae can be septate (compartments are separated by cross walls) or non-septate. The hyphae's tips are where mushrooms grow. The hyphae are numerous

and interwoven. The ends of the hyphae are where fungi develop. The basic structure of the fungus, the mycelium, is made up of numerous interconnected hyphae. The mycelium, which is finely and intricately branched, fills a significant portion of the soil and generates a wide range of enzymes that interact with soil organic matter and mineral compounds in order to produce the nutrients and energy the fungus requires for growth.

The wide range of fungi can be categorised either taxonomic or according to their functional roles. The most recent classification schemes (Hibbett et al., 2007; James et al., 2006a,b) comprise six phyla and an extra four unplaced subphyla. Fungi frequently function in ways that traverse taxonomic boundaries. The differential between saprotrophic and mycorrhizal fungi is one that is significant. The waste or dead tissue of plants and animals serves as a source of energy and nutrients for saprotrophs. According to Smith and Read (1997), mycorrhizal fungi create mutualistic relationships with phototrophic organisms in which they directly obtain carbohydrates from their living plant partner in exchange for nutrients.

The ability to see fungal spores is another way to distinguish macrofungi from microfungi. No matter how they are classified, all fungi have the same trait of being heterotrophic, which means they depend on and are very integrated with the biota with whom they form communities. Basidiomycetes, which are saprotrophic fungi, are among the few microbes that can create enzymes that can break down lignins, the intricate phenolic compounds that preserve plant cell walls (Cairney and Meharg, 2002). Decomposed material can be transformed into minerals and made available for plant absorption in the soil water, or it can be used as energy or structural elements of the decomposer community (Dighton, 2003). In order to get nutrients from rocks, some fungi also emit organic acids (Chen et al., 2000). Fungi are also very significant. By entangling soil particles with hyphae and releasing insoluble exudates, fungi also play a significant role in the production of large stable soil aggregates (>250 m) (Rillig, 2004).

By boosting the availability of nutrients and aiding in the creation of a permeable, well-structured soil, fungi are beneficial for plants. Fungi degrade plant and animal remains in cooperation with a variety of other microorganisms in the soil, recycling nutrients as well as energy (Smith and Read, 1997). The direct provision of mineral nutrients to their symbiotic plant partner is one of the key ecological functions of mycorrhizal fungus. Around 95% of terrestrial plant species produce mycorrhizae, which emphasises the importance of this relationship (Brundrett, 1991). Fungal hyphae, which have a much smaller diameter than even fine roots (2-20 m compared to 200 m) and are capable of investigating microscopic soil holes, can acquire nutrients that are inaccessible to plants (Marschner and Dell, 1994). Common mycelial networks between plants are possible, and the transmission of nutrients from other plants on the network can aid in the establishment of seedlings or weakly competitive species (Nara, 2006; Simard and Durall, 2004).

They are essential for the movement of nutrients and maintaining ecosystem health. By improving their nutritional status and boosting their survival under adverse situations including pathogen attack, potentially toxic elements, drought, and high temperatures thanks to this ectomycorrhizal relationship, plants improve their fitness (Smith & Read, 2008). These advantages are mostly attributable to the external mycorrhizal mycelium, which serves as the link between the soil and plant components in forest ecosystems, and the mantle in the case of pathogen protection (Read & PérezMoreno, 2003). According to estimates, three square metres of forest soil contain enough ectomycorrhizal mycelium to stretch across the equator of the planet (Pérez-Moreno, 2005).

Although there is a trophic continuum in nature, three trophic groups are recognised: (a) saprotrophic, which uses organic debris as its primary source of energy; (b) parasitic, which attacks other living things and causes harm in order to survive; and (c) mutualistic, which forms advantageous symbioses and shares benefits with other organisms in order to survive. The mutualistic fungi that form mycorrhizal relationships with plants are one of the most significant structural and functional biotic components in forest ecosystems. 2008 (Smith & Read).

Fungi, on the other hand, are a vital, intriguing, and biotechnologically useful group of organisms with a huge opportunity for industrial utilisation in biotechnology. Despite this, they are also comparatively understudied. However, the same bacterial and fungal taxa have previously received attention, such as *Streptomyces* in the *Actinobacteria* and *Aspergillus* and *Penicillium* in the filamentous fungi, which are common soil moulds (Karwehl and Stadler 2017).

2. Fungal Diversity

Methods based on molecular biology are now frequently used for taxonomic identification of species to comprehend shifts in their richness and composition along different environments because many fungi are uncultivable and rarely produce visible sexual structures (Persoh 2015; Balint et al. 2016; Tedersoo and Nilsson 2016). For trustworthy assignment of ecological and functional attributes to taxa for additional ecophysiological and biodiversity analyses, accurate systematic identification to species, genus, and higher taxonomic levels is essential (Ko ljalg et al. 2013, Jeewon and Hyde 2016, etc.).

Additionally, molecular techniques have fundamentally changed the morphology-based system of classification and revolutionised our understanding of the genetic interactions among the Fungi (Hibbett et al. 2007, Wijayawardene et al. 2018). The order of divergence and classification of the major fungal groupings have been further revised because to the accessibility of fulllength rRNA gene and protein-encoding marker genome sequences (James et al. 2006a) and the development of high-resolution genomics methods (Spatafora et al. 2016, 2017).

According to the target organisms and taxonomic resolution, plant, microbial, and fungal ecologists typically test the significance of environmental factors on fungal diversity at the level of orders, classes, or phyla, but not their subranks or various ranks mixed together due to simplicity and the prevention of confusion (e.g. Tedersoo et al. 2014; Maestre et al. 2015).

3. Life Cycle and Growth: From Spore to Mycelium

A spore, that has a circumference of only a few microns (m), is often the first stage of a filamentous fungus's life. In a moist, rich in nutrients environment, the spore begins to grow and germinate. A germ tube is created, which develops into a threadlike, filamentous cell referred as a hypha. The hypha eventually forms a network of interwoven hyphal threads known as a mycelium after growing and elongating for some time. The mycelium begins to explore the air
and space so as to develop reproductive structures when nutrients in the substrate in which it lives become scarce. Ascomycetes (moulds) can develop fruiting bodies, which contain sexual spores, or conidiophores, which release asexual spores at their ends. These fruiting bodies are made up of mycelia that are different in composition from the substrate mycelia, which is more loosely packed and forms multidimensional net-like structures simulating the worldwide system of interrelated computer networks. There are visible and unseen filamentous fungus. A collection of mm- to cmlong hyphae makes up a fungal mycelium, and the diameter of a fungal hyphae can range from 2 to 10 m. The honey mushroom Armillaria bulbosa, also known as a honey mushroom in English and Hallimasch in German, has colonised over 1 000 hectares of forests soil, making it the largest and oldest organism on Earth. This example shows how the mycelia of mushroom-forming fungi are capable of colonising large surface areas (smith et al., 1992).

On numerous byproducts and waste streams from forestry and agriculture, mycelia of fungus that produce mushrooms can also develop. The composition and physical characteristics of the substrate, the ambient growth circumstances (temperature, humidity, and pH), and the genetic makeup of the fungus all affect how effectively it colonises and produces biomass. For instance, a strain of Schizophyllum commune (also known as split gill) that has two regulatory genes for mushroom development inactivated generates three times as much fungal tissue biomass as the parental strain (pelkmans et al., 2017).

In order to feed on and degrade organic matter and polymeric materials, filamentous fungus have evolved to become incredibly efficient decomposers (spatafora et al., 2017). Most of the plant

matter on Earth (450 out of 550 gigatons of carbon) is made up of polysaccharides from plant life (Bar-on et al., 2018).

4. Mushroom Magic: The Marvels of Mycology

The macrofungi known as mushrooms have fleshy, subfleshy, or occasionally leathery sporophores that resemble umbrellas and have fertile surfaces on lamellate or lining the tubes, opening out through pores. Depending on whether they are edible or poisonous, the lamellate parts are frequently referred to as "mushrooms" or "toadstools," while the tube-bearing or poroid members are referred to as "boletes" Rahi et al., 2005). The variety of mushrooms varies widely and includes anything from the common Agaricus mushrooms, which have an umbrella-shaped top and a stalk, to polypores, Earth Stars (Geastrum), Stink Horns (phalloides), and Puff Balls (Lycoperdon). Either edible mushrooms are taken from the wild or, more frequently, they are grown, picked, and processed under strict quality control standards for size, shape, tenderness, and flavour. Secondary metabolites are often created after active growth, and many of them have peculiar chemical structures. Given the significance of numerous natural products in medicine, industry, and/or agriculture, interest in these molecules is high. The production of lipids, vitamins, polysaccharides, and precursors for the synthesis of amino acids and nucleotides, which are components of cells, is also of significant economic value in modern industry (Bano et al., 1976, Chang et al., 1982, Gupta, 1989, (Sohi, 1988).

Several biotechnological breakthroughs, such as mass cultivation for various reasons, have their roots in pure culture investigations on mushrooms and similar basidiomycetous fungus. A standardisation of cultivating conditions is necessary to ensure proper mycelial colonisation of the substrates, which will be followed by differentiation of the fruit bodies. These steps include strain improvement and selection among edible fungi like *Agaricus*, making mushroom spawns and applying them to composted natural substrates, and preparation of mushroom spawns (Rahi and malik, 2016).

The basidiomycetes differentiate themselves from other fungus by a few very distinctive features. They are the most developed fungi in terms of development, and even the hyphae they produce have a unique "cellular" structure. Basidiospores are produced by basidiomycetes on a unique structure known as the basidium. At maturity, basidiospores can be uninucleate or binucleate despite being haploid. These basidiospores germinate to create hyphae when they are discharged from fully developed fruiting bodies. Mycelium is made up of hyphae as a whole. Hyphae produced from two separate mating types' spores unite to create secondary mycelium, that includes two nuclei and is essentially in charge of producing fruiting bodies (Rahi and malik, 2016).

Fungal Order/Genus	Therapeutic Properties	Bioactive Components	Mode of Action	References
Order: Agaricales	Food sources, potential nutraceuticals	Various secondary metabolites, polysaccharides	Immune modulation, antioxidant effects, anti-tumor properties	(Roberts and Evans, 2013) (Alexopolous, 2002) (Chang, 1993)

Fungal Order/Genus	Therapeutic Properties	Bioactive Components	Mode of Action	References
Order: Aphyllophorales	Decomposers, medicinal uses	Medicinally active compounds, antibiotics	Degradation of cellulose and lignin, mycorrhizal association, parasitic/pathogenic activity	(Alexopolous, 2002),Stamets, 1993), Vaidya and Rabba, 1993
Order: Auriculariales	Saprotrophs, medicinal uses	Polysaccharides, bioactive compounds	Potential anti- inflammatory, antioxidant, and immunomodulatory effects	Wang et al.,2004, Ukai et al.,1983
Gasteromycetes	Medicinal uses	Various secondary metabolites	Potential for anti- inflammatory, antioxidant, and immunomodulatory effects	Jones and Jones, 1993

5. Fungi in Action: Nature's Recyclers

Due to their eukaryotic background, fungi are more closely linked to animals and plants than they are to bacteria or archaea. The nuclei of fungal cells are attached to the membrane and contain DNAcontaining chromosomes, just like those of all eukarya, including humans (Lilly and Barnett, 1951). They also include organelles that are membrane-bound, such mitochondria. Glucans and chitin make up the cell wall of fungiⁱ. Being a saprobe, or an organism that feeds on decaying matter, is the "default" nutritional approach for fungi, which are heterotrophic creatures. While some fungi are singlecelled creatures that are typically referred to as yeasts, many of them develop as hyphae, which are cylindrical structures with a diameter of 2 to 10 micrometres. The hyphae can either be septate, in which case they are composed of compartments divided by cross walls, or nonseptate (Naranjo-Ortiz et al., 2019).

The tips of the hyphae are where fungi develop. The basic structure of the fungus, the mycelium, is made up of numerous interconnected hyphae. The mycelium, which is finely and intricately branched, fills a significant portion of the soil and generates a wide range of enzymes that interact with soil organic matter and mineral compounds to release the nutrients and energy the fungus requires to thrive. Fungi can reproduce sexually or asexually. Spores, a broad word for resilient resting structures, are created by both processes. Yeasts divide by binary fission or budding. In a normal fungal life cycle, sexual reproduction occurs when compatible spores mate, and the "imperfect stage" occurs when asexual reproduction results in the budding of spores (Watkinson et al., 2015)

Although fungus undertake a wide range of tasks, the saprotrophs, mycorrhizas, and lichens are three functional groupings of fungi that are particularly significant in soil ecological systems. The large variety of enzymes produced by saprophytic fungus includes amylases, proteases, lipases, and phosphatases. The hyphae at the top of the mycelium, which is growing through its substrate, create these enzymes. The mycelium will frequently spread outward in a radial pattern from a single germinated spore, forming a circular structure of metabolic activity. The sugars, peptides, amino acids, and lipids released by the fungal enzymes may not always be consumed by this fungus; instead, bacteria, plants, and other soil biota, including other fungi, struggle fiercely for them (Corbu et al., 2023).

Therefore, saprotrophic fungi increase the biomass and variety of soils and are essential to degradation by supplying substrates to other soil organisms. This is especially true for types of saprotrophic fungi that are experts in breaking down resistant plant and animal materials like cellulose (found in plant fibres) and lignin (found in plants) as well as chitin (found in other fungus and insect exoskeletons) (Crowther et al., 2012). For instance, 'white-rot' fungi are special because they are able to break down lignin into less resistant compounds that can be acted upon by enzymes from a wider range of organisms. Fungi that are saprotrophic are essential to the global greenhouse gas cycle (Shah et al., 1992).

Mycorrhizal fungi associate with living plant roots in mutually beneficial symbiotic relationships. The symbiotic relationship is based on the transfer of resources: the plant gives the fungus sugars as an alternative source of carbon, and the fungus gives the plant nutrients from the soil(Huey et al., 2020). Mycorrhizal associations are formed by the vast majority of terrestrial plants, and they enable plants to occupy a considerably wider range of soil habitats than would otherwise be conceivable. The greatest variety of host plants can establish symbioses with arbuscular (AM) and ectomycorrhizal (EM) fungus. About 80% of the species of plant are colonised by AM fungus, which are particularly common in herbaceous species like many significant agricultural crops (Huey et al., 2020).

The mycelia of AM fungal species are typically smaller than those of EM fungi, but these are especially crucial for allowing plants access to soil that contains inorganic phosphorus. Most prominent trees and woody plants in temperate regions, including the commercially significant pine, spruce, fir, oak, beech, poplar, and willow, develop relationships with EM fungus. In EM relationships, the fungus primarily adheres to the root's surface and only reaches the spaces between root cells, although it has the potential to grow a substantial extra-radical mycelium. EM fungi play a crucial role in the decomposition of organic matter in soils, just like saprotrophic fungi. EM fungi may be able to manufacture enzymes that are more expensive to produce energetically than ordinary saprotrophs since they are powered by carbon from the plant (Lindahl et al. 2007).

Because EM fungi have a large amount of mycelium, which allows their vegetative hyphae to fuse with one another (anastomose), and because EM fungi tend to be non-specific to host plants, EM fungus frequently develop huge, intricate underground connections known as mycorrhizal networks. All major terrestrial ecosystems have mycorrhizal networks (MNs), which allow resources including carbon, nutrients, water, defensive signals, and allelochemicals to be exchanged between plants. A mycorrhizal network that already exists is where nearly all seeds that germinate in soil do so, enabling the young plant to swiftly use this resource-transfer channel (Teste et al. 2009).

As a result, MNs have significant effects on the establishment, survival, and growth of plants. They also have ramifications for the diversity and stability of plant communities in response to external stress. Because they offer pathways for ideas and cross-scale interactions that result in self-organisation and emergent features, MNs are thought to be crucial to ecosystems as complex adaptive systems (Simard et al., 2012). Lichens are symbiotic, mutualistic relationships between a fungus and a green alga (bipartite symbiosis), and cyanobacteria (tripartite symbiosis) on occasion. The fungus provides the 'body' of the symbiosis by shielding the photobionts from radiation and dehydration and secreting organic acids that release insoluble minerals from the substrate (Simard et al., 2012).

Algae use photosynthesis to create carbon, while cyanobacteria, if any are present, fix nitrogen in the atmosphere into ammonium, a useful form of Nitrogen. Lichens are an example of a symbiosis that is nutritionally independent, extremely resilient to temperature and humidity extremes, and uniquely adapted to desiccation. They may survive in a variety of ecosystems that are inhospitable to plants because of this, such as the High Arctic, the Antarctic, and alpine and desert regions. Lichens often adopt one of three growth types on these bare substrates: foliose (leafy), fruticose (lacy), or crustose (producing a crust). Through the breakdown of primary substrates caused by the organic acids they secrete, they aid in the development of a soil profile and the initial succession of plants onto these new soil (Aislabie et al., 1999).

Symbiotic Partnerships: Fungi's Cooperative Relationships

The symbiotic associations that can be developed between heterotrophic fungus and photoautotrophic partners, including plants, mosses, cyanobacteria, and algae, are diverse. Lichens are just one example. Fungi and vascular plants have a wide variety of ecologically significant relationships. Some of these connections are known. such ectomycorrhizas well (Agerer, 1991). endomycorrhizas, and the particular orchid mycorrhizal associations (Rasmussen, 2002). Algicolous fungi and bacteria or algae (Hawksworth, 1987) and lichens (Hawksworth, 1988) are two examples of well-known and varied connections between fungi and cyanobacteria algae. Algicolous fungus can coexist or mutualistically, as in the case of mycophycobioses (Kohlmeyer and Kohlmeyer, 1972), or they may parasitize algae or cyanobacteria (Kohlmeyer and Demoulin, 1981), (Sønstebø and Rohrlack, 2011).

Lichen-forming fungi may form symbiotic relationships with algae or cyanobacteria to create the lichen, which has its own distinct identity. Lichens are one example of the incredibly numerous relationships between fungi and photosynthetic organisms, despite being unique. The term "symbiosis" was coined by the German mycologist Anton de Bary to describe the circumstance in which different species coexist (Oulhen et al.,2016) corroborating Schwendener's hypothesis that lichen is made up of two distinct creatures, a fungus and an alga. Since then, lichens have been understood to be an obligatory association between a fungus (mycobiont) and a partner that uses light for photosynthesis, such as cyanobacteria or green algae (photobiont) (Nash, 2008), (Rikkinen, 2013).

The viability of this mutual connection is dependent on the mutualistic-antagonistic interactions between a large number of interconnected organisms, also referred to as the "holobiont" (Margulis and Fester, 1991), (Douglas and werren, 2016). The stability of the environment, the availability of partners, and the interactional intimacy of the symbionts all play a role in this complicated relationship (Rafferty et al., 2015), (Chomicki and Renner, 2017).

From a Multi-Species Symbiotic Relationship to a Dual Partnership The photobiont delivers carbon fixed by photosynthesis as the system's energy source, and the fungal partner provides the water, mineral nutrients, and protective structures for the system. Generalist fungi have been similarly frequently defined (Wietz et al., 2003), (Blaha et al., 2003), despite the fact that the fungus can be highly particular when choosing its photobiont (Piercey and DePriest, 2001), (Magain et al., 2017). According to the variety of photobionts that they can lichenize, Yahr et al., (2006) divided photobionts into three categories: specialists, intermediates, and generalists. According to this approach, photobiont generalists accept a wide range of algal partners and can establish associations with a variety of strains depending on the environment, whereas photobiont specialists partner with specific algal lineages.

A smaller number of algal companions create symbiotic interactions with intermediates. Photobiont specialists are usually linked to lichens with broad ecological niches and geographic distributions, which are able to link up with locally adapted photobionts in various climatic zones (Blaha et al., 2006), (Fernández-Mendoza et al., 2011), (Vargas Castillo and Beck, 2012). The precise variables that affect photobiont choice are well understood and appear to be influenced by phylogenetic specialisation, fungus reproductive approach, photobiont cell availability, and ecological variables like climate or substrate (Scheidegger, 1985). In a surprising number of investigations (del Campo, 2010, Moya et al., 2020), different algae species have been seen in conjunction with the same thallus. For instance, Backor et al.2018, and Casano et al.,2011 both confirmed the existence of many algal genotypes in a single lichen thallus.

A transient phase of interactions with more than one photobiont in individual thalli may have occurred during the ecological diversification and speciation of lichen microorganisms in various habitats, according to Del Campo et al. and Casano et al.'s findings on Ramalina farinacea thalli. The current pattern of algal cooperation is likely encouraged by their various and complimentary ecophysiological reactions, which let the lichen flourish in a variety of environments and geographical locations (Casano et al., 2011).

Employing next-generation sequencing methods, bacterial and secondary fungal communities living in lichens have been identified (e.g., Grube et al., Tuovinen et al.,). These unrecognised species may play important functions as functional elements in organising the thalli and influencing the response to external stimuli (Grube et al.,2009, Cernava et a., 2017). Individual bacterial strains have been recovered from lichens since the first half of the 20th century, with Alpha proteobacteria constituting the dominant group and being noted for the first time by Cardinale et al.,2008. According to research on the variety of bacteria that are linked with lichens, the thallus's various regions can affect microbial colonisation by providing various chemical and physiological micro-niches (Grube et al., 2009, Hodkinson and Lutzoni, 2009). The fungal companion is another important component influencing the makeup of bacteria.

In context with this, Aschenbrenner et al. 2016 showed that the lichen *Lobaria pulmonaria* displayed a core and common fraction of its bacterial biome as well as a transient fraction. Additionally, they showed that the lichen's vegetative propagules contained bacteria that permitted asexual vertical transmission. It has long been believed that the mycobiont forms the thallus by a specific contact with an appropriate alga, and that a large number of linked and potentially interaction bacterial and other companions then more or less specifically colonise the lichen. Recently, it has been proposed that the microbiota influences the development of lichenization (Speibille et al., 2020). Similar concerns have also been raised in connection to the function and specialisation of bacteria within lichens.

In two epiphytic lichens, Spribille et al., 2016 discovered that a particular group of basidiomycetous yeasts contributed to the microbiome and that the quantity of the yeast within the lichen was correlated with concentrations of vulpinic acid, a secondary metabolite linked to lichen defences. More recently, in the northern hemisphere, several *Cladonia* species included hitherto unidentified *cystobasidiomycete* symbionts that Cernajova and Skaloud, 2019 found. These experiments consistently found basidiomycetous yeast within lichens, indicating that this organism represents the third partner in the lichen complex. *Cyphobasidium spp.* was not recognised by Millanes et al., 2016 as a third mutualistic partner, but rather as a fungus related to lichens that can produce galls on the thalli.

Oberwinkler, 2017, who explored the significance of these lichenrelated yeasts, stated that "it is obvious that basidiomycetous yeasts in lichen thalli are not a third component of symbiosis, but rather the vegetative propagules of mycoparasites."

Two main categories of fungi that are symbionts with plants are endophytic fungi and mycorrhizal fungi. Endophytic fungi are totally contained within plant tissues and may be found on roots, stems, or leaves. Mycorrhizal fungi, on the other hand, are found solely in roots and spread out into the rhizosphere. Mutualism, commensalism, and parasitism are just a few of the different symbiotic lives that fungi exhibit as symbionts. Mutualistic symbioses give host fitness benefits that can lead to stress and metal tolerance,(Read, 1999) resistance to diseases, thermotolerance, growth enhancement,(Redman et al., 2002,Marks and Clay, 1990, Varma et al., 1999) herbivore resistance, and increased nutrient acquisition (Read, 1999). In contrast to commensal symbioses, which have no positive nor negative impacts on their hosts, parasitic symbioses have an adverse impact on host fitness by reducing growth rates and/or fertility or by causing illness symptoms that may be fatal (Redman et al., 2001). Endophytes may receive mutualistic benefits from obtaining nutrients from hosts, avoiding biotic and abiotic stress, and spreading via seed transfer (Schardl et al., 2004, Schulz, 2006).

When a plant or its host tissue reaches senescence, endophytic fungi, which may develop inside of roots, stems, and/or leaves, emerge to sporulate (Sherwool and caroll, 1974, Stone et al., 2004). Thus, a vast variety of fungi, including dormant saprophytes and latent pathogens, are included in the category of endophytes. Recent phylogenetic data, however, show that some endophytes, despite sharing a physical identity with known parasites in the same host, are genetically separate (Caroll, 1988). Constitutive mutualists (Class I endophyte) and inducible mutually beneficial (Class II endophyte) are two separate categories of endophytic fungus, according to Carroll, 1988. Typically, it is hypothesised that the clavicipitaceous majority of Class I endophytes (Epichlo/neotyphodium) solely infect grass and are systemic and vertically distributed through seeds. Contrarily, nonsystemic Class II endophytes have a wide range of taxonomic classifications, are horizontally spread from one plant to another, and colonise practically all plants in environments (Rodriguez et al., 2008). Based on host range, colonisation pattern, conveyance, and ecological role, endophytes can currently be categorised into four classes.

Fungal endophytes give stress tolerance. Numerous studies have demonstrated that fungal endophytes help host species become more stress-tolerant and are crucial for at least certain plants' survival in high-stress settings (Rodriguez et al., 2004). Class II endophytes, for instance, provide heat tolerance to plants growing in geothermal soils (Redman et al., 2002), the degree of endophyte colonisation of tree leaves correlates with the host's resistance to pathogens (Arnold et al., 2003), and endophytes confer drought tolerance to a variety of host species (Waller et al., 2005). Class I endophytes or clavipitaceous. Class I endophytes are restricted to a few cool- and and comprise small number of warm-season grasses а phylogenetically related clavicipitaceous taxa (Dighton and White, 2005). These endophytes are unique in that they complete their whole life cycle inside the host grass's aerial section, developing nonpathogenic, systemic, and typically intercellular connections (Baccon and Battista, 1991). It is common for Class I endophytes to boost plant biomass, impart drought resistance, and create compounds that are poisonous to animals and inhibit herbivory (Clay, 1988). The advantages offered by these fungi, however, seem to be influenced by the host species, host genotype, and environmental factors.90-92(Saikkonen et al., 1999, Faeth et al., 2006). Infected populations of grass frequently expand over time, which shows that endophytes, despite growing at the expense of host metabolism, give their hosts an adaptive benefit. Perennial ryegrass and tall fescue both grow more quickly when they are endophyteinfected (Clay, 1988). Neotyphodium endophytes also improve grasses' ability to withstand drought by osmoregulation and stomatal regulation (Baccon and Hill, 1996).

Conclusion

The world of fungi is a fascinating and important realm that holds the key to the balance of ecosystems and offers numerous benefits to human life. Fungi, such as mushrooms and molds, are essential in nature's recycling and play a vital role in breaking down organic matter, facilitating decomposition and nutrient cycling. They also play a significant role in medicine, materials, and industries, making them silent partners in our everyday lives.

Fungi's cooperative spirit is evident in their symbiotic relationships, such as mycorrhizae and lichens, which promote mutual growth and survival. However, fungi face threats such as habitat loss and pollution, which threaten their existence and the delicate balance of our ecosystems. Recognizing the urgency of fungal conservation and preservation is vital to safeguarding the delicate web of life on Earth. These unassuming organisms are fundamental to the health and sustainability of our planet, and through continued exploration and commitment to their protection, we can ensure that the marvels of fungi remain an enduring legacy for generations to come.

References

- Agerer, R. (1991). 2 Characterization of Ectomycorrhiza. In *Methods in microbiology* (Vol. 23, pp. 25-73). Academic Press.
- Aislabie, J., Davison, A., Boul, H., Franzmann, P., Jardine, D., & Karuso, P. (1999).
 Isolation of Terrabacter sp. strain DDE-1, which metabolizes 1, 1dichloro-2, 2-bis (4-chlorophenyl) ethylene when induced with biphenyl. *Applied and Environmental Microbiology*, 65(12), 5607-5611.
- Alexopoulos, C. J., Mims, C. W., & Blackwell, M. (1996). *Introductory mycology*: John Wiley and Sons.
- Arnold, A. E., Mejía, L. C., Kyllo, D., Rojas, E. I., Maynard, Z., Robbins, N., & Herre, E. A. (2003). Fungal endophytes limit pathogen damage in a tropical tree. *Proceedings of the National Academy of Sciences*, 100(26), 15649-15654.
- Aschenbrenner, I. A., Cernava, T., Berg, G., & Grube, M. (2016). Understanding microbial multi-species symbioses. *Frontiers in Microbiology*, 7, 180.
- Bačkor, M., Peksa, O., Škaloud, P., & Bačkorová, M. (2010). Photobiont diversity in lichens from metal-rich substrata based on ITS rDNA sequences. *Ecotoxicology and environmental safety*, 73(4), 603-612.
- Bacon, C. W., & De Battista, J. (1991). Endophytic fungi of grasses.

- Bacon, C. W., & Hill, N. S. (1996). Symptomless grass endophytes: products of coevolutionary symbioses and their role in the ecological adaptations of grasses. *Endophytic fungi in grasses and woody plants: systematics,* ecology, and evolution. APS Press, St Paul, MN, 155-178.
- Bálint, M., Bahram, M., Eren, A. M., Faust, K., Fuhrman, J. A., Lindahl, B., . . . Unterseher, M. (2016). Millions of reads, thousands of taxa: microbial community structure and associations analyzed via marker genes. *FEMS microbiology reviews*, 40(5), 686-700.
- Bano, Z. (1976). The nutritive value of mushrooms, in Proceeding of First Symposium on Survey and Cultivation of Edible Mushroom of India, vol. 2, pp. 148–160, Regional Research Laboratory, Srinagar, India.
- Bar-On, Y. M., Phillips, R., & Milo, R. (2018). The biomass distribution on Earth. Proceedings of the National Academy of Sciences, 115(25), 6506-6511.
- Barea, J.-M., Ferrol, N., Azcón-Aguilar, C., & Azcón, R. (2008). Mycorrhizal symbioses. *The ecophysiology of plant-phosphorus interactions*, 143-163.
- Blaha, J., Baloch, E., & Grube, M. (2006). High photobiont diversity associated with the euryoecious lichen-forming ascomycete Lecanora rupicola (Lecanoraceae, Ascomycota). *Biological Journal of the Linnean Society*, 88(2), 283-293.
- Brundrett, M. (1991). Mycorrhizas in natural ecosystems *Advances in ecological research* (Vol. 21, pp. 171-313): Elsevier.
- Cairney, J. W., & Meharg, A. A. (2002). Interactions between ectomycorrhizal fungi and soil saprotrophs: implications for decomposition of organic matter in soils and degradation of organic pollutants in the rhizosphere. *Canadian Journal of Botany*, 80(8), 803-809.
- Cardinale, M., Vieira de Castro Jr, J., Müller, H., Berg, G., & Grube, M. (2008). In situ analysis of the bacterial community associated with the reindeer lichen Cladonia arbuscula reveals predominance of Alphaproteobacteria. *FEMS microbiology ecology*, 66(1), 63-71.
- Carroll, G. (1988). Fungal endophytes in stems and leaves: from latent pathogen to mutualistic symbiont. *Ecology*, 69(1), 2-9.
- Casano, L. M., del Campo, E. M., García-Breijo, F. J., Reig-Armiñana, J., Gasulla, F., Del Hoyo, A., ... & Barreno, E. (2011). Two Trebouxia algae with different physiological performances are ever-present in lichen thalli of Ramalina farinacea. Coexistence versus competition?. *Environmental microbiology*, 13(3), 806-818.
- Castillo, R. V., & Beck, A. (2012). Photobiont selectivity and specificity in Caloplaca species in a fog-induced community in the Atacama Desert, northern Chile. *Fungal Biology*, 116(6), 665-676.
- Černajová, I., & Škaloud, P. (2019). The first survey of Cystobasidiomycete yeasts in the lichen genus Cladonia; with the description of Lichenozyma pisutiana gen. nov., sp. nov. *Fungal Biology*, *123*(9), 625-637.

- Cernava, T., Erlacher, A., Aschenbrenner, I. A., Krug, L., Lassek, C., Riedel, K., ... & Berg, G. (2017). Deciphering functional diversification within the lichen microbiota by meta-omics. *Microbiome*, 5(1), 1-13.
- Chang, S. T. (1993). Mushroom and mushroom biology, in Genetics and Breeding of Edible Mushrooms, S. T. Chang, J. A. Bushwell, and P. G. Miles, Eds., pp. 1–3, CRC Press.
- Chang, S.-t., & Quimio, T. H. (1982). *Tropical mushrooms: biological nature and cultivation methods*: Chinese University Press.
- Chen, J., Blume, H.-P., & Beyer, L. (2000). Weathering of rocks induced by lichen colonization—a review. *Catena*, *39*(2), 121-146.
- Chomicki, G., & Renner, S. S. (2017). Partner abundance controls mutualism stability and the pace of morphological change over geologic time. *Proceedings of the National Academy of Sciences*, 114(15), 3951-3956.
- Clay, K. (1988). Fungal endophytes of grasses: a defensive mutualism between plants and fungi. *Ecology*, 69(1), 10-16.
- Corbu, V. M., Gheorghe-Barbu, I., Dumbravă, A. Ş., Vrâncianu, C. O., & Şesan, T. E. (2023). Current Insights in Fungal Importance—A Comprehensive Review. *Microorganisms*, 11(6), 1384.
- Crowther, T. W., Boddy, L., & T Jones, H. (2012). Functional and ecological consequences of saprotrophic fungus–grazer interactions. *The ISME journal*, 6(11), 1992-2001.
- Del Campo, E. M., Casano, L. M., Gasulla, F., & Barreno, E. (2010). Suitability of chloroplast LSU rDNA and its diverse group I introns for species recognition and phylogenetic analyses of lichen-forming Trebouxia algae. *Molecular phylogenetics and evolution*, 54(2), 437-444.
- Dighton, J. (2018). Fungi in ecosystem processes: CRC press.
- Dighton, J., & White, J. F. (2005). Evolutionary Development of the Clavicipitaceae. In *The Fungal Community* (pp. 525-538). CRC Press.
- Douglas, A. E., & Werren, J. H. (2016). Holes in the hologenome: why hostmicrobe symbioses are not holobionts. *MBio*, 7(2), 10-1128.
- Faeth, S. H., Gardner, D. R., Hayes, C. J., Jani, A., Wittlinger, S. K., & Jones, T. A. (2006). Temporal and spatial variation in alkaloid levels in Achnatherum robustum, a native grass infected with the endophyte Neotyphodium. *Journal of chemical ecology*, 32, 307-324.
- Fernández-Mendoza, F., Domaschke, S., García, M. A., Jordan, P., Martín, M. P., & Printzen, C. (2011). Population structure of mycobionts and photobionts of the widespread lichen Cetraria aculeata. *Molecular Ecology*, 20(6), 1208-1232.
- Ganley, R. J., Brunsfeld, S. J., & Newcombe, G. (2004). A community of unknown, endophytic fungi in western white pine. *Proceedings of the National Academy of Sciences*, 101(27), 10107-10112.

- Grube, M., Cardinale, M., de Castro, J. V., Müller, H., & Berg, G. (2009). Speciesspecific structural and functional diversity of bacterial communities in lichen symbioses. *The ISME journal*, 3(9), 1105-1115.
- Grube, M., Cernava, T., Soh, J., Fuchs, S., Aschenbrenner, I., Lassek, C., ... & Berg, G. (2015). Exploring functional contexts of symbiotic sustain within lichen-associated bacteria by comparative omics. *The ISME journal*, 9(2), 412-424.
- Gupta, V. P. (1989). Mushroom yield- a rich food-a profitable commercial crop, Kisan World, vol. 37.
- Hawksworth, D. L. (1987). Observations on three algicolous microfungi. *Notes* from the Royal Botanic Garden Edinburgh.
- Hawksworth, D. L. (1988). The variety of fungal-algal symbioses, their evolutionary significance, and the nature of lichens. *Botanical journal of the Linnean Society*, 96(1), 3-20.
- Hawksworth, D. L., & Lücking, R. (2017). Fungal diversity revisited: 2.2 to 3.8 million species. *Microbiology spectrum*, 5(4), 10.1128/microbiolspec. funk-0052-2016.
- Hibbett, D. S., Binder, M., Bischoff, J. F., Blackwell, M., Cannon, P. F., Eriksson, O. E., . . . Lücking, R. (2007). A higher-level phylogenetic classification of the Fungi. *Mycological research*, 111(5), 509-547.
- Hodkinson, B. P., & Lutzoni, F. (2009). A microbiotic survey of lichen-associated bacteria reveals a new lineage from the Rhizobiales. *Symbiosis*, 49, 163-180.
- Hudson, H. J. (1991). Fungal biology: CUP Archive.
- Huey, C. J., Gopinath, S. C., Uda, M., Zulhaimi, H. I., Jaafar, M. N., Kasim, F. H., & Yaakub, A. R. W. (2020). Mycorrhiza: a natural resource assists plant growth under varied soil conditions. *3 Biotech*, *10*, 1-9.
- Hyde, K. D., Xu, J., Rapior, S., Jeewon, R., Lumyong, S., Niego, A. G. T., . . . Brooks, S. (2019). The amazing potential of fungi: 50 ways we can exploit fungi industrially. *Fungal Diversity*, 97, 1-136.
- Ingold, C. T. (2012). The biology of fungi: Springer Science & Business Media.
- James, T. Y., Kauff, F., Schoch, C. L., Matheny, P. B., Hofstetter, V., Cox, C. J., . . Miadlikowska, J. (2006). Reconstructing the early evolution of Fungi using a six-gene phylogeny. *Nature*, 443(7113), 818-822.
- James, T. Y., Letcher, P. M., Longcore, J. E., Mozley-Standridge, S. E., Porter, D., Powell, M. J., . . Vilgalys, R. (2006). A molecular phylogeny of the flagellated fungi (Chytridiomycota) and description of a new phylum (Blastocladiomycota). *Mycologia*, 98(6), 860-871.
- Jeewon, R., & Hyde, K. D. (2016). Establishing species boundaries and new taxa among fungi: recommendations to resolve taxonomic ambiguities. *Mycosp*, 7(11), 1669-1677.
- Jones, A., & Jones, E. (1993). Observations on the marine gasteromycete Nia vibrissa. *Mycological research*, 97(1), 1-6.

- Karwehl, S., & Stadler, M. (2016). Exploitation of fungal biodiversity for discovery of novel antibiotics. How to Overcome the Antibiotic Crisis: Facts, Challenges, Technologies and Future Perspectives, 303-338.
- Kohlmeyer, J. A. N., & Kohlmeyer, E. (1972). Is Ascophyllum nodosum lichenized?.
- Kohlmeyer, J., & Demoulin, V. (1981). Parasitic and symbiotic fungi on marine algae.
- Köljalg, U., Larsson, K. H., Abarenkov, K., Nilsson, R. H., Alexander, I. J., Eberhardt, U., . . . Larsson, E. (2005). UNITE: a database providing webbased methods for the molecular identification of ectomycorrhizal fungi. *New Phytologist*, 166(3), 1063-1068.
- Latch, G. C. (1993). Physiological interactions of endophytic fungi and their hosts. Biotic stress tolerance imparted to grasses by endophytes. Agriculture, Ecosystems & Environment, 44(1-4), 143-156.
- Lilly, V. G., & Barnett, H. L. (1951). Physiology of the fungi. *Physiology of the fungi*.
- Lindahl, B. D., Ihrmark, K., Boberg, J., Trumbore, S. E., Högberg, P., Stenlid, J., & Finlay, R. D. (2007). Spatial separation of litter decomposition and mycorrhizal nitrogen uptake in a boreal forest. *New Phytologist*, 173(3), 611-620.
- Maestre, F. T., Delgado-Baquerizo, M., Jeffries, T. C., Eldridge, D. J., Ochoa, V., Gozalo, B., . . Ulrich, W. (2015). Increasing aridity reduces soil microbial diversity and abundance in global drylands. *Proceedings of the National Academy of Sciences*, 112(51), 15684-15689.
- Magain, N., Miadlikowska, J., Goffinet, B., Sérusiaux, E., & Lutzoni, F. (2017). Macroevolution of specificity in cyanolichens of the genus Peltigera section Polydactylon (Lecanoromycetes, Ascomycota). Systematic Biology, 66(1), 74-99.
- Margulis, L., & Fester, R. (Eds.). (1991). Symbiosis as a source of evolutionary innovation: speciation and morphogenesis. MIT press.
- Marks, S., & Clay, K. (1990). Effects of CO 2 enrichment, nutrient addition, and fungal endophyte-infection on the growth of two grasses. *Oecologia*, 84, 207-214.
- Marschner, H., & Dell, B. (1994). Nutrient uptake in mycorrhizal symbiosis. *Plant and Soil, 159*, 89-102.
- Millanes, A. M., Diederich, P., & Wedin, M. (2016). Cyphobasidium gen. nov., a new lichen-inhabiting lineage in the Cystobasidiomycetes (Pucciniomycotina, Basidiomycota, Fungi). *Fungal Biology*, 120(11), 1468-1477.
- Moya, P., Molins, A., Chiva, S., Bastida, J., & Barreno, E. (2020). Symbiotic microalgal diversity within lichenicolous lichens and crustose hosts on Iberian Peninsula gypsum biocrusts. *Scientific Reports*, 10(1), 14060.

- Muggia, L., Vancurova, L., Škaloud, P., Peksa, O., Wedin, M., & Grube, M. (2013). The symbiotic playground of lichen thalli–a highly flexible photobiont association in rock-inhabiting lichens. *FEMS Microbiology Ecology*, 85(2), 313-323.
- Nara, K. (2006). Ectomycorrhizal networks and seedling establishment during early primary succession. *New Phytologist*, *169*(1), 169-178.
- Naranjo-Ortiz, M. A., & Gabaldón, T. (2019). Fungal evolution: diversity, taxonomy and phylogeny of the Fungi. *Biological Reviews*, 94(6), 2101-2137.
- Nash, T.H.I. (2008). Lichen Biology, 2nd ed.; Cambridge University Press: Cambridge, UK, ISBN 9780521871624.
- Noh, H. J., Park, Y., Hong, S. G., & Lee, Y. M. (2021). Diversity and physiological characteristics of Antarctic lichens-associated bacteria. *Microorganisms*, 9(3), 607.
- Oberwinkler, F. (2017). Yeasts in pucciniomycotina. *Mycological Progress*, 16, 831-856.
- Oulhen, N., Schulz, B. J., & Carrier, T. J. (2016). English translation of Heinrich Anton de Bary's 1878 speech, 'Die Erscheinung der Symbiose' ('De la symbiose'). *Symbiosis*, 69(3), 131-139.
- Pardo-De la Hoz, C. J., Magain, N., Lutzoni, F., Goward, T., Restrepo, S., & Miadlikowska, J. (2018). Contrasting symbiotic patterns in two closely related lineages of trimembered lichens of the genus Peltigera. *Frontiers* in Microbiology, 9, 2770.
- Pelkmans, J. F., Patil, M. B., Gehrmann, T., Reinders, M. J., Wösten, H. A., & Lugones, L. G. (2017). Transcription factors of Schizophyllum commune involved in mushroom formation and modulation of vegetative growth. *Scientific reports*, 7(1), 310.
- Pérez-Moreno, J. (2005). Transferencia de nutrientes: Hongos ectomicorrízicos. Scientific American Latinoamerica, 33, 32–33.
- Peršoh, D. (2015). Plant-associated fungal communities in the light of meta'omics. *Fungal Diversity*, 75(1), 1-25.
- Piercey-Normore, M. D., & DePriest, P. T. (2001). Algal switching among lichen symbioses. *American journal of botany*, 88(8), 1490-1498.
- Rafferty, N. E., CaraDonna, P. J., & Bronstein, J. L. (2015). Phenological shifts and the fate of mutualisms. *Oikos*, *124*(1), 14-21.
- Rahi, D. K., Rajak, R. C., Shukla, K. K., & Pandey, A. K. (2005). Diversity and nutraceutical potential of wild edible mushrooms of Central India, in Microbial Diversity: Current Perspectives and Potential Applications, pp. 967–980.
- Rahi, D. K., & Malik, D. (2016). Diversity of mushrooms and their metabolites of nutraceutical and therapeutic significance. *Journal of Mycology*, 2016.

- Rahi, D. K., Shukla, K. K., Rajak, R. C., & Pandey, A. K. Agaricales of Central India II: a new species of Cantharellus," Indian Journal of Mushrooms, In press.
- Rasmussen, H. N. (2002). Recent developments in the study of orchid mycorrhiza. *Plant and soil*, 244, 149-163.
- Read, D. J. (1999). Mycorrhiza—the state of the art. *Mycorrhiza: structure, function, molecular biology and biotechnology*, 3-34.
- Read, D., & Perez-Moreno, J. (2003). Mycorrhizas and nutrient cycling in ecosystems–a journey towards relevance? *New Phytologist*, 157(3), 475-492.
- Redman, R. S., Dunigan, D. D., & Rodriguez, R. J. (2001). Fungal symbiosis from mutualism to parasitism: who controls the outcome, host or invader?. *New Phytologist*, 151(3), 705-716.
- Redman, R. S., Sheehan, K. B., Stout, R. G., Rodriguez, R. J., & Henson, J. M. (2002). Thermotolerance generated by plant/fungal symbiosis. *Science*, 298(5598), 1581-1581.
- Rikkinen, J. (2013). Molecular studies on cyanobacterial diversity in lichen symbioses. *MycoKeys*.
- Rillig, M. C. (2004). Arbuscular mycorrhizae and terrestrial ecosystem processes. *Ecology letters*, 7(8), 740-754.
- Roberts, P., & Evans, S. (2013). The Book of Fungi, Ivy Press, Lewes, UK.
- Rodriguez, R. J., Henson, J., Van Volkenburgh, E., Hoy, M., Wright, L., Beckwith, F., ... & Redman, R. S. (2008). Stress tolerance in plants via habitatadapted symbiosis. *The ISME journal*, 2(4), 404-416.
- Rodriguez, R. J., Redman, R. S., & Henson, J. M. (2004). The role of fungal symbioses in the adaptation of plants to high stress environments. *Mitigation and adaptation strategies for global change*, 9, 261-272.
- Rodriguez, R. J., White Jr, J. F., Arnold, A. E., & Redman, A. R. A. (2009). Fungal endophytes: diversity and functional roles. *New phytologist*, 182(2), 314-330.
- Saikkonen, K., Helander, M., Faeth, S. H., Schulthess, F., & Wilson, D. (1999). Endophyte-grass-herbivore interactions: the case of Neotyphodium endophytes in Arizona fescue populations. *Oecologia*, 121, 411-420.
- Schardl, C. L., Leuchtmann, A., & Spiering, M. J. (2004). Symbioses of grasses with seedborne fungal endophytes. *Annu. Rev. Plant Biol.*, 55, 315-340.
- Scheidegger, C. (1985). Systematische Studien zur Krustenflechte Anzina carneonivea (Trapeliaceae, Lecanorales). Nova Hedwigia, 41(1-4), 191-218.
- Schulz, B. (2006). Mutualistic interactions with fungal root endophytes. *Microbial root endophytes*, 261-279.

- Shah, M. M., Barr, D. P., Chung, N., & Aust, S. D. (1992). Use of white rot fungi in the degradation of environmental chemicals. *Toxicology letters*, 64, 493-501.
- Sherwood, M., & Carroll, G. (1974). Fungal succession on needles and young twigs of old-growth Douglas fir. *Mycologia*, 66(3), 499-506.
- Simard, S. W., Beiler, K. J., Bingham, M. A., Deslippe, J. R., Philip, L. J., & Teste, F. P. (2012). Mycorrhizal networks: mechanisms, ecology and modelling. *Fungal Biology Reviews*, 26(1), 39-60.
- Simard, S. W., & Durall, D. M. (2004). Mycorrhizal networks: a review of their extent, function, and importance. *Canadian Journal of Botany*, 82(8), 1140-1165.
- Smith, M. L., Bruhn, J. N., & Anderson, J. B. (1992). The fungus Armillaria bulbosa is among the largest and oldest living organisms. *Nature*, 356(6368), 428-431.
- Smith, S. E., & Read, D. J. (2010). Mycorrhizal symbiosis: Academic press.
- Sohi, H. S. (1988). Mushroom culture in India, recent research findings, Indian Phytopathology, vol. 41, pp. 313–326.
- Sønstebø, J. H., & Rohrlack, T. (2011). Possible implications of chytrid parasitism for population subdivision in freshwater cyanobacteria of the genus Planktothrix. *Applied and environmental microbiology*, 77(4), 1344-1351.
- Spatafora, J. W., Aime, M. C., Grigoriev, I. V., Martin, F., Stajich, J. E., & Blackwell, M. (2017). The fungal tree of life: from molecular systematics to genome-scale phylogenies. *The fungal kingdom*, 1-34.
- Spatafora, J. W., Chang, Y., Benny, G. L., Lazarus, K., Smith, M. E., Berbee, M. L., . . Gryganskyi, A. (2016). A phylum-level phylogenetic classification of zygomycete fungi based on genome-scale data. *Mycologia*, 108(5), 1028-1046.
- Spribille, T., Tagirdzhanova, G., Goyette, S., Tuovinen, V., Case, R., & Zandberg, W. F. (2020). 3D biofilms: in search of the polysaccharides holding together lichen symbioses. *FEMS Microbiology Letters*, 367(5), fnaa023.
- Spribille, T., Tuovinen, V., Resl, P., Vanderpool, D., Wolinski, H., Aime, M. C., ... & McCutcheon, J. P. (2016). Basidiomycete yeasts in the cortex of ascomycete macrolichens. *Science*, 353(6298), 488-492.
- Stamets, P. (1993). Growing Gourmet and Medicinal Mushrooms, Ten Speed Press, Berkeley, Calif, USA.
- Stone, J. K., Polishook, J. D., & White, J. F. (2004). Endophytic fungi. *Biodiversity* of fungi: inventory and monitoring methods, 241, 270.
- Tedersoo, L., Bahram, M., Põlme, S., Kõljalg, U., Yorou, N. S., Wijesundera, R., . . . Suija, A. (2014). Global diversity and geography of soil fungi. *science*, 346(6213), 1256688.

- Tedersoo, L., & Nilsson, R. H. (2016). Molecular identification of fungi. *Molecular mycorrhizal symbiosis*, 299-322.
- Teste, F. P., Simard, S. W., Durall, D. M., Guy, R. D., Jones, M. D., & Schoonmaker, A. L. (2009). Access to mycorrhizal networks and roots of trees: importance for seedling survival and resource transfer. *Ecology*, 90(10), 2808-2822.
- Tuovinen, V., Ekman, S., Thor, G., Vanderpool, D., Spribille, T., & Johannesson, H. (2019). Two basidiomycete fungi in the cortex of wolf lichens. *Current Biology*, 29(3), 476-483.
- Ukai, S., KIHo, T., HARA, C., MORITA, M., GOTO, A., IMAIZUMI, N., & HASEGAWA, Y. (1983). Polysaccharides in fungi. XIII. Antitumor activity of various polysaccharides isolated from Dictyophora indusiata, Ganoderma japonicum, Cordyceps cicadae, Auricularia auricula-judae, and Auricularia species. *Chemical and pharmaceutical bulletin*, 31(2), 741-744.
- Vaidya, J., & Rabba, A. (1993). Fungi in folk medicine. Mycologist, 7(3), 131-133.
- Varma, A., Verma, S., Sudha, Sahay, N., Bütehorn, B., & Franken, P. (1999). Piriformospora indica, a cultivable plant-growth-promoting root endophyte. *Applied and environmental Microbiology*, 65(6), 2741-2744.
- Waller, F., Achatz, B., Baltruschat, H., Fodor, J., Becker, K., Fischer, M., ... & Kogel, K. H. (2005). The endophytic fungus Piriformospora indica reprograms barley to salt-stress tolerance, disease resistance, and higher yield. *Proceedings of the National Academy of Sciences*, 102(38), 13386-13391.
- Wang, H., Ng, T., & Liu, Q. (2004). Alveolarin, a novel antifungal polypeptide from the wild mushroom Polyporus alveolaris. *Peptides*, 25(4), 693-696.

Watkinson, S. C., Boddy, L., & Money, N. (2015). The fungi: Academic Press.

- Wijayawardene, N. N., Hyde, K. D., Lumbsch, H. T., Liu, J. K., Maharachchikumbura, S. S., Ekanayaka, A. H., . . . Phookamsak, R. (2018). Outline of ascomycota: 2017. *Fungal Diversity*, 88, 167-263.
- Wirtz, N., Lumbsch, H. T., Green, T. A., Türk, R., Pintado, A., Sancho, L., & Schroeter, B. (2003). Lichen fungi have low cyanobiont selectivity in maritime Antarctica. *New Phytologist*, 160(1), 177-183.
- Yahr, R., Vilgalys, R., & DePriest, P. T. (2006). Geographic variation in algal partners of Cladonia subtenuis (Cladoniaceae) highlights the dynamic nature of a lichen symbiosis. *New phytologist*, 171(4), 847-860.